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10 ORIGINAL ARTICLE / ARTÍCULO ORIGINAL

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31 Running Head: Apidemiological aspects of parasite infection in cats

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## 45 **ABSTRACT**

46 This study investigated the presence of bronchopulmonary and gastrointestinal parasites  
47 in cats from the state of Sergipe, Brazil, highlighting their medical and veterinary  
48 importance. Fecal samples from 100 felines were analysed using coproparasitological  
49 techniques, showing a 62% positivity rate for parasites, with a significant prevalence of  
50 gastrointestinal species. Notably, bronchopulmonary parasites, particularly  
51 *Aelurostrongylus* sp., were detected for the first time in Sergipe. Samples were analysed  
52 through the Mini-FLOTAC® and Baermann techniques, which proved effective in

53 detecting various parasite types, including eggs, larvae, and cysts. Clinical evaluations  
54 were performed, with clinical signs observed in less than 25% of infected cats. The study  
55 identified eight parasite genera, with *Ancylostoma* sp. and *Strongyloides* sp. among the  
56 most prevalent gastrointestinal parasites, while *Aelurostrongylus* sp. represented  
57 bronchopulmonary infections. Both monoparasitism and polyparasitism were recorded,  
58 with Mini-FLOTAC® showing superior diagnostic performance. This research presents  
59 evidence of *Aelurostrongylus* sp. in domestic cats in Sergipe and underscores the need  
60 for enhanced veterinary surveillance. Findings provide a foundation for understanding  
61 the transmission dynamics and parasite-host relationships of these diseases,  
62 contributing significantly to the epidemiological knowledge in the region.

63 **Keywords:** Bronchopulmonary parasites - clinical alterations - feline parasites - Mini-  
64 FLOTAC®

## 66 **RESUMEN**

67 Este estudio investigó la presencia de parásitos broncopulmonares y gastrointestinales  
68 en gatos del estado de Sergipe, Brasil, destacando su importancia médica y veterinaria.  
69 Se analizaron muestras fecales de 100 felinos mediante técnicas coproparasitológicas,  
70 revelando una tasa de positividad del 62% para parásitos, con una prevalencia  
71 significativa de especies gastrointestinales. De forma relevante, los parásitos  
72 broncopulmonares, particularmente *Aelurostrongylus* sp., fueron detectados por primera  
73 vez en Sergipe. Las muestras se analizaron mediante las técnicas Mini-FLOTAC® y  
74 Baermann, que demostraron ser eficaces en la detección de diversos tipos de parásitos,  
75 incluidos huevos, larvas y quistes. Se realizaron evaluaciones clínicas, observándose  
76 signos clínicos en menos del 25% de los gatos infectados. El estudio identificó ocho  
77 géneros de parásitos, siendo *Ancylostoma* sp. y *Strongyloides* sp. los parásitos  
78 gastrointestinales más prevalentes, mientras que *Aelurostrongylus* sp. representó las

79 infecciones broncopulmonares. Se registraron casos de monoparasitismo y  
80 poliparasitismo, presentando la técnica Mini-FLOTAC® un rendimiento diagnóstico  
81 superior. Esta investigación aporta evidencias de la presencia de *Aelurostrongylus* sp.  
82 en gatos domésticos de Sergipe y refuerza la necesidad de una vigilancia veterinaria  
83 más rigurosa. Los resultados proporcionan una base para la comprensión de la dinámica  
84 de transmisión y de las relaciones parásito-hospedador de estas enfermedades,  
85 contribuyendo de manera significativa al conocimiento epidemiológico de la región.

86 **Palabras clave:** Alteraciones clínicas - parásitos broncopulmonares - parásitos de  
87 felinos - Mini-FLOTAC®

88

## 89 RESUMO

90 Este estudo investigou a presença de parasitas broncopulmonares e gastrointestinais  
91 em gatos do estado de Sergipe, Brasil, destacando a sua importância médica e  
92 veterinária. Amostras fecais de 100 felinos foram analisadas através de técnicas  
93 coproparasitológicas, revelando uma taxa de positividade de 62% para parasitas, com  
94 prevalência significativa de espécies gastrointestinais. De forma relevante, parasitas  
95 broncopulmonares, particularmente *Aelurostrongylus* sp., foram detectados pela  
96 primeira vez em Sergipe. As amostras foram analisadas pelas técnicas Mini-FLOTAC®  
97 e Baermann, que se mostraram eficazes na detecção de diversos tipos de parasitas,  
98 incluindo ovos, larvas e quistos. Foram realizadas avaliações clínicas, tendo sido  
99 observados sinais clínicos em menos de 25% dos gatos infetados. O estudo identificou  
100 oito gêneros de parasitas, sendo *Ancylostoma* sp. e *Strongyloides* sp. os parasitas  
101 gastrointestinais mais prevalentes, enquanto *Aelurostrongylus* sp. representou as  
102 infecções broncopulmonares. Registaram-se casos de monoparasitismo e  
103 poliparasitismo, com a técnica Mini-FLOTAC® a apresentar desempenho diagnóstico  
104 superior. Esta investigação apresenta evidências da presença de *Aelurostrongylus* sp.

105 em gatos domésticos em Sergipe e reforça a necessidade de uma vigilância veterinária  
106 mais rigorosa. Os resultados fornecem uma base para a compreensão da dinâmica de  
107 transmissão e das relações parasita-hospedeiro destas doenças, contribuindo de forma  
108 significativa para o conhecimento epidemiológico na região.

109 **Palavras-chave:** Alterações clínicas - parasitos broncopulmonares - parasitos de felinos  
110 - Mini-FLOTAC®

111

## 112 INTRODUCTION

113 In feline medicine, parasitic diseases are increasingly prevalent and considered  
114 significant causes of clinical manifestations. Cats can be affected by various  
115 gastrointestinal helminths, such as *Ancylostoma* sp., *Toxocara cati* (Schrank, 1788), and  
116 *Dipylidium caninum* (Linnaeus, 1758). However, species of nematodes *Ancylostoma* sp.  
117 and *Toxocara cati* (Schrank, 1788) are more prevalent (Abbas *et al.*, 2022; Filho *et al.*,  
118 2022; Silva *et al.*, 2023). The diagnosis and study of emerging or neglected parasitic  
119 infections have been one of the major challenges in small animal medicine (Abbas *et al.*,  
120 2022).

121 Bronchopulmonary parasites, for example, have sparked great interest among  
122 veterinarians and parasitologists worldwide due to the increase in the number of cases  
123 of felines infected with bronchopulmonary nematodes in several European countries and  
124 the occurrence of the parasitosis in regions that were previously non-endemic (Traversa  
125 & Guglielmini, 2008; Ferraz *et al.*, 2019).

126 The reasons for the emergence of new cases of bronchopulmonary parasitosis  
127 in felines are still unknown; however, some factors such as climate change, intense  
128 movement of companion animals, diversity, and susceptibility of potential intermediate  
129 hosts (vectors) have been taken into consideration (Livia *et al.*, 2023; Yildirim *et al.*,  
130 2023).

131 Pulmonary metastrongylids are among the main agents involved in the infection  
132 of companion animals, where *Aelurostrongylus abstrusus* (Railliet, 1898) (Nematoda,  
133 Metastrongyloidea, Angiostrongylidae), *Troglostrongylus brevior* (Gerichter, 1948), and  
134 *Troglostrongylus subcrenatus* (Railliet, 1913) (Nematoda, Strongylida, Crenosomatidae)  
135 are listed as the main pulmonary parasites of domestic felines (Livia *et al.*, 2023; Yildirim  
136 *et al.*, 2023). The adult forms of these nematodes reside in nodules in the bronchioles  
137 and alveolar ducts (Nonnis *et al.*, 2023; Napoli *et al.*, 2023).

138 As clinical signs in felines infected with bronchopulmonary parasites are not  
139 always detectable, as they may present the disease in its asymptomatic or subclinical  
140 form. Parasitological examinations, such as the Baermann and Mini-FLOTAC® methods,  
141 should be used for confirmation of the diagnosis (Ianniello *et al.*, 2020; Cardoso *et al.*,  
142 2021; Marchiori *et al.*, 2024). However, it's worth noting that young, debilitated, and/or  
143 immunosuppressed animals may present with coughing, sneezing, mucopurulent nasal  
144 discharge, dyspnea, tachypnea, exercise intolerance, anorexia, weight loss, and even  
145 death (Nonnis *et al.*, 2023). When present, clinical signs result from the intense  
146 inflammatory response caused by the elimination of eggs by adult females and the  
147 migration of first-stage larvae (L1) to the bronchial tree, causing lesions in the pulmonary  
148 alveoli, bronchioles, and local arteries (Vismarra *et al.*, 2023; Yildirim *et al.*, 2023).

149 Given the impacts that pulmonary and gastrointestinal parasites can have on  
150 animal health, the zoonotic potential of some of them, their geographic expansion, and  
151 recent advances in epidemiology, clinical, and control of these agents (Abbas *et al.*,  
152 2022), the present study aims to describe the clinical and epidemiological aspects of  
153 bronchopulmonary and gastrointestinal parasite infection in naturally infected felines in  
154 the state of Sergipe, Brazil.

## 155 **MATERIALS AND METHODS**

### 156 ***Study area***

157           The study included felines residing in rural and/or urban areas of the  
158 municipalities of Aracaju, Carira, Nossa Senhora Aparecida, Nossa Senhora das Dores,  
159 Nossa Senhora da Glória, and Itabaiana, located in the state of Sergipe, Brazil  
160 (10°13'06" S/37°25'13" W) (Figure 1). Sergipe is situated in the Northeast Region and is  
161 bordered by the Atlantic Ocean to the east and the states of Bahia to the west and south,  
162 and Alagoas to the north, separated by the São Francisco River (IBGE, 2015).

### 163 ***Experimental design and animals***

164           To achieve the proposed objectives, fecal samples were collected from 100  
165 domestic felines of different breeds, sexes, and ages over 2 months old, residing in  
166 households, shelters, or Non-Governmental Organizations (NGOs). All animals were  
167 clinically evaluated, following three basic steps: 1) Anamnesis and clinical history; 2)  
168 Clinical examination; and 3) Fecal sample collection for diagnostic purposes. It is worth  
169 noting that all clinical alterations of the animals were described in individual records.

### 170 ***Coproparasitological evaluation***

171           Fecal samples were collected from all felines through spontaneous defecation,  
172 with the biological material deposited in properly labeled collection tubes and stored in a  
173 cool box at 4°C until further processing at the Clinical Analysis Laboratory of the  
174 Universidade Federal de Sergipe - Sertão campus, Brazil. All samples were analyzed  
175 using the Mini-FLOTAC® technique (Cringoli *et al.*, 2013) and the Baermann "gold  
176 standard" method (Willesen, 2021).

177           The Mini-FLOTAC® technique was performed by weighing two grams of feces,  
178 which were then deposited into the Fill-FLOTAC® containing 18 mL of zinc sulfate  
179 flotation solution (ZnCl<sub>2</sub> at 1.350; S7). Subsequently, the process of homogenization and  
180 sieving was carried out, and then 5 mL of the solution was transferred to the chamber of  
181 the Mini-FLOTAC® until a positive meniscus formed at the end of each chamber. After  
182 10 minutes, the central disk of the Mini-FLOTAC® was rotated at a 90° angle with the

183 aid of a key located at the end of the plate, which was then removed so that the chamber  
184 could be attached to the optical microscope adapter. Identification and counting of  
185 parasitic structures were then carried out at magnifications of 100X and 400X (Lima *et*  
186 *al.*, 2015).

187 The Baermann technique was performed by weighing approximately 10g of  
188 feces, which were then wrapped in gauze to form a pouch, attached to a rod, and then  
189 deposited over a sedimentation glass containing water, ensuring that the fecal sample  
190 was fully submerged. After an overnight process, 1.5 mL of sediment from the bottom of  
191 the glass was pipetted and deposited into a Falcon tube, which was then centrifuged at  
192 a speed of 1500-2000 rpm for 3 min. Following centrifugation, the supernatant was  
193 discarded, and approximately 0.1 mL of sediment from the tube was pipetted onto a slide  
194 and cover slip (in triplicate), then examined under an optical microscope at  
195 magnifications of 100X and 400X. If necessary, one or two drops of Lugol's solution were  
196 added to fix the larvae (Alho *et al.*, 2013).

197 All parasites observed in the fecal samples were photographed and analyzed  
198 using ImageJ software to obtain morphological and morphometric characteristics, and  
199 were then identified based on the taxonomic keys provided by Bowman *et al.* (2006) and  
200 Taylor *et al.* (2010).

### 201 **Statistical analysis**

202 All data were analyzed using InStat software (GraphPad Software), calculating  
203 absolute frequency (n/N) and relative frequency (%). The Chi-square test ( $X^2$ ) was used  
204 to determine if there was significance between the diagnostic methods and clinical  
205 alterations observed in the animals. The independent variables considered in the model  
206 will be those with statistical significance less than 0.20. EpiInfoTM7 will also be used to  
207 perform statistical calculations, with a significance level established at  $p < 0.05$ .

208 **Ethical aspects**

209 The present research was submitted to and approved by the Ethics Committee  
210 on Animal Use of the Universidade Federal de Sergipe (approval number 4813080621).  
211 Animal participation in the research was voluntary, with felines becoming participants  
212 after the responsible guardian signed the Informed Consent Form, following the  
213 guidelines outlined in Resolution No. 466/2012 of the National Commission for Ethics in  
214 Research of the National Health Council of the Ministry of Health.

215

216 **RESULTS**

217 In this study, parasitological evaluation revealed positivity for gastrointestinal  
218 parasites (93.65%), bronchopulmonary parasites (4.76%), and pseudoparasites (1.59%)  
219 in 62.00% (n=62/100;  $p < 0.0001$ ) of fecal samples from felines in the municipalities of  
220 Aracaju (29.03%; 18/62), Nossa Senhora Aparecida (1.61%; 1/62), Nossa Senhora das  
221 Dores (3.23%; 2/62), and Nossa Senhora da Glória (66.13%; n=41/62) ( $p < 0.0008$ ).

222 All biological samples from cats in the municipalities of Carira (0.00%; 0/5) and  
223 Itabaiana (0.00%; 0/3) were considered negative. The Venn diagram (Figure 2)  
224 schematically demonstrates the presence of bronchopulmonary and gastrointestinal  
225 parasites in 16.67% and 66.67% ( $p < 0.0001$ ) of felines from the Sergipe municipalities,  
226 respectively.

227 Parasitic structures were detected in the forms of eggs (59.30%), oocysts  
228 (18.60%), cysts (4.65%), larvae (16.28%), and proglottids (1.16%) ( $p < 0.0003$ ) (Figure  
229 12). Eight genera of parasites were identified in the fecal samples of felines (Table 1),  
230 with five types of helminths (62.50%) and three protozoa (37.50%) ( $p < 0.0026$ ).  
231 Highlighted among the gastrointestinal parasites were *Ancylostoma* sp. (44.00%; 44/62;  
232  $p < 0.0011$ ), *Dipylidium* sp. (1.00%; 1/62;  $p < 0.0005$ ), *Cystoisospora* sp. (19.00%; 19/62;  
233  $p < 0.0019$ ), *Entamoeba* sp. (3.00%; 3/62;  $p < 0.0001$ ), *Giardia* sp. (3.00%; 3/62;  $p <$

234 0.0001), *Strongyloides* sp. (25.00%; 25/62;  $p < 0.0063$ ), and *Toxocara* sp. (2.00%; 2/62;  
235  $p < 0.0006$ ) as the most frequent, followed by the bronchopulmonary parasite  
236 *Aelurostrongylus* sp. (3.00%; 3/62;  $p < 0.0001$ ).

237 Monoparasitism (33.87%; 21/62) was observed exclusively in felines infected  
238 only with gastrointestinal parasite genera, while polyparasitism (66.13%; 41/62) ( $p <$   
239  $0.0293$ ) was identified in animals with two or more types of enteroparasites, in which  
240 symbiotic interactions: bronchopulmonary parasite + gastrointestinal parasite (4.84%;  $p$   
241  $< 0.0001$ ) and gastrointestinal parasite + gastrointestinal parasite (61.29%;  $p < 0.0233$ )  
242 were reported.

243 Regarding the effectiveness of the parasitological methods employed, 98.41%  
244 and 25.19% of the samples were positive in the Mini-FLOTAC® and Baermann  
245 techniques, respectively ( $p < 0.0001$ ). 100% of eggs, larvae, cysts, oocysts, and  
246 proglottids were diagnosed using the Mini-FLOTAC® technique, while the Baermann  
247 method was able to recover only *Strongyloides* sp. larvae in 12.50% of the analyzed  
248 feline samples ( $p < 0.0001$ ).

249 Regarding the profile of felines positive for enteroparasites, it was observed that  
250 72.00% were females and 28.00% were males, adults, without defined breed, with an  
251 age range between 3 months and 6 years ( $p < 0.0153$ ). Clinically, only 3.23% (2/62) of  
252 the animals had a history of respiratory and/or gastroenteritis symptoms in the last 30  
253 days; however, in the clinical examination, 24.19% (15/62) of the felines presented  
254 noteworthy alterations ( $p < 0.0004$ ) (Figure 3). Clinical alterations were observed in  
255 felines positive for bronchopulmonary and/or gastrointestinal parasites (Figure 4).

## 256 **DISCUSSION**

257 This study reports for the first time the natural infection of domestic felines by  
258 bronchopulmonary parasites in Sergipe, in addition to the description of different  
259 gastrointestinal parasites that have an impact on both animal and human health.

260 The positivity rate (62.00%) for parasitic structures detected in fecal samples of  
261 felines from Sergipe was higher than those reported in studies conducted in Australia  
262 (8.6%) (Mcglade *et al.*, 2003), China (41.39%) (Yang & Liang, 2015), Egypt (52.40%)  
263 (Abbas *et al.*, 2022), United States (5.1%) (Hoggard *et al.*, 2019), France (Bourgoin *et*  
264 *al.*, 2022), Greece (50.70%) (Symeonidou *et al.*, 2018), Italy (22.00%) (Sauda *et al.*,  
265 2019), India (19.00%) (Vincy & Tresamol, 2023), Russia (18.20%) (Kurnosova *et al.*,  
266 2023), and Zimbabwe (1.1%) (Pfukenyi *et al.*, 2010).

267 When comparing the overall positivity rate for bronchopulmonary and/or  
268 gastrointestinal parasites in patients from our study with published data from  
269 epidemiological surveys conducted in Brazil, we observed that studies conducted in  
270 Pernambuco using the FLOTAC® technique showed a higher percentage of positivity  
271 (65.00% to 100%) (Monteiro *et al.*, 2016; Lima *et al.*, 2017). However, research using  
272 traditional techniques (e.g., sucrose flotation, spontaneous sedimentation, and direct  
273 examination) showed lower positivity rates: Rio de Janeiro (49.50%) (Pereira *et al.*,  
274 2017), São Paulo (18.10%) (Gennari *et al.*, 2016), (26.03%), Goiás (17.00% - 60.00%)  
275 (Lima *et al.*, 2018), Rio Grande do Sul (24.63%) (Moreira *et al.*, 2018), and Santa  
276 Catarina (37.80%) (Stalliviere *et al.*, 2009).

277 Clinically, less than 25% of naturally infected felines in Sergipe by bronchopulmonary  
278 or gastrointestinal parasites presented clinical alterations. Some authors emphasize that  
279 certain parasitized animals may be asymptomatic; however, they may develop  
280 cutaneous, gastrointestinal, hepatic, and neurological alterations (Simpson, 1998; Lima  
281 *et al.*, 2021). Arslan *et al.* (2019) also highlight that cats are commonly affected by  
282 toxocariasis, a gastrointestinal parasitic disease caused by larvae of the ascarid  
283 nematodes, *Toxocara canis* (Stiles, 1905) and *T. cati*, which can lead to gastroenteritis,  
284 peritonitis, eosinophilic ascites due to peritoneal inflammation, abdominal pain, and  
285 nausea in infected felines.

286 In feline medicine, respiratory parasitic infections pose a particular challenge for  
287 veterinarians due to the number of pathologies with similar clinical manifestations  
288 (Stepanović *et al.*, 2020). Respiratory alterations were observed exclusively in felines  
289 parasitized by the bronchopulmonary helminth *Aelurostrongylus* sp., which presented  
290 nasal discharge, coughing, sneezing, and dyspnea, clinical signs that, according to  
291 Traversa *et al.* (2010), should be carefully evaluated and considered in the differential  
292 diagnosis of feline cardiopulmonary diseases.

293 Several coproparasitological techniques from the FLOTAC® group are available for  
294 the qualitative and/or quantitative diagnosis of gastrointestinal and pulmonary  
295 parasitoses in felines. However, some studies highlight the importance of using the  
296 Baermann technique in detecting larvae of bronchopulmonary parasites  
297 *Aelurostrongylus abstrusus* and *T. brevior*, which were detected only by the Baermann  
298 test (Colombo *et al.*, 2022). Despite the Mini-FLOTAC® technique showing the best  
299 performance in diagnosing mixed infections (gastrointestinal and pulmonary), the  
300 Baermann test is considered the gold standard for diagnosing cardiopulmonary  
301 nematode parasites in animals, due to the positive hydro/thermotropism demonstrated  
302 by first-stage larvae present in fecal samples (Morelli *et al.*, 2022).

303 The findings of this study highlight the presence of bronchopulmonary parasites in  
304 the state of Sergipe, as well as the presence of gastrointestinal parasite genera with  
305 medical and veterinary significance. Additional studies are warranted to elucidate the  
306 parasite-host relationship and the transmission dynamics of these parasitic diseases in  
307 Sergipe.

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315 **Author contributions: CRediT (Contributor Roles Taxonomy)**

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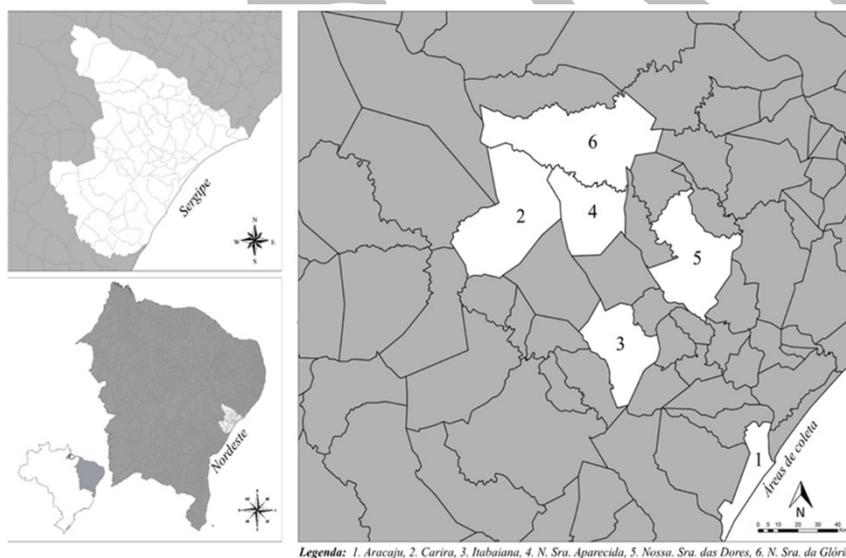
483 **Table 1.** Percentage of positivity by type of parasite detected in populations of cats in  
 484 Northern Brazil.

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486	<b>Parasite/Parasitic structure detected</b>	<b>Absolute Frequency (n/N)</b>	<b>Relative Frequency (%)</b>	<b>Prevalence</b>
487	<i>Aelurostrongylus</i> sp. <sup>L</sup>	3/62	3.00	3
488	<i>Ancylostoma</i> sp. <sup>E</sup>	44/62	44.00	44
	<i>Cystoisospora</i> sp. <sup>O</sup>	19/62	19.00	19
489	<i>Dipylidium</i> sp. <sup>E,P</sup>	1/62	1.00	1
	<i>Entamoeba</i> sp. <sup>C</sup>	3/62	3.00	3
490	<i>Giardia</i> sp. <sup>C</sup>	3/62	3.00	3
	<i>Strongyloides</i> sp. <sup>E</sup>	25/62	25.00	25
491	<i>Toxocara</i> sp. <sup>E</sup>	2/62	2.00	2

492 Note: E – Eggs, C – Cyst, O – Oocyst, L – Larvae. P – Proglottis

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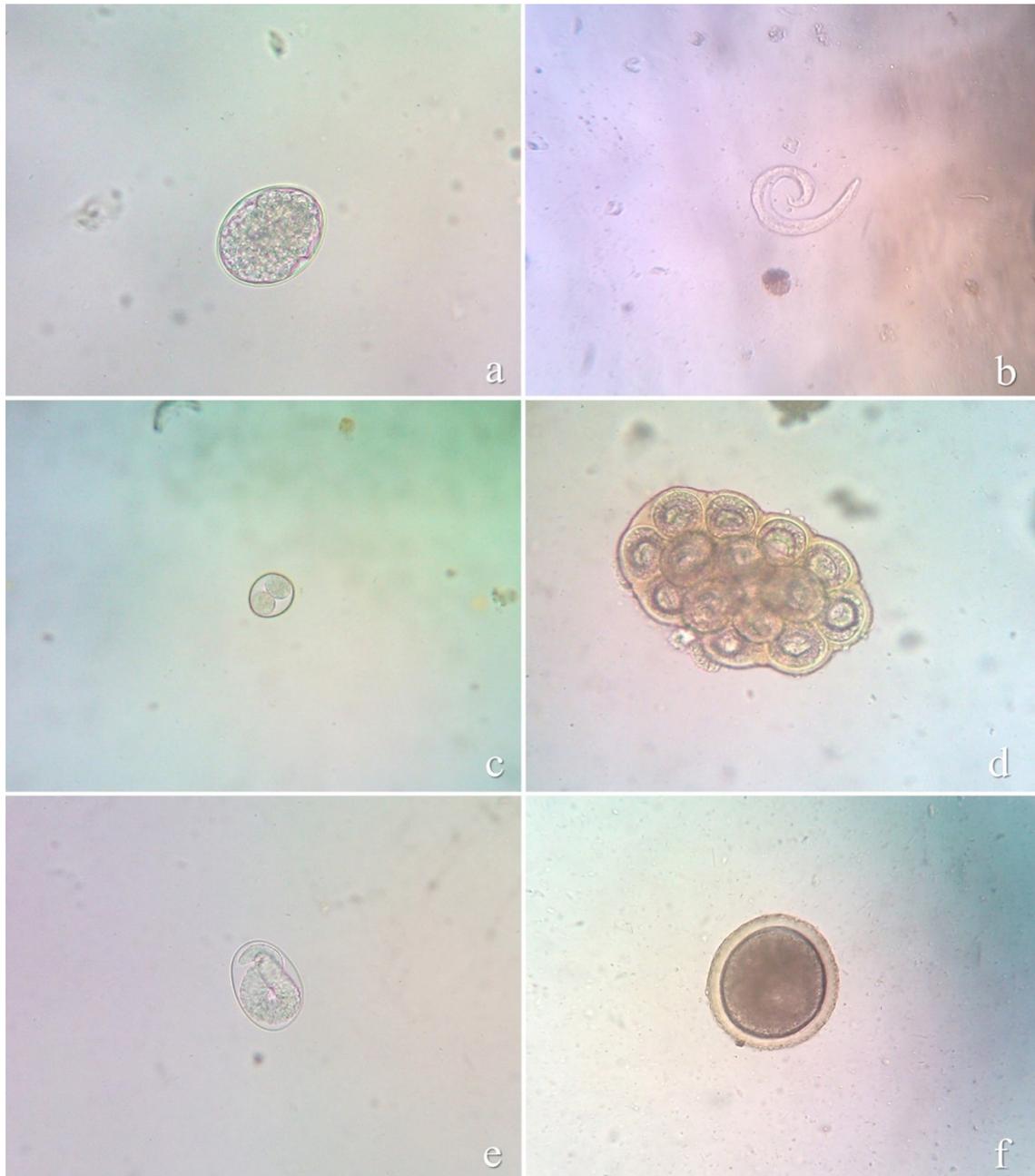
495 **Figure 1.** Topographic location of the collection areas in the state of Sergipe, Northern  
 496 Brazil.

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512 **Figure 3.** Parasitic bronchopulmonary and gastrointestinal parasites diagnosed in cats.

513 a - *Ancylostoma* sp. egg; b - *Aelurostrongylus* sp. larva; c - *Cystoisospora* sp. oocyst; d

514 - *Dipylidium* sp. egg capsule; e - *Strongyloides* sp. larvated egg; f - *Toxocara* sp. egg.

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519 **Figure 4.** Clinical alterations observed in felines positive for bronchopulmonary and/or  
520 gastrointestinal parasites. a) Animal presenting ocular and nasal discharge; b) Feline  
521 presenting cachexia; c) Cat presenting apathy and prostration; and d) Feline presenting  
522 weight loss and skin lesions throughout the body.

523