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# HELMINTHS OF *CHIASMOCLEIS ALBOPUNCTATA* (BOETTGER, 1885) (ANURA: MICROHYLIDAE) AND *DENDROPSOPHUS NANUS* (BOULENGER, 1889) (ANURA: HYLIDAE) IN CERRADO, SOUTHEASTERN BRAZIL

HELMINTOS DE *CHIASMOCLEIS ALBOPUNCTATA* (BOETTGER, 1885) (ANURA: MICROHYLIDAE) E *DENDROPSOPHUS NANUS* (BOULENGER, 1889) (ANURA: HYLIDAE) EM UMA ÁREA DE CERRADO, CENTRO-OESTE DO BRASIL

## HELMINTOS DE *CHIASMOCLEIS ALBOPUNCTATA* (BOETTGER, 1885) (ANURA: MICROHYLIDAE) Y *DENDROPSOPHUS NANUS* (BOULENGER, 1889) (ANURA: HYLIDAE) EN CERRADO, SURESTE DEL BRASIL

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# ABSTRACT

Brazil is the country with the largest amphibian richness in the world with about 1,136 described species, of which 1,093 are anurans. However, related studies on the knowledge of the associated helminth fauna are scarce, representing only 8% of anurans. This study aimed to identify the parasite communities, and their parasitic parameters in two amphibian species: *Chiasmocleis albopunctata* (Boettger, 1885) and *Dendropsophus nanus* (Boulenger, 1889) found in eastern Mato Grosso do Sul State. The hosts were collected through pitfall traps with drift fences (PTDF) and the Active Search method at the breeding sites, euthanized, and the helminths removed and identified. A total of 172 amphibians were collected: 70 individuals from *C. albopunctata* (Boettger, 1885) and 102 individuals from *D. nanus* (Boulenger, 1889). Of this total, 84 were parasitised by at least one taxon. The total number of helminths found was 675. Here we report ten helminth taxa, including two new records for the *D. nanus* host species, and five records for *C. albopunctata* host species. Thus, this study represents an important contribution to the knowledge of parasite and host diversity, since the amount of existing studies is insufficient and not representative.

Keywords: Anurans - Diversity Conservation - Helminthology - Parasitology - Wild Animals

# RESUMEN

Brasil es el país con la mayor riqueza de anfibios del mundo con alrededor de 1.136 especies descritas, de las cuales 1.093 son anuros. Sin embargo, los estudios relacionados sobre el conocimiento de la fauna de helmintos asociados son escasos, y representan solo el 8% de los anuros. Este estudio tuvo como objetivo identificar las comunidades de parásitos y sus parámetros parasitarios en dos especies de anfibios: *Chiasmocleis albopunctata* (Boettger, 1885) y *Dendropsophus nanus* (Boulenger, 1889) que se encuentran en el este del estado de Mato Grosso do Sul. Los hospedadores fueron recolectados a través de trampas de caídas con cercas de deriva (PTDF) y por el método de búsqueda activa en los sitios de reproducción, sacrificados, y los helmintos fueron removidos e identificados. Se recolectaron un total de 172 anfibios: 70 individuos de *C. albopunctata* (Boettger, 1885) y 102 individuos de *D. nanus* (Boulenger, 1889). De este total, 84 fueron parasitados por al menos un taxón. El número total de helmintos encontrados fue de 675. Aquí informamos diez taxones de helmintos, incluidos dos nuevos registros para la especie hospedadora *D. nanus* y cinco registros para la especie hospedadora *C. albopunctata*. Por lo tanto, este estudio representa una contribución importante al conocimiento de la diversidad de parásitos y huéspedes, ya que la cantidad de estudios es insuficiente y no representativa.

Palabras clave: Anuros - Conservación de la Diversidad - Helmintología - Parasitología - Animales salvajes

# INTRODUCTION

Brazil is the country with the largest amphibian diversity in the world with about 1,136 species described, of which 1,093 are anurans (Martins-Sobrinho et al., 2017; Segalla et al., 2019). However, related studies on the knowledge of the associated helminth fauna are scarce, representing only 8% of anurans (Campião et al., 2014, 2016; Graça et al., 2017). Given that Brazil has a rich helminth fauna, with about 164 taxa (57% of the rates found for South America) (Campião et al., 2014; Graça et al., 2017), faunal surveys are necessary since the vast majority of anurans have not had their helminth fauna described (Graça et al., 2017). Faunal surveys are urgently needed because the number of newly discovered organisms is disappearing faster than new organisms are described (Greene & Losos, 1988; Dobson et al., 2008; Muniz-Pereira et al., 2009). This is especially true in areas such as the Cerrado, which has a very large diversity of species, mainly endemic, but has a high degree of degradation recorded, being constantly impacted by anthropic activities which causes extinction of biodiversity, invasion of exotic species, soil erosion, and

fragmentation of habitats (Klink & Machado, 2005; Myers *et al.*, 2000).

Anurans are animals widely diverse in their ways of life and can spend their lives close to bodies of water, as well as use them only in the larval stages and breeding periods. They can even live in environments with low water availability. Anurans' way of life has a great influence on their parasitic composition, just as parasites are major determinants of this host group. However, amphibian helminths constitute a poorly sampled group in taxonomic survey studies (Anjos, 2011; Campião *et al.*, 2014).

Knowledge of the associated helminth fauna is important in understanding the ecological interactions between the various species of a community, understanding the ecosystem functions (McCallum & Dobson, 1995; Poulin & Morand, 2000, 2004), biogeography (Rohde, 2005), and environmental quality assessment of ecosystems (Thomas *et al.*, 2005). Projects related to environmental conservation and recovery require knowledge of ecology and the systematics of organisms and ecosystems (Scott *et al.*, 1987). Given these assumptions, knowledge of wildlife fauna, associated parasites, and ecological interactions is an important tool for conservation, recovery, and rational use of the environment (McCallum & Dobson, 1995; Poulin & Morand, 2000; Santos, 2003; Thomas *et al.*, 2005, 2009).

This study aimed to reduce the gap in the helminthological knowledge of anurans by identifying parasite communities and their parasitic parameters in two amphibian species *Chiasmocleis albopunctata* (Boettger, 1885) and *Dendropsophus nanus* (Boulenger, 1889) in an area of Cerrado Biome, found in eastern Mato Grosso do Sul state in the Midwest region of Brazil.

# MATERIAL AND METHODS

Study Area

The collections were performed in an area of the Gallery Forest adjacent to the Stream of Véstia, at UNESP Research and Extension Teaching Farm (FEPE), Ilha Solteira Campus (20 ° 21'48" S, 051 ° 24'17" W), Selvíria Municipality, Mato Grosso do Sul State, Brazil.

#### Host Collection

The hosts were collected through six sets of Pitfall Traps with Drift Fence (PTDF) (Corn, 1993), consisting of a Y-shaped fence with a centre bucket and a bucket at each "Y" vertex, usually in places near anuran breeding sites. We also used the searching in breeding sites method described by Scott & Woodward (1994), which consists of systematically walking around and through the given collection site and searching for the animals of interest (Halliday, 2006). In this work, the active search collections were always performed by an average of three people for three consecutive hours (from 20:00 to 23:00 hours). The collections took place monthly between September 2013 and November 2015 (SisBio 36667-2; CEUA no. 06/2014).

At UNESP's Ecology Laboratory of Parasitism (Lecop), the hosts were euthanised with sodium thiopental solution. All internal organs, the cellomatic cavity, and hind limb musculature were evaluated for the presence of helminths.

The hosts were deposited at the Museum of

Zoology "Adão José Cardoso" State University of Campinas (ZUEC).

#### Helminths Collection

Helminths were collected, fixed, and processed following commonly employed methodologies (Amato *et al.*, 1991). The nematodes were clarified with Aman's lactophenol (Andrade, 2000), and species identification was based on articles and classification keys (Vicente *et al.*, 1990; Gibbons, 2010; Anderson *et al.*, 2009). Trematodes were submitted to the hydrochloric carmine staining technique (Amato *et al.*, 1991), diaphanized with clove oil, and identified according to Fernandes & Kohn (2014). Parasitic parameters such as Prevalence (P) and Mean Infection Intensity (IMI) were calculated according to Bush *et al.* (1997), with the aid of the SigmaEstat 3.1 program.

The morphometric, morphological, and photomicrograph data of the helminths were obtained using a computerised LAS V4 image analysis system (Leica Application Suite) adapted to the DM 2500-Leica microscopes with phase interferential contrast system.

The parasites were deposited in the Helminthological Collection of the Department of Parasitology, Institute of Biosciences, Paulista State University (UNESP) (CHIBB 8798 - 8818), Botucatu, State of São Paulo, Brazil.

#### Ethic aspects

All international, national, and institutional guidelines applicable to the care and use of animals were followed.

#### RESULTS

A total of 172 amphibians, 70 *C. albopunctata* (Boettger, 1885) individuals and 102 *D. nanus* (Boulenger, 1889) individuals were collected. Of these, 84 were parasitised by at least one taxon (P = 48.8%). The total number of helminths found was 675, with a mean infection intensity of  $5.8 \pm 1.4$  (1-154).

In the host species *C. albopunctata* (Boettger, 1885), 214 helminths were found, of which 207

were nematodes, and seven were trematodes. The overall prevalence was 62.8%, and the IMI was 2.8  $\pm$  0.3 (1-14). The nematodes were the most prevalent helminths (97.7%) with an IMI of 2.8  $\pm$  0.3 (1-14), while trematodes had a prevalence of 2.3% and a total of seven individuals collected in a single host.

In the host species *D. nanus* (Boulenger, 1889), 461 helminths were found, of which 452 were trematodes, five were acanthocephalans, and four

were nematodes. The total prevalence of helminths was 39.2%, and the IMI was  $11 \pm 3.7$  (1-154). The prevalence of trematode infection in this host species was 90% with an IMI of  $12.2 \pm 4.2$  (1-154); for acanthocephalans, the prevalence was 5% with an IMI of  $2.5 \pm 1.5$  (1-4), and for nematodes the prevalence was 7.5% with an IMI of  $1.3 \pm 0.3$  (1-2). The taxa found in each host species, infection sites, their prevalence (P) and mean infection intensity (IMI), and abundance are described in Table 1.

**Table 1.** Parasitic parameters of nematodes and trematodes associated with anurans *Chiasmocleis albopunctata* (Boettger, 1885) and *Dendropsophus nanus* (Boulenger, 1889). ID = Small Intestine; Cav = Cavity; GI = Large Intestine; P = Overall Prevalence; IMI = Mean Intensity of Infection; SI = Site of Infection; E = stomach.

Ch	iasmocleis albopunc	tata (Boettger, 1885)	
Helminths	P (%)	IMI	SI
Cosmocercidae	60.5	$3.2 \pm 0.46$ (1-12)	ID, IG, Ca
Aplectana sp.	2.3	3	ID
Cosmocerca parva	67.4	$2.5 \pm 0.4$ (1-14)	ID, IG, Cav
Mesocoelium monas	2.3	7	ID
Larva of Nematoda	2.3	1	IG
P (%)	58.6		
IMI	$5.2 \pm 2.7$		
D	endropsophus nanus	r (Boulenger, 1889)	
Helminths	P (%)	IMI	SI
Cosmocercidae	7.5	$1.3 \pm 0.33$ (1-2)	ID, IG
Lophosicyadiplostomum sp.	85	13.1 ± 4.5 (1-154)	Rim
Cyst of Trematoda digenea	2.5	1	Rim
Metacercaria unidentified	2.5	3	Е
Neohaematoloechus neivai	2.5	2	Pulmão
Echinorhynchidae	5	2.5 ± 1.5 (1-4)	E, ID
P(%)	41.2		
IMI	$11.5\pm24.5$		

	Dendropsophus nanus (Boulenger, 1889)	
Family Cosmocercidae	Campião et al. (2014, 2016)	
Lophosicyadiplostomum sp.	Hamann & González (2009)	
Cyst of Trematoda Digenea	Unidentified	
Metacercaria unidentified	Unidentified	
Neohaematoloechus neivai	First report	
Family Echinorhynchidae	First report	
	Chiasmocleis albopunctata (Boettger, 1885)	
Family Cosmocercidae	First report	
<i>Aplectana</i> sp.	First report	
Cosmocerca parva	First report	
Mesocoelium monas	First report	
Larva of Nematoda	First report	

 Table 2. List of taxa found for each host species, highlighting what has been reported in the literature and what is being reported for the first time.

# DISCUSSION

We report here ten helminth rates and seven new records, in addition to the host species *C. albopunctata* (Boettger, 1885), which had not been studied so far (Table 2).

The anuran *D. nanus* (Boulenger, 1889) is found from Suriname and French Guiana, to Uruguay, Argentina, Paraguay and Bolivia, and in Brazil it occurs from the Northeast to the South (Frost, 2019). The registration of two new helminth taxa for this species demonstrates the lack of studies in this area.

Existing parasitological studies revealed 11 parasitic helminth taxa of *D. nanus* (Boulenger, 1889), these being *Centrorhyncus* sp., Unidentified Acanthocephala, *Contracaecum* sp., *Cosmocerca podicipinus* Baker & Vaucher, 1984, Cosmocercoidea gen. sp., *Cylindrotaenia* sp., *Creptotrema* sp., Digenea gen. sp., Plagiorchiata gen. sp, Diplostomidae gen. sp., and *Brevimulticaecum* sp. (Hamann & Kehr, 1998; González & Hamann, 2011; Aguiar *et al.*, 2014; Campião *et al.*, 2014, 2016; Graça *et al.*, 2017). The anuran *D. nanus* (Boulenger, 1889) has an aquatic larval phase, remains in the vegetation near the water as an adult, and uses the aquatic environment in reproduction (Uetanabaro *et al.*, 2008). Proximity to water during the anuran life cycle is highly related to the high prevalence of trematode parasites (McAlpine, 1997; Muzzall *et al.*, 2001; Bolek & Coggins, 2001, 2003; Paredes-Calderón *et al.*, 2006; Todd, 2007; Toledo *et al.*, 2018), which explains the helminth fauna described for *D. nanus* (Boulenger, 1889) in this work. Thus, the use of water bodies during the life cycle of *D. nanus* (Boulenger, 1889) contributed to determining the parasite community of this anuran.

The Cosmocercidae family comprises of several nematode species and have been reported in several South American countries parasitising the intestine, rectum, stomach, and lungs of their hosts. Unidentified individuals of the family have been reported in *Dermatonotus muelleri* (Boettger, 1885), *Physalaemus nattereri* (Steindachner, 1863) (*=Eupemphix nattereri*), *Boana prasina* (Burmeister, 1856) (*=Hypsiboas prasinus*), *Leptodactylus latrans* (Steffen, 1815), *Leptodactylus pentadactylus* (Laurenti, 1768),

Leptodactylus podicipinus (Cope, 1862), Pethecopus azureus (Cope, 1862) (=Phyllomedusa azurea), Rhinella crucifer (Wied-Neuwied, 1821), Rhinella icterica (Spix, 1824) and Rhinella diptycha (Cope, 1862)(=Rhinella schneideri) (Campião *et al.*, 2014). In the present study, for the species D. nanus (Boulenger, 1889), individuals of this family were found in the small and large intestines of the anurans. Acanthocephalans, belonging to the family Echinorhynchidae, have been reported in Paraguay and Brazil parasitising the intestine and body cavity of the hosts, being some species from this family Acanthocephalus acutispinus Machado Filho, 1968, Acanthocephalus caspanensis Fernandez and Ibarra, 1989, Acanthocephalus correalimai Machado Filho, 1970, Acanthocephalus ula Lent and Santos, 1989 and Pseudoacanthocephalus lutzi (Hamann, 1891) (Campião et al., 2014). However, in this study, individuals of this family were found in the small intestine and the stomach of the anurans. Trematoda Lophosicyadiplostomum is an anuran parasite in its larval phase and has been found in Argentina and Brazil parasitising the kidneys of these hosts. Individuals of the genus have been reported in Scinax nasicus (Cope, 1862), Lysapsus limellum Cope, 1862 (=Pseudis limellum), and D. nanus (Boulenger, 1889) (=Hyla nana) (Hamann & González, 2009; Campião et al., 2014). In the present study, this species was found parasitising the kidneys. The trematode Neohaematoloechus neivai (Travassos & Artigas, 1927), have been found in Brazil and Venezuela parasitising the lungs of their hosts (Campião et al., 2014; Fernandes & Kohn, 2014), having already been reported in Leptodactylus latrans (Steffen, 1815), Leptodactylus labyrinthicus (Spix, 1824), Leptodactylus pentadactylus (Laurenti, 1768), Pseudis paradoxa (Linnaeus, 1758), and Lithobates palmipes (Spix, 1824) (=Rana palmipes).

In this study, with the registration of the trematode *Neohaematoloecus neivai* (Travassos & Artigas, 1927) and acanthocephalans of the family Echinorhynchidae, we increased the number of helminth taxa of *D. nanus* (Boulenger, 1889) to 13.

The frog *C. albopunctata* (Boettger, 1885) is found in Bolivia, Argentina, Paraguay, and the Midwest and Southeast Regions of Brazil (Frost, 2019). Although their distribution covers four different countries, the associated helminth fauna of this anuran has never been studied.

The species *C. albopunctata* (Boettger, 1885) is considered terrestrial and uses the aquatic environment only in the larval phase and during breeding periods (Uetanabaro *et al.*, 2008; Giaretta *et al.*, 2008). Contact with soil may favour nematode infection (Barton, 1999; Bolek & Coggins, 2000, 2003; Iannacone, 2003; Luque *et al.*, 2005; Sena *et al.*, 2018; Toledo *et al.*, 2018), which explains this marked prevalence of these helminths for this host species. Despite this prevalence of nematodes, the richness of parasite species for this host was low with only three species identified.

For the species C. albopunctata (Boettger, 1885), nematodes of the family Cosmocercidae were found parasitising the body cavity of the hosts. For the Cosmocerca parva Travassos, 1925 helminth, there are studies indicating its presence in Argentina, Brazil, Colombia, Paraguay, Guyana, and Peru (Campião et al., 2014). These parasites are found mainly in the intestine and rectum of their hosts, and have been reported in several species of various genera such as Ameerega, Colostethus, Dendropsophus, Edalorhina, Elachistocleis, Hamptophryne, Hypsoboas, Hylodes, Leptodactylus, Odontophrynus, Oreobates, Phyllomedusa, Physalaemus, Pristimantis, Rhaeboophion, Scella, Procella, Rhinella, Proceratophrys, Scarthyla, and Scinax (Campião et al., 2014). In this study, we found Cosmocerca parva Travassos, 1925 in the body cavity of the hosts. Individuals of the genus Aplectana have been documented in several South American countries parasitising the stomach, intestine and rectum of their hosts. Unidentified individuals of the genus have been found parasitising Ceratophrys Cranwelli Barrio, 1980, Dendropsophus microps (Peters, 1872), Dermatonotus muelleri (Boettger, 1885), Haddadus binotatus (Spix, 1824), Boana albopunctata (Spix, 1824) (=Hypsiboas albopunctata), Boana pardalis (Spix, 1824) (=Hypsiboas pardalis), Leptodactylus bufonius Boulenger, 1894, Leptodactylus chaquensis Cei, 1950, Leptodactylus elenae Heyer, 1978, Leptodactylus fuscus (Schneider, 1799), Adenomera marmorata Steindachner, 1867

Helminths of *Chiasmocleis* and *Dendropsophus* 

(=Leptodactylus marmoratus), Leptodactylus mystacinus (Burmeister, 1861), Leptodactylus podicipinus (Cope, 1862), Leptodactylus syphax Bokermann, 1969, Physalaemus signifier (Girard, 1853), Rhinella granulosa (Spix, 1824), Rhinella icterica (Spix, 1824), Rhinella marina (Linnaeus, 1758), Scinax acuminatus (Cope, 1862), Thoropa miliaris (Spix, 1824), and Trachycephalus mesophaeus (Hensel, 1867) (Campião et al., 2014). In this study, Aplectana sp. was found in the small intestine of the host. The Mesocoelium monas (Rudolphi, 1819) Trematoda, according to Campião et al. (2014) and Fernandes & Kohn (2014), have been found in Argentina, Brazil, Colombia, Paraguay, Peru and Venezuela parasitising the small intestine of their hosts, and have been reported in Siphonops annulatus (Mikan, 1822), Leptodactylus fuscus (Schneider, 1799), Leptodactylus mystaceus (Burmeister, 1861), Leptodactylus mystacinus (Burmeister, 1861), Leptodactylus latrans (Steffen, 1815), Leptodactylus pentadactylus (Laurenti, 1768), Rhinella arenarum (Hensel, 1867), Rhinella crucifer (Wied-Neuwied, 1821), Rhinella icterica (Spix, 1824), Rhinella marina (Linnaeus, 1758), Rhinella diptycha (Cope, 1862) (=Rhinella schneideri), Incilius nebulifer (Girard, 1854) and Scinax nasicus (Cope, 1862). In the present study it was also found in the small intestine of the host.

All taxa found in *C. albopunctata* (Boettger, 1885) have never been reported for this host species, as this is the first study using this approach. Here we highlight the five taxa record for this host species. Additional studies with populations from other locations may show different patterns from those found here.

This study represents an important contribution to the knowledge of the diversity of parasites and hosts since the number of studies is insufficient and not representative. It also helps to complement information for ecosystem studies with a view to facilitating conservation projects.

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### **BIBLIOGRAPHIC REFERENCES**

- Aguiar, A, Morais, DH, Cicchi, PJP & Silva, RJ. 2014. Evaluation of Helminths Associated with 14 Amphibian Species from a Neotropical Island Near the Southeast Coast of Brazil. Herpetological Review, vol. 45, pp. 227-236.
- Amato, JFR, Boeger, WA & Amato, SB. 1991. Protocolos para laboratório – Coleta e processamento de parasitos de pescado. Imprensa Universitária – UFRRJ, Seropédica, pp.81.
- Anderson, R, Chabaud, A & Willmott, S. 2009. *Keys to the Nematode parasites of vertebrates*. CABI, Wallingford, CABI, pp. 463.
- Andrade, C. 2000. *Meios e soluções comumente empregados em laboratórios*. Editora Universidade Rural, Seropédica, pp. 353.
- Anjos, LA. 2011. Herpetoparasitology in Brazil: what we know about endoparasites, how much we still do not know. Neotropical Helminthology, vol. 5, pp. 107-111.
- Barton, DP. 1999. Ecology of helminth communities in tropical Australian amphibians. International Journal of Parasitology, vol. 29, pp. 921-926.
- Bolek, MG & Coggins, JR. 2000. Seasonal occurrence and community structure of helminth parasites from the Eastern American toad Bufo americanus americanus, from Southeastern Wisconsin, U.S.A. Comparative Parasitology, vol. 67, pp. 202-209.
- Bolek, MG & Coggins, JR. 2001. Seasonal occurrence and community structure of helminth parasites from the green frog, Rana clamitans melanota, from southeastern Wisconsin, U.S.A. Comparative Parasitology, vol. 68, pp. 164-172.
- Bolek, MG & Coggins, JR. 2003. Helminth Community structure of sympatric Eastern

American toad, Bufo americanus, Northern Leopard frog, Rana pipiens, and Bluespotted salamander, Ambystoma laterale, from Southeastern Wisconsin. Journal of Parasitology, vol. 89, pp. 673-680.

- Bush, AO, Lafferty, KD, Lotz, JM & Shostak, AW. 1997. Parasitology meets ecology on its own terms: Margolis et al. Revisited. The Journal of Parasitology, vol. 83, pp. 575-583.
- Campião, KM, Morais, DH, Dias, OT, Aguiar, A, Toledo, G, Tavares, LER & Silva, RJ. 2014. Checklist of Helminth parasites of Amphibians from South America. Zootaxa, vol. 3843, pp. 1-93.
- Campião, KM, Silva, ICO, Dalazen, GT, Paiva, F & Tavares, LE. 2016. Helminth parasites of 11 anuran species from the Pantanal wetland, Brazil. Comparative Parasitology, vol. 83, pp. 92-100.
- Corn, PS. 1993. Straingth-Line drift fences and pitfall traps. In:Heyer, WR, Donnelly, MA, McDiarmid, RW, Hayek, LC & Foster, MS (eds). Measuring and monitoring biological diversity: standard methods for amphibians. Smithsonian Institution Press.
- Dobson, A, Lafferty, K, Kuris, A, Hechinger, R & Jetz, W. 2008. *Homage to Linnaeus: How many parasites? How many hosts?* Proceedings of the National Academy of Sciences, vol. 105, pp. 11482-11489.
- Fernandes, BMM & Kohn, A. 2014. South American trematodes parasites of amphibians and reptiles. FIOCRUZ, Rio de Janeiro, pp. 225.
- Frost, DR. 2019. Amphibian Species of the World: An Online Reference, Version 6.0. American Museum of Natural History, New York, USA, consulted in 16 of July of 2019, <http://research.amnh.org/herpetology/am phibia/index.html>
- Giaretta, AA, Menin, M, Facure, KG, Kokubum, MNC & Filho, JCO. 2008. Species richness, relative abundance, and habitat of reproduction of terrestrial frogs in the Triângulo Mineiro region, Cerrado biome, southeastern Brazil. Iheringia, Série Zoologia, vol. 98, pp. 181-188.
- Gibbons, LM. 2010. Keys to the nematode parasites of vertebrates: Supplementary volume. CABI Publishing, Wallingford, Reino Unido, pp. 416.

- González, CE & Hamann, MI. 2011. Cosmocercid Nematodes of three species of frogs (Anura: Hylidae) from Corrientes, Argentina. Comparative Parasitology, vol. 78, pp. 212-216.
- Graça, RJ, Oda, FH, Lima, FS, Guerra, V, Gambale, PG & Takemoto, RM. 2017. Metazoan endoparasites of 18 anuran species from the mesophytic semideciduous Atlantic Forest in southern Brazil. Journal of Natural History, vol. 51, pp. 705-729.
- Greene, HW & Losos, JB. 1988. Systematics, natural history, and conservation. BioScience, vol 38, pp. 458-462.
- Halliday, T. 2006. *Amphibians. In*:Sutherland, WJ (ed). *Ecological census techniques*. Cambridge University Press, Cambridge, pp. 218-226.
- Hamann, MI, González, C & Kehr, AI. 2006a. Helminth community structure of the oven frog Leptodactylus latinasus (Anura, Leptodactylidae) from Corrientes, Argentina. Acta Parasitologica, vol. 51, pp. 294-299.
- Hamann, MI, Kehr, AI & González, CE. 2006b. Species affinity and infracommunity ordination of helminths of Leptodactylus chaquensis (Anura: Leptodactylidae) in two contrasting environments from northeastern Argentina. Journal of Parasitology, vol. 92, pp. 1171-1179.
- Hamann, MI & González, CE. 2009. Larval digenetic trematodes in tadpoles of six amphibian species from Northeastern Argentina. Journal of Parasitology, vol. 95, pp. 623-628.
- Hamann, MI & Kehr, AI. 1998. Variación espacio temporal en infrapoblaciones de helmintos y su relación con las fluctuaciones poblacionales de Hyla nana (Anura, Hylidae). Cuadernos de Herpetología, vol. 12, pp. 23-33.
- Iannacone, J. 2003. Helmintos parasitos de Atelopus bomolochus Peters 1973 (Anura: Bufonidae) de Piura, Peru. Gayana, vol. 67, pp. 9-15.
- Klink, CA & Machado, RB. 2005. *A conservação do Cerrado Brasileiro*. Megadiversidade, vol. 1, pp.147-155.
- Luque, JL, Martins, NA & Tavares, LER. 2005. Community structure of metazoan parasites of the yellow Cururu toad, Bufo ictericus

(Anura, Bufonidae) from Rio de Janeiro, Brazil. Acta Parasitology 50: 215-220.

- Martins-Sobrinho, PM, Silva, WGO, Santos, EG, Moura, GJB & Oliveira, JB. 2017. Helminths of some tree frogs of the families Hylidae and Phyllomedusidae in an Atlantic rainforest fragment, Brazil. Journal of Natural History, vol. 51, pp. 1639-1648.
- McAlpine, DF. 1997. Helminth communities in bullfrogs (Rana catesbeiana), green frogs (Rana clamitans), and leopard frogs (Rana pipiens) from New Brunswick, Canada. Canadian Journal of Zoology, vol. 75, pp. 1883–1890.
- McCallum, H & Dobson, A. 1995. Detecting disease and parasite threats to endangered species and ecosystems. Trends in Ecology and Evolution, vol. 10, pp. 190-194.
- Muniz-Pereira, LC, Vieira, FM & Luque, JL. 2009. Checklist of helminth parasites of threatened vertebrate species from Brazil. Zootaxa, vol. 2123, pp.1-45.
- Muzzall, PM, Gillilland, MG, Summer, CS & Mehne, CJ. 2001. Helminth communities of green frogs Rana clamitans Latreille, from southwestern Michigan. Journal of Parasitology, vol. 87, pp. 962-968.
- Myers, N, Mittermeier, RA, Mittermeier, CG, da Fonseca, GAB & Kent, J. 2000. *Biodiversity hotspots for conservation priorities*. Nature, vol. 403, pp. 853-858.
- Paredes-Calderón, L, León-Regagnon, V & Gárcia-Prieto, L. 2004. Helminth infracommunities of Rana vaillanti Brocchi (Anura: Ranidae) in Los Tuxtlas, Veracruz, Mexico. Journal of Parasitology, vol. 90, pp. 692-696.
- Poulin, R & Morand, S. 2000. *The diversity of parasites*. Quarterly Review of Biology, vol. 75, pp. 277-293.
- Poulin, R & Morand, S. 2004. Parasite Biodiversity. Smithsonian Books, Washington, pp.216.
- Rohde, K (ed.). 2005. *Parasitologia Marinha*. Publicação CSIRO, Collingwood IC 3066, Austrália e CABI Publishing, Wallingford, Reino Unido, pp. 565.
- Schaefer, EF, Hamann, MI, Kehr, AI, González, CE & Duré, MI. 2006. Trophic, reproductive and parasitological aspects of the ecology of Leptodactylus chaquensis (Anura: Leptodactylidae) in Argentina.

Herpetological Journal, vol. 16, pp. 387-394.

- Santos, AJ. 2003. Estimativas de riqueza em espécies. In: Cullen, LJR, Valladares-Padua, C & Rudran, R (eds.) Métodos de estudos em Biologia da Conservação & Manejo da vida Silvestre. Ed. da UFPR, Fundação o Boticário de Proteção à natureza, Curitiba, pp. 19-41.
- Segalla, MV, Caramaschi, U, Cruz, CAG, Garcia, PCA, Grant, T, Haddad, CFB, Santana, DJ, Toledo, LF & Langone, JA. 2019. *Brazilian amphibians: list of Species*. Herpetologia Brasileira, vol. 8, pp. 65-96.
- Sena, PA, Conceição, BM, Silva, PF, Silva, WGO, Ferreira, WB, Júnior, VAS, Moura, GJB & Oliveira, JB. 2018. Helminth communities of Pithecopus nordestinus (Anura: Phyllomedusidae) in forest remnants, Brazil. Herpetology Notes, vol. 11, pp. 565-572.
- Scott, JM, Csuti, B, Jacobi, JD & Estes, JE. 1987. Species richness: a geographical approach to protecting future biological diversity. BioScience, vol. 37, pp. 782-788.
- Scott, NJ & Woodward, BD. 1994. Inventory and monitoring. In: Heyer, WR, Donnelly, MA, McDiarmid, RW, Hayek, LA & Foster, MS (eds.). Measuring and monitoring biological diversity – Standard methods for amphibians. Smithsonian Institution, Washington.
- Thomas, F, Guégan, JF & Renaud, F. (eds.). 2009. Ecology and evolution of parasitism. Oxford University Press, USA.
- Thomas, F, Renaud, F & Guégan, JF. (eds.). 2005. *Parasitism and ecosystems*. Oxford University Press, USA.
- Todd, BD. 2007. Parasites lost? An overlooked hypothesis for the evolution of alternative reproductive strategies in Amphibians. The American Naturalist, vol. 170, pp. 793-799.
- Toledo, GM, Schwarts, HO, Nomura, HAQ, Aguiar, A, Velota, RAMV, Silva, RJ & Anjos, LA. 2018. Helminth community structure of 13 species of anurans from Atlantic rainforest remnants, Brazil. Journal of Helminthology, vol. 92, pp. 438-444.
- Uetanabaro, M, Prado, CPA, Rodrigues, DJ, Gordo, M & Campos, Z. (eds.). 2008. Guia de Campo dos Anuros do Pantanal e

Planaltos de Entorno. Ed. UFMS, Campo Grande; Ed UFMT, Cuiabá, pp. 196.

Vicente, JJ, Rodrigues, HO, Gomes, DC & Pinto, RM. 1990. Nematoides do Brasil. Parte II: Nematoides de anfibios. Revista Brasileira Zoologia, vol. 7, pp. 549-626. Received September 27, 2019. Accepted November 30, 2019.