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ORIGINAL ARTICLE / ARTÍCULO ORIGINAL

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PARASITES OF ELASMOBRANCHS FROM THE SOUTHERN GULF OF MEXICO: THE

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BULL SHARK *CARCHARHINUS LEUCAS* (MÜLLER & HENLE, 1839) AND THE SOUTHERN

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STINGRAY *HYPANUS AMERICANUS* (HILDEBRAND & SCHROEDER, 1928)

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PARÁSITOS DE ELASMOBRANQUIOS DEL SUR DEL GOLFO DE MÉXICO: EL TIBURÓN

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TORO *CARCHARHINUS LEUCAS* (MÜLLER & HENLE, 1839) Y LA RAYA LÁTIGO *HYPANUS*

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AMERICANUS (HILDEBRAND & SCHROEDER, 1928)

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40 Running Head: Elasmobranch Parasites from Southern Gulf of Mexico

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51 ABSTRACT

52 Elasmobranchs play an essential ecological role as top predators in marine ecosystems, and
53 parasitological studies allow the assessment of both host health and the environmental conditions they
54 inhabit. This study analyzed the parasitic diversity of two representative species from the southern
55 coast of the Gulf of Mexico: the bull shark *Carcharhinus leucas* (Müller & Henle, 1839), captured at
56 several fishing localities in the states of Campeche and Tabasco (Seybaplaya, Champotón, Ciudad
57 del Carmen, and Barra de San Pedro), and the southern stingray *Hypanus americanus* (Hildebrand &
58 Schroeder, 1928), collected in Seybaplaya, Campeche. In *C. leucas*, all ten examined specimens
59 showed a 100% prevalence of parasitic infection, with a total of 175 individuals belonging to the
60 groups Monogenea, Cestoda, Nematoda, and Copepoda. The identified taxa were *Nesippus orientalis*
61 (Heller, 1865), *Nemesis* sp. (Risso, 1826), *Paralebion elongatus* (Wilson, 1911), *Erpocotyle*
62 *carcharhini* (Watson & Thorson, 1976), *Granulinema carcharhini* (Moravec & Little, 1988), and
63 *Nybelinia* sp. (Poche, 1926), with cestodes being the most abundant group (64.57%). In 14 examined
64 specimens of *H. americanus*, a total of 1,775 endoparasites were recorded (953 in males and 822 in
65 females), belonging to the groups Cestoda and Nematoda. In this species, the genera
66 *Acanthobothrium* (Van Beneden, 1849), *Nybelinia* (Poche, 1926), *Oncomegas* (Dollfus, 1929),
67 *Pterobothrium* (Diesing, 1850), *Phyllobothrium* (Van Beneden, 1850), *Grillotia* (Guiart, 1927), and
68 *Anisakis* (Dujardin, 1845) were identified. *Oncomegas* showed the highest prevalence (100%) and
69 *Phyllobothrium* the lowest (7.14%), and *Grillotia* is reported for the first time in *H. americanus* from

70 the Gulf of Mexico. Overall, the results reveal a high parasitic diversity dominated by cestodes, which
71 may reflect alterations in coastal habitats. This study provides new parasite records and reinforces the
72 value of parasitology as a tool for understanding biodiversity and assessing the ecological status of
73 elasmobranchs from the southern Gulf of Mexico.

74 **Keywords:** Elasmobranchs – *Carcharhinus leucas* – *Hypanus americanus* – Parasitic diversity –
75 Cestodes – Nematodes – Monogeneans – Copepods – Gulf of Mexico

76 RESUMEN

77 Los elasmobranquios desempeñan un papel ecológico esencial como depredadores tope en los
78 ecosistemas marinos, y los estudios parasitológicos permiten evaluar tanto la salud de los hospederos
79 como las condiciones ambientales que habitan. En este estudio se analizó la diversidad parasitaria de
80 dos especies representativas de la costa sur del Golfo de México: el tiburón toro *Carcharhinus leucas*
81 (Müller & Henle, 1839), capturado en diversas localidades pesqueras de los estados de Campeche y
82 Tabasco (Seybaplaya, Champotón, Ciudad del Carmen y Barra de San Pedro), y la raya látigo
83 *Hypanus americanus* (Hildebrand & Schroeder, 1928), colectada en Seybaplaya, Campeche. En *C.*
84 *leucas*, los diez ejemplares examinados presentaron una prevalencia del 100% de infección
85 parasitaria, con un total de 175 individuos pertenecientes a los grupos Monogenea, Cestoda,
86 Nematoda y Copepoda. Los taxones identificados fueron *Nesippus orientalis* (Heller, 1865), *Nemesis*
87 sp. (Risso, 1826), *Paralebion elongatus* (Wilson, 1911), *Erpocotyle carcharhini* (Watson & Thorson,
88 1976), *Granulinema carcharhini* (Moravec & Little, 1988) y *Nybelinia* sp. (Poche, 1926), siendo los
89 cestodos el grupo más abundante (64.57%). En 14 ejemplares examinados de *H. americanus* se
90 registraron un total de 1,775 endoparásitos (953 en machos y 822 en hembras), pertenecientes a los
91 grupos Cestoda y Nematoda. En esta especie se identificaron los géneros *Acanthobothrium* (Van
92 Beneden, 1849), *Nybelinia* (Poche, 1926), *Oncomegas* (Dollfus, 1929), *Pterobothrium* (Diesing,
93 1850), *Phyllobothrium* (Van Beneden, 1850), *Grillotia* (Guiart, 1927) y *Anisakis* (Dujardin, 1845).
94 *Oncomegas* presentó la mayor prevalencia (100%) y *Phyllobothrium* la menor (7.14%); además,

95 *Grillotia* se reporta por primera vez en *H. americanus* para el Golfo de México. En conjunto, los
96 resultados evidencian una alta diversidad parasitaria dominada por cestodos, lo cual podría reflejar
97 alteraciones en los hábitats costeros. Este estudio aporta nuevos registros de parásitos y refuerza el
98 valor de la parasitología como herramienta para comprender la biodiversidad y evaluar el estado
99 ecológico de los elasmobranquios del sur del Golfo de México.

100 **Palabras clave:** *Carcharhinus leucas* – Céstodos – Copépodos – Diversidad parasitaria –
101 Elasmobranquios – Golfo de México – *Hypanus americanus* – Monogéneos – Nemátodos.

102 INTRODUCTION

103 Elasmobranchs have a wide distribution in tropical and subtropical seas worldwide. In coastal
104 environments they can be found in estuaries, shallow freshwater streams, and coastal lagoon systems
105 (Compagno, 1984). In the Mexican region of the Gulf of Mexico, there is a high diversity of sharks
106 and rays, with more than 80 species recorded, of which approximately 20 have commercial
107 importance. Along the coast of Campeche (Mexico), artisanal fisheries report elasmobranchs among
108 the most abundant resources, particularly shark species such as *Rhizoprionodon terraenovae*
109 (Richardson, 1836) and *Sphyrna tiburo* (Linnaeus 1758), which together represent 88% of shark
110 landings in the state (Martínez-Cruz *et al.*, 2011, 2012).

111 Within this group, the bull shark *Carcharhinus leucas* (Müller & Henle, 1839) and the southern
112 stingray *Hypanus americanus* (Hildebrand & Schroeder, 1928) are species of ecological and
113 economic relevance. The bull shark is a diadromous fish with a broad distribution in tropical and
114 subtropical continental coasts worldwide, including coastal areas, estuaries, and lacustrine systems.
115 It is commercially exploited along the Gulf of Mexico coastline, where it contributes approximately
116 2% of total shark landings (Bravo-Zavala *et al.*, 2022). *H. americanus* is distributed in the western
117 Atlantic Ocean, from New Jersey (USA) to southern Brazil, and is one of the most abundant ray

118 species in the elasmobranch fisheries of the southern Gulf of Mexico. This species primarily inhabits
119 shallow coastal waters, bays, and seagrass meadows (FishBase, 2022). In recent years, its population
120 has shown an estimated decline of 20–29% over the last three generations and is currently considered
121 “Near Threatened” (Carlson *et al.*, 2020).

122 Over recent decades, coastal fisheries of the Gulf of Mexico have been affected by overexploitation,
123 pollution, habitat modification, and other factors that compromise population health (Vidal-Martínez
124 *et al.*, 2015; Morales-Serna *et al.*, 2019). In this context, the study of parasites associated with
125 elasmobranchs represents a useful tool for assessing the ecological status of marine ecosystems, as
126 parasites can act as bioindicators of environmental disturbances (Sures *et al.*, 2017). Sharks and rays,
127 by occupying high trophic levels, provide an exceptional habitat for a wide diversity of parasitic
128 fauna, particularly intestinal helminths such as cestodes (Rodríguez-Santiago *et al.*, 2019, Kleinertz
129 *et al.*, 2022).

130 In the case of *C. leucas*, more than 50 parasite species have been recorded worldwide (Rodríguez-
131 Santiago *et al.*, 2019, Solano-Barquero *et al.*, 2023). However, local studies are scarce, and most have
132 focused on endoparasites, leaving the ectoparasitic fauna poorly documented despite recent reports
133 of parasitic copepods in sharks from the Gulf of Mexico (Rodríguez-Santiago *et al.*, 2015; Rodríguez-
134 Santiago *et al.*, 2016). Similarly, for *H. americanus*, parasitological knowledge remains limited
135 despite its ecological and fishery relevance (Méndez & Galván-Magaña, 2016; Merlo-Serna &
136 García-Prieto, 2016; Pozos-Carré *et al.*, 2020; Rodríguez-Santiago *et al.*, 2019).

137 Given this scenario, the aim of the present study is to report the parasitic fauna recorded in two
138 elasmobranch species commonly captured along the southern coast of the Gulf of Mexico: the bull
139 shark (*C. leucas*), sampled in localities of Campeche and Tabasco (Seybaplaya, Champotón, Ciudad
140 del Carmen, and Barra de San Pedro), and the southern stingray (*H. americanus*), collected in

141 Seybaplaya, Campeche. This study contributes to the knowledge of parasitological biodiversity in the
142 region, as well as to the ecological understanding of the host species and their marine environment.

143 MATERIALS AND METHODS

144 Study area

145 The habitats of the southern Gulf of Mexico encompass a wide variety of coastal ecosystems,
146 including lagoons, estuaries, dunes, mangroves, seagrass beds, and coral reefs, which support more
147 than one thousand fish species. This region is part of the marine ecoregion known as the Southern
148 Gulf of Mexico, a semi-enclosed basin influenced by tropical currents that includes the waters off the
149 states of Veracruz, Tabasco, Campeche, and Yucatán (Wilkinson *et al.*, 2009). The economy of this
150 zone depends largely on three main activities: oil and gas extraction, fisheries, and tourism.

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160 **Figure 1.** Geographic location of the four sampling sites along the southern coast of the Gulf of
161 Mexico: Seybaplaya, Champotón, Ciudad del Carmen, and Barra de San Pedro. The inset shows the
162 position of the study area within the Mexican Republic.

163 For the present study, four representative localities along the southern coast of the Gulf of Mexico
164 were selected: Seybaplaya (19° 38' 02" N, 90° 41' 02" W), Champotón (19° 21' N, 90° 43' W), and
165 Ciudad del Carmen (18° 38' 32" N, 91° 50' 29" W) in the state of Campeche, and Barra de San Pedro
166 (18° 39' 30" N, 92° 28' 12" W) in the state of Tabasco (Fig. 1). Sampling of *C. leucas* was conducted
167 in all four localities, whereas specimens of *H. americanus* were collected exclusively in the fishing
168 zone of Seybaplaya, Campeche.

169 **Host collection**

170 ***Carcharhinus leucas* (bull shark)**

171 Two sampling campaigns were conducted during two collection periods: the first between February
172 and April, and the second between September and November of 2014. A total of 10 specimens were
173 obtained from four localities in the states of Campeche and Tabasco, with the support of local fishing
174 cooperatives and fish vendors from the municipal market of Ciudad del Carmen. Specimens were
175 identified using the field guides of SAGARPA (2008), recording both males and females of different
176 sizes.

177 For each individual, morphometric data—including total length (cm) and body weight (kg)—were
178 recorded. Dissections were performed to remove the gills and spiral valve for parasitological
179 examination. These organs were placed in properly labeled plastic bags and preserved in 70% ethanol
180 for storage and transport to the Environmental Parasitology Laboratory at the Environmental Sciences
181 Research Center of the Universidad Autónoma del Carmen (UNACAR) in Ciudad del Carmen,
182 Campeche, Mexico.

183 ***Hypanus americanus* (southern stingray)**

184 Specimens were obtained from the locality of Seybaplaya, Campeche, during two sampling events
185 conducted in August 2020 and April 2021. Individuals were identified using the key of Mejía-Falla
186 *et al.* (2011). For each specimen, morphometric data such as disk length and width (cm) were recorded
187 (Fig. 2), and dissections were performed to remove the gills and spiral valve for parasitological
188 examination. These organs were placed in properly labeled plastic bags, fixed in 70% ethanol, and
189 kept under cold conditions in a portable cooler for transport to the Environmental Parasitology
190 Laboratory.

191 **Processing of biological samples**

192 Parasitological analyses were conducted following the procedures described by Guzmán-Cornejo *et*
193 *al.* (2012).

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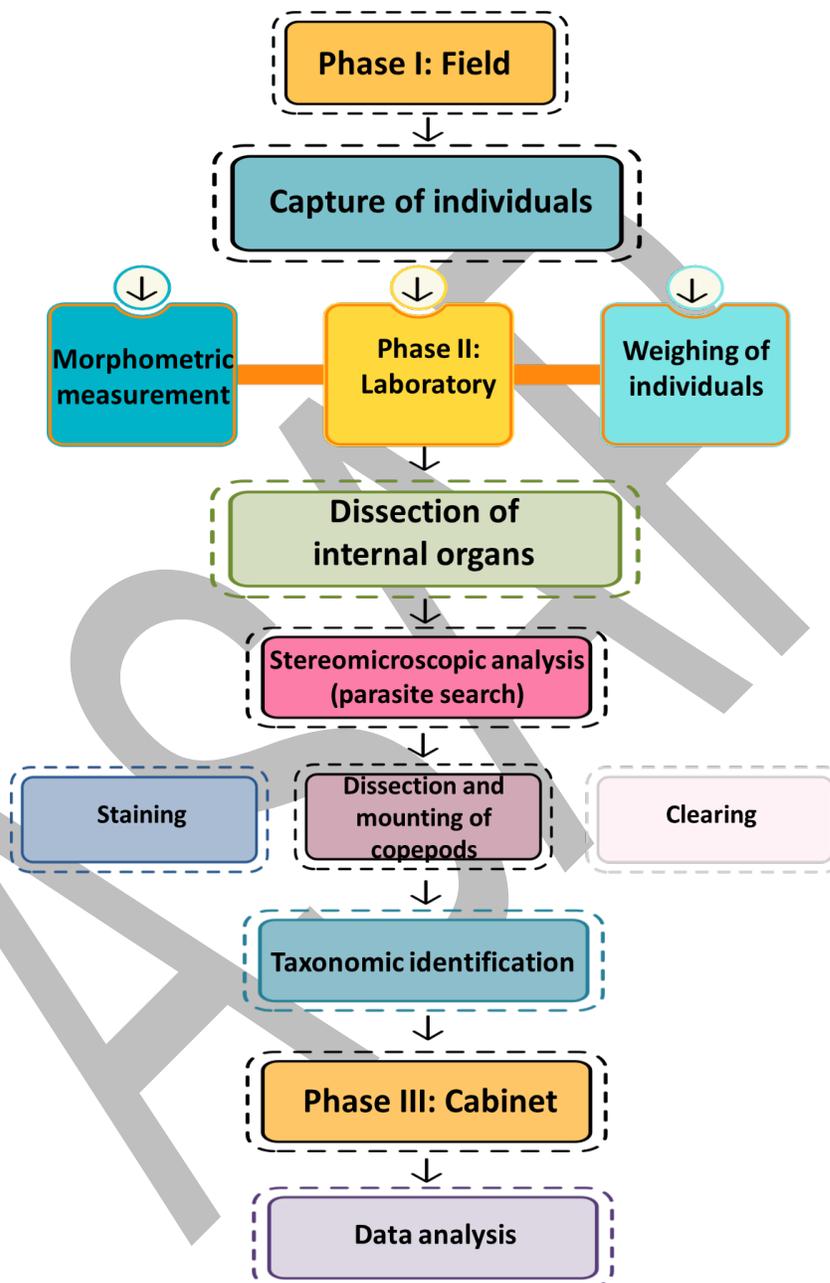
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220 **Figure 2.** Flowchart illustrating the parasitological procedure divided into three phases. Phase I

221 (Field) includes the capture of individuals, morphometric measurements, and weighing. Phase II

222 (Laboratory) involves the dissection of internal organs, stereomicroscopic examination for parasite

223 detection, staining and clearing procedures, and the dissection and mounting of copepods, followed
224 by taxonomic identification. Phase III (Cabinet) corresponds to data analysis and final processing.

225

226 **Parasite identification**

227 Parasite identification was conducted through detailed observation of the characteristic
228 morphological structures of each specimen, with emphasis on diagnostic features. Taxonomic
229 determination was carried out to the levels of family, genus, and species through comparison with
230 previously published morphological descriptions, the use of specialized taxonomic keys, and relevant
231 scientific literature for each metazoan parasite group (Yamaguti, 1959, 1961, 1963, 1971; Anderson
232 *et al.* 1974-1983; Khalil *et al.*, 1994; Walter & Boxshall, 2008, among others). Graphic illustrations
233 of each identified genus were produced using CorelDraw X8, once the helminths had been identified.
234 Two observation techniques were employed: (1) staining using Meyer's paracarmine method
235 (Salgado-Maldonado, 1979) for monogeneans and cestodes, and (2) the technique proposed by
236 Moravec *et al.* (1993) for nematodes and copepods, which involves gradual glycerine solutions to
237 clear the cuticle and allow visualization of internal taxonomic structures of interest (Fig. 2).

238 **Scanning Electron Microscopy (SEM)**

239 For ultrastructural analysis, selected representative specimens of the recovered parasites were
240 processed for scanning electron microscopy. Samples were fixed in 2.5% glutaraldehyde in phosphate
241 buffer, dehydrated through a graded ethanol series, subjected to critical point drying, and
242 subsequently coated with a gold-palladium layer. Observations were carried out using a scanning
243 electron microscope at the facilities of CINVESTAV, Mérida, México.

244 **Data analysis**

245 The characterization of the parasitic communities of *C. leucas* and *H. americanus* was performed
246 following the quantitative ecological descriptors proposed by Bush *et al.* (1997), including
247 prevalence, mean abundance, and mean intensity.

248 **Ethical aspects**

249 All specimens analyzed in this study were obtained from artisanal fisheries and local markets, and no
250 organisms were captured or sacrificed exclusively for research purposes. The collection and handling
251 of hosts complied with national regulations and ethical guidelines for the use of animals in scientific
252 research. Parasitological examinations were conducted following established protocols to minimize
253 unnecessary handling of biological material.

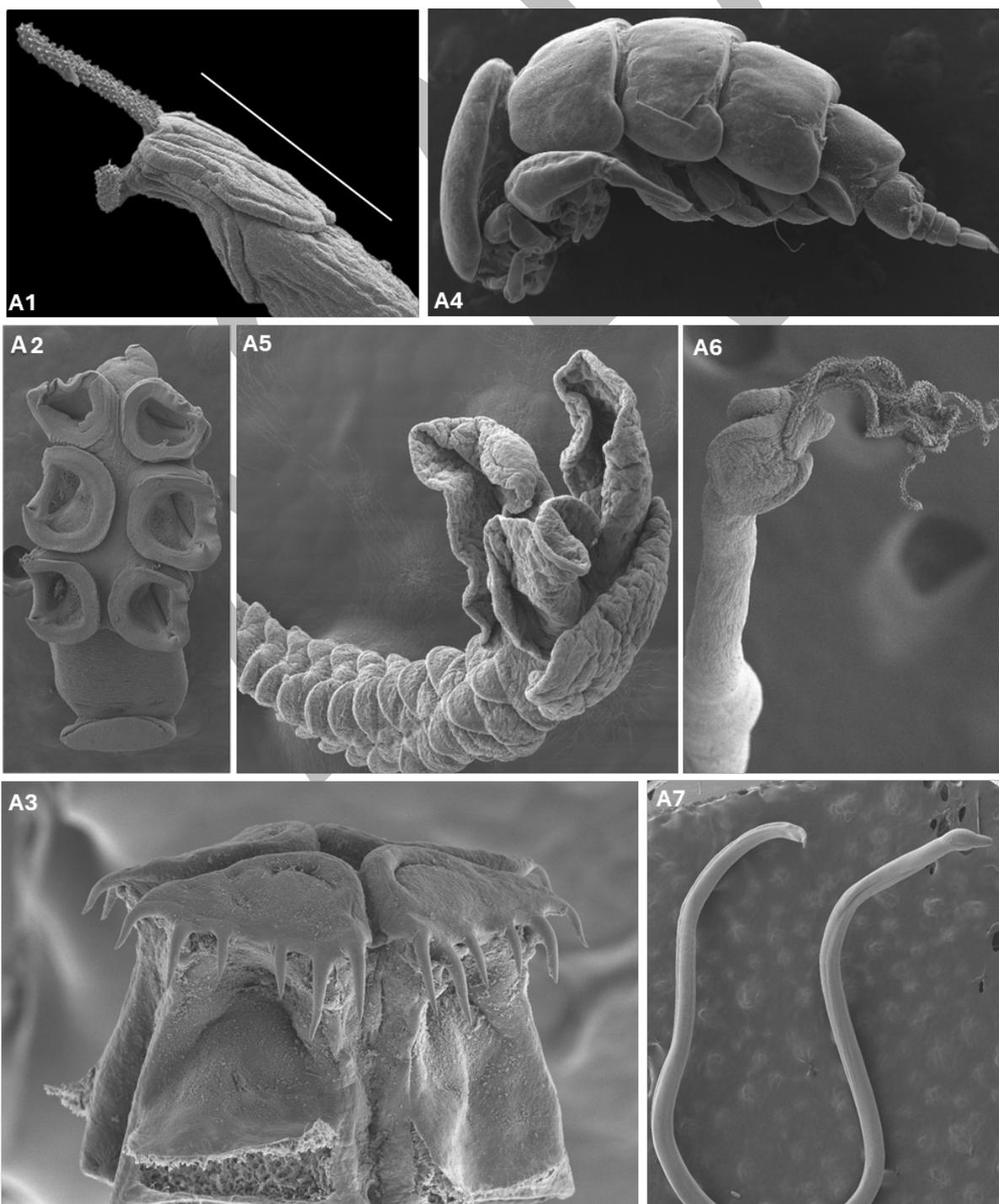
254 **RESULTS**

255 A total of 10 specimens of bull shark (*C. leucas*) were examined: six individuals from Champotón,
256 one from Seybaplaya, one from Ciudad del Carmen, and two from Barra de San Pedro. The mean (\pm
257 standard deviation) total length of these individuals was 132.37 ± 55.67 cm, and mean body weight
258 was 69.6 ± 30.32 kg. A total of 175 parasites were recorded, corresponding to six parasitic species:
259 one cestode (*Nybelinia* sp.; Fig. 2. A1), one monogenean (*Erpocotyle carcharhini* (Watson &
260 Thorson, 1976) (Fig. 2. A2), one nematode (*Granulinema carcharhini* (Moravec & Little, 1988)) Fig.
261 3. B1), and three copepods (*Nemesis* sp., (Fig. 2. A3) *Nesippus orientalis* (Heller, 1865) (Fig. 3. B3),
262 and *Paralebion elongates* (Wilson, 1911)) (Fig. 3, B5). The organs with the highest levels of infection
263 were the stomach and spiral valve, primarily due to cestodes.

264 In the case of the southern stingray (*H. americanus*), a total of 14 specimens from Seybaplaya,
265 Campeche, were examined. Disk length and width (cm) were recorded or estimated for each
266 individual. The mean (\pm standard deviation) disk width was 56.71 ± 8.91 cm, and the disk length was
267 estimated at approximately 42.5 ± 6.7 cm, based on the proportional relationship between disk width

268 and disk length reported for dasyatid rays. A total of 1,775 parasites were recorded, representing
269 seven parasitic species: six cestodes (*Oncomegas* sp., (Fig. 4, C1), *Pterobothrium* sp., (Fig. 4, C2),
270 *Grillotia* sp., (Fig. 4, C3), *Phyllobothrium* sp., (Fig. 4, C1), *Nybelinia* sp., *Acanthobothrium* sp.) and
271 one nematode (*Anisakis* sp.) (Fig. 2, 3 y 4). Cestodes were the predominant helminth group during
272 both sampling periods, occurring in all fourteen spiral valves.

273 Information regarding prevalence, mean intensity, and mean abundance of the parasites found in both
274 elasmobranch species is presented in Table 1.



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291 **Figure 3.** Imágenes obtenidas mediante microscopía electrónica de barrido (MEB). (A1) *Nybelinia*
292 sp. de *C. leucas*, mostrando el escólex en vista lateral con los tentáculos en las regiones basal y
293 metabasal. (A2) *Erpocotyle carcharhini* de *C. leucas*, apreciándose el organismo completo. (A3)
294 *Acanthobothrium* sp. procedente de la válvula espiral de *H. americanus*. (A4) *Nemesis* sp. de *C.*
295 *leucas*. (A5) *Phyllobothrium* sp. obtenido de la válvula espiral de *H. americanus*. (B6) *Nybelinia* sp.
296 de *H. americanus*. (B7) Representación morfológica de *Anisakis* sp. procedente de la válvula espiral
297 de *H. americanus*.

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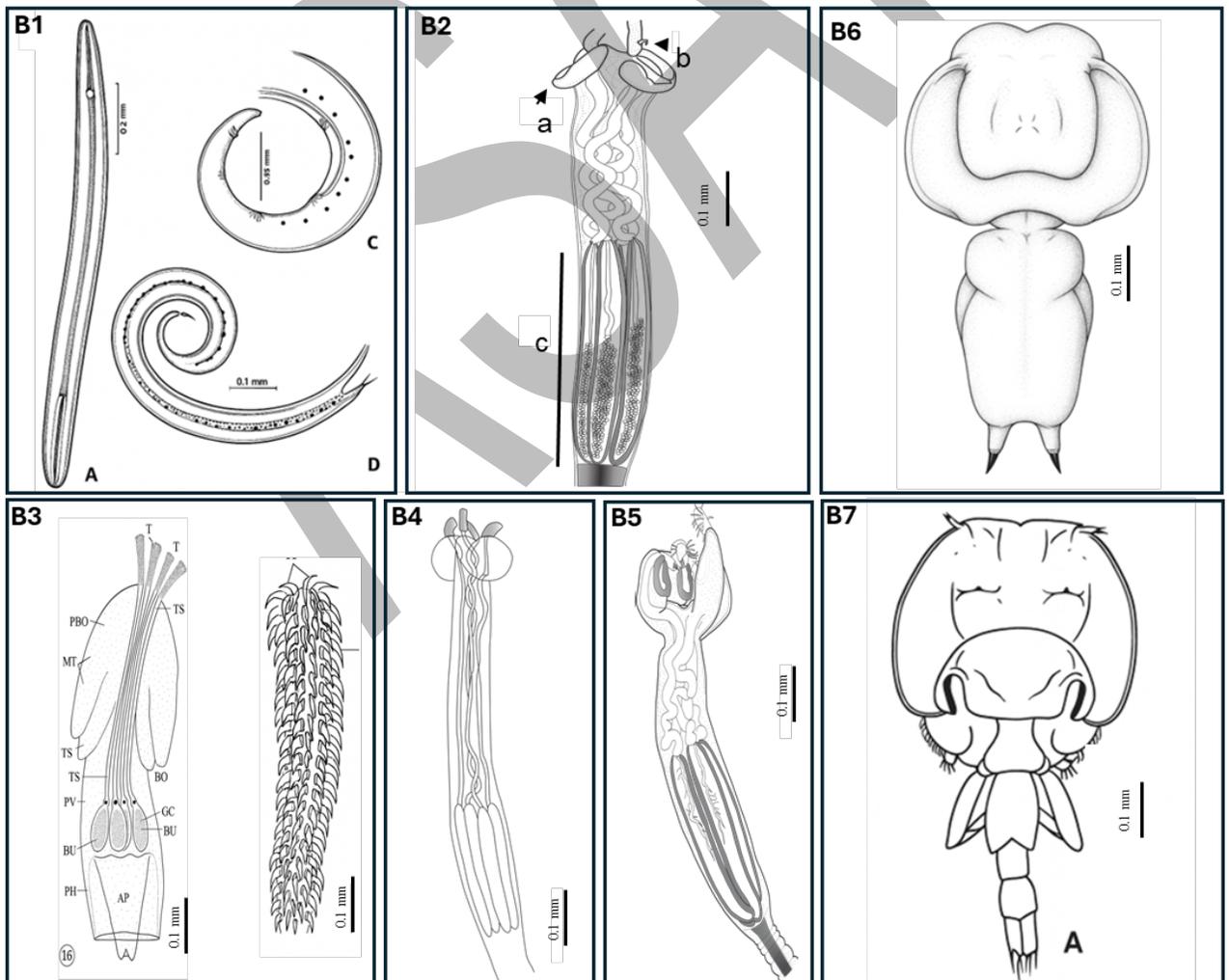
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310 **Figure 4.** (B1) *Granulinema carcharhini* (male): A) esophageal region; B) anterior part of the body;
 311 C) tail; D) posterior part of the body (modified from Moravec & Little, 1988). (B2) Morphological
 312 representation of *Oncomegas* sp. obtained from the spiral valve of *H. americanus*. (B3) *Nybelinia* sp.
 313 from *H. americanus*. (B4) Morphological representation of *Grillotia* sp. from the spiral valve of *H.*
 314 *americanus*. (B5) Morphological representation of *Pterobothrium* sp. obtained from the spiral valve
 315 of *H. americanus*. (B6) Dorsal view of *Nesippus orientalis* observed under the microscope. (B7)
 316 *Paralebion elongatus*: male specimen in dorsal view. Morphological drawings were digitally
 317 prepared using CorelDRAW 2024 based on microscopic observations.

318 **Table 1.** Infection parameters for each parasite species recorded in *Carcharhinus leucas* and *Hypanus*
 319 *americanus*.

Parasite group	Parasite species	Total number of parasites	Prevalence (%)	Mean abundance \pm Standard deviation	Mean intensity \pm Standard deviation
Bull shark (<i>Carcharhinus leucas</i>)					
Monogenea	<i>Erpocotyle carcharhini</i>	25	30	2.5 \pm 1.5	11.3 \pm 5.4
Cestoda	<i>Nybelinia</i> sp.	113	100	11.3 \pm 3.36	2 \pm 0
Nematoda	<i>Granulinema carcharhini</i>	2	10	0.2 \pm 0.45	8.3 \pm 4.9
Copepoda	<i>Nemesis</i> sp.	6	10	0.6 \pm 0.77	6 \pm 0
	<i>Nesippus orientalis</i>	12	30	1.2 \pm 1	4 \pm 2.1
	<i>Paralebion elongatus</i>	17	30	1.7 \pm 1.3	5.6 \pm 0.9
Southern stingray (<i>Hypanus americanus</i>)					
Cestoda	<i>Oncomegas</i> sp.	956	100	68.28 \pm 66.98	68.28 \pm 66.98
	<i>Pterobothrium</i> sp.	620	92.85	44.28 \pm 41.17	47.69 \pm 40.75
	<i>Phyllobothrium</i> sp.	2	7.14	0.14 \pm 0.53	2 \pm 0
	<i>Nybelinia</i> sp.	2	14.28	0.14 \pm 0.36	1 \pm 0
	<i>Acanthobothrium</i> sp.	47	35.71	3.35 \pm 6.87	9.4 \pm 9.09
Nematoda	<i>Grillotia</i> sp.	144	71.42	10.28 \pm 9.44	14.4 \pm 7.93
	<i>Anisakis</i> sp.	4	21.42	0.28 \pm 0.61	1.33 \pm 0.57

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324 **DISCUSSION**

325 This study represents a significant contribution to the parasitological knowledge of elasmobranchs
326 along the southern coast of the Gulf of Mexico, constituting the first record of parasites in *C. leucas*
327 from the localities of Seybaplaya, Champotón, and Ciudad del Carmen in Campeche, and Barra de
328 San Pedro in Tabasco, as well as a new parasitological record for *H. americanus* captured in
329 Seybaplaya, Campeche. These results expand the understanding of the geographic distribution of
330 parasitic species in the Gulf of Mexico and suggest the presence of environmental and ecological
331 conditions that favor their establishment in this region, without implying that they are exclusive to
332 these areas.

333 In *C. leucas*, the ecto- and endoparasite species recorded had not been previously reported for this
334 shark along the coasts of Mexico, allowing it to be considered a new host for several taxa. When
335 compared with studies from other regions of the Gulf, such as that of Méndez & Dorantes-González
336 (2013) in Veracruz, where eight cestode species were reported, only one cestode species (*Nybelinia*
337 sp.) was identified in the present study. This may be attributed to differences in feeding habits,
338 availability of intermediate hosts, or local environmental variations. *C. leucas* is an opportunistic
339 predator (Compagno, 1984) whose diet includes bony fishes potential intermediate hosts of cestodes
340 which explains the presence of helminths such as *Granulinema carcharhini*, a species previously
341 reported in the Gulf of Mexico but with limited taxonomic information (Moravec & Little, 1988).
342 Likewise, the absence of *Erpocotyle carcharhini* in *C. leucas* captured in low-salinity environments
343 such as Laguna de Términos and the San Pedro River aligns with Watson & Thorson (1976), who
344 suggested low tolerance of this parasite to freshwater conditions.

345 Regarding copepods, genera such as *Nemesis* and *Nesippus* are common gill parasites of sharks, and
346 their presence in this region represents the first record for Mexico, although they have been previously
347 documented in the western Atlantic and African waters (Cressey, 1970; Oldewage, 1993). These
348 observations reinforce the hypothesis that *C. leucas*, by moving between marine and estuarine
349 environments, may act as a vector of interoceanic parasite dispersal.

350 In *H. americanus*, seven helminth genera were identified, with a prevalence of 100% for the Class
351 Cestoda and 21.42% for the Phylum Nematoda. The high frequency of cestodes is consistent with
352 reports from other elasmobranchs in the Gulf of Mexico (Méndez & Dorantes-González, 2017;
353 Dorantes-González & Méndez, 2018; Pozos-Carré *et al.*, 2020), where this group constitutes the
354 dominant fraction of the parasitic fauna. This high prevalence is associated with the benthic diet of
355 the ray, which includes mollusks, crustaceans, and small fishes (FishBase, 2022), making it a suitable
356 host for larval stages of numerous cestodes (Palm, 2011).

357 The genera *Oncomegas* and *Pterobothrium* were the most abundant, with prevalences of 100% and
358 92.85%, respectively—results comparable to those observed in Veracruz (Pozos-Carré *et al.*, 2020).
359 *Oncomegas wagneri*, in particular, has been associated with areas containing aromatic hydrocarbons
360 (Vidal-Martínez *et al.*, 2015), which may indicate exposure to contaminants in Seybaplaya. Other
361 genera, such as *Phyllobothrium* and *Acanthobothrium*, exhibited lower prevalences, although their
362 presence reaffirms their role as typical parasites of Gulf elasmobranchs (Méndez *et al.*, 2018). Also
363 noteworthy is the first record of *Grillotia* in *H. americanus* for the Gulf of Mexico, expanding the
364 known distribution of this genus, previously reported in other rays from the Atlantic and southern
365 Africa (Álvarez *et al.*, 2006; Oosthuizen *et al.*, 2021).

366 The low representation of nematodes in both species agrees with observations from other studies and
367 may be attributed to the high concentrations of urea present in elasmobranchs, which hinder the
368 development of these helminths (Modzelesky, 2018; Solano-Barquero *et al.*, 2025).

369 Overall, the results obtained for both species represent only a portion of the parasitic diversity known
370 in chondrichthyans from the Gulf of Mexico, where at least 47 cestode species have been recorded in
371 27 of the 128 described species (Chandler, 1954; McEachran, 2009; Adán-Torres *et al.*, 2025).
372 Nonetheless, the findings of this study suggest the existence of ecological barriers or limiting factors
373 that shape the distribution of certain parasites within the Gulf, potentially influenced by salinity,
374 habitat type, feeding habits, and anthropogenic disturbance. Therefore, expanding sampling areas and
375 increasing sample sizes in future studies will provide a better understanding of the biogeographic
376 patterns and parasite-host relationships in elasmobranchs from the southern (Gulf of Mexico)
377 (Solano-Barquero *et al.*, 2025).

378 This study provides a valuable contribution to the parasitological knowledge of elasmobranchs along
379 the southern coast of the Gulf of Mexico, documenting for the first time the parasitic composition of
380 the bull shark in Campeche and Tabasco, and of the southern stingray in the fishing port of
381 Seybaplaya, Campeche. Six parasite species were recorded in the bull shark, two of which were
382 endoparasites (*Nybelinia* sp. and *G. carcharhini*) and four ectoparasites (*E. carcharhini*, *Nemesis* sp.,
383 *N. orientalis*, and *P. elongatus*). In the southern stingray, seven parasite species were recorded six
384 belonging to the Class Cestoda (*Oncomegas*, *Pterobothrium*, *Phyllobothrium*, *Nybelinia*, *Grillotia*,
385 and *Acanthobothrium*) and one to the Phylum Nematoda (*Anisakis*).

386 The record of *Grillotia* sp. represents the first report of this genus in the southern stingray and in the
387 Gulf of Mexico, extending its known distribution. Overall, the results of this study reveal a high
388 parasitic diversity in the elasmobranchs of the region and highlight the ecological importance of these
389 species as definitive hosts within the trophic networks of the Gulf of Mexico. This work provides a
390 foundation for future research on parasite biogeography, environmental factors influencing helminth
391 distribution, and the potential role of coastal conditions such as salinity and anthropogenic activity in
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435 **BIBLIOGRAPHIC REFERENCES**

436 Adán-Torres, B., García-Prieto, L., & León-Règagnon, V. (2025). Molecular and morphological
437 characterization, phylogenetic affinities and new records of Trypanorhyncha (Cestoda) from
438 the Mexican Atlantic. *Parasitology International*, 109, 103095.

439 Álvarez, M.F., Aragort, W., Leiro, J.M. & Sanmartín, M.L. (2006). Macroparasites of five species of
440 ray (genus *Raja*) on the northwest coast of Spain. *Diseases of Aquatic Organisms*, 70, 93–100.

441 Anderson, R.C., Chabaud A.G., & Willmott S. (1974). *CIH keys to the nematode parasites of*
442 *vertebrates*. Commonwealth Agricultural Bureaux, Farnham Royal, Buck. Vol. 1-10.

443 Anderson, R.C. (1984). The origins of zooparasitic nematodes. *Canadian Journal of Zoology*, 62,
444 317-328.

445 Anderson, R.C. (1988). Nematode transmission patterns. *Journal of Parasitology*, 74, 30-45.

446 Bravo-Zavala, F.G., Pérez-Jiménez, J.C., Tovar-Ávila, J., & Arce-Ibarra, A.M. (2022). Vulnerability
447 of 14 elasmobranchs to various fisheries in the southern Gulf of Mexico. *Marine and*
448 *Freshwater Research*, 73, 1064-1082.

- 449 Bush, A.O., Lafferty, K.D., Lotz, J.M., & Shostak, A.W. (1997). Parasitology meets ecology on its
450 own terms: Margolis *et al.* revisited. *Journal of Parasitology*, 83, 575–583.
- 451 Carlson, J., Charvet, P., Blanco-Parra, M.P., Briones Bell-Iloch, A., Cardenosa, D., Derrick, D.,
452 Espinoza, E., Morales-Saldaña, J.M., Naranjo-Elizondo, B., Pacoureaux, N., Schneider, E.V.C.,
453 Simpson, N.J., Pollom, R., & Dulvy, N.K. (2020). *Hypanus americanus*. *The IUCN Red List*
454 *of Threatened Species* 2020: e.T181244884A104123787.
- 455 Compagno, L.J.V. (1984). *FAO Species Catalogue. Vol. 4. Sharks of the world. An annotated and*
456 *illustrated catalogue of shark species known to date. Part 2 – Carcharhiniformes*. Rome: FAO,
457 pp. 478–486.
- 458 Chandler, A.C. (1954). Cestoda. In: Fish and Wildlife Service, U.S. Department of the Interior (Ed.),
459 *Gulf of Mexico, its origin, waters, and marine life. Fishery Bulletin of the Fish and Wildlife*
460 *Service*, 55, 351–353.
- 461 Cressey, R.F. (1970). Copepods parasitic on sharks from the west coast of Florida. *Smithsonian*
462 *Contributions to Zoology*, 381, 1–30.
- 463 Dorantes-González, M.A., & Méndez, O. (2018). Helmintos parásitos intestinales del tiburón puntas
464 negras *Carcharhinus limbatus* (Müller & Henle, 1839) (Carcharhiniformes: Carcharhinidae),
465 en playa Chachalacas, Veracruz, México. *Neotropical Helminthology*, 12, 213–222.
- 466 FishBase. (2022). *Hypanus americanus* (Hildebrand & Schroeder, 1928) *Mantarraya del sur*. En:
467 *Hypanus americanus, raya del sur: pesquerías, peces de caza*.
468 <https://www.fishbase.se/summary/1247>
- 469 Guzmán-Cornejo, C., García, L., Rivas, G., Mendoza-Garfias, B., Osorio, D. & Montiel, G. (2012).
470 *Manual de prácticas de metazoarios parásitos de vertebrados*. Prensas de Ciencias,
471 Universidad Nacional Autónoma de México, 141 pp.
- 472 Khalil, L.F., Jones, A., & Bray, R.A. (Eds.). (1994). *Keys to the cestode parasites of vertebrates*.
473 CAB International. 751 pp.
- 474 Kleinertz, S., Yulianto, I., Kurschat, C., Koepper, S., Simeon, B. M., Klimpel, S., ... & Palm, H. W.
475 (2022). Elasmobranchs from Indonesian waters: feeding ecology and trypanorhynch cestode
476 fauna composition to support efforts in shark and ray conservation. *Acta parasitologica*, 67,
477 1612–1625.
- 478 Martínez-Cruz, L.E., Balan-Ché, L.I., Seca-Escalante, J.M., Oviedo-Pérez, J.L. & González-
479 Ocaranza, L. (2011). *Caracterización de la pesquería de elasmobranquios en el estado de*
480 *Campeche*. Centro Regional de Investigación Pesquera de Lerma, Campeche, 27 pp.

- 481 Martínez-Cruz, L.E., Oviedo-Pérez, J.L., González-Ocaranza, L., Balan-Ché, L.I. & Maldonado-
482 Martín, J.I. (2012). *Caracterización de la pesquería de elasmobranquios en el estado de*
483 *Campeche*. Centro Regional de Investigación Pesquera de Lerma, Campeche, 59 pp.
- 484 McEachran, J.D. (2009). Fishes (Vertebrata: Pisces) of the Gulf of Mexico. In: Felder, D.L. & Camp,
485 D.K. (Eds). *Gulf of Mexico origin, waters, and biota. Vol. 1. Biodiversity*. Corpus Christi, USA:
486 Texas A&M University Press, 1393 pp.
- 487 Mejía-Falla, P., Navia, A. & Puentes, V. (2011). *Guía para la identificación de especies de tiburones,*
488 *rayas y quimeras de Colombia*. Colombia: Ministerio de Ambiente y Desarrollo Sostenible.
- 489 Méndez, O. & Dorantes-González, M.A. (2013). Cestodes of the bull shark *Carcharhinus leucas* in
490 Chachalacas beach, Veracruz, Mexico. *Neotropical Helminthology*, 7, 167–171.
- 491 Méndez, O. & Dorantes-González, M. (2017). Helmintos parásitos intestinales de tiburones en la
492 costa central del estado de Veracruz, México. *Ciencia Pesquera*, 25, 51–61.
- 493 Méndez, O. & Galván-Magaña, F. (2016). Cestodes of the blue shark, *Prionace glauca* (Linnaeus,
494 1758) (Carcharhiniformes: Carcharhinidae), off the west coast of Baja California Sur. *Zootaxa*,
495 4085, 438–444.
- 496 Méndez, O., Valero-Pacheco, E. & Dorantes-González, M.A. (2018). Helmintos parásitos intestinales
497 de algunos tiburones (Pisces: Elasmobranchii) del Golfo de México. *Neotropical*
498 *Helminthology*, 12, 223–231.
- 499 Merlo-Serna, A. & García-Prieto, L. (2016). A checklist of helminth parasites of Elasmobranchii in
500 Mexico. *ZooKeys*, 563, 73–128.
- 501 Modzelesky, A. S. (2018). *Meta-Analytic Summary of Parasites and Diseases of Elasmobranchs*
502 *Found in Florida Waters* (Nova Southeastern University). NSUWorks. 94p.
- 503 Moravec, F., Kohn, A., & Fernandes, B.M.M. (1993). Nematode parasites of fishes of the Paraná
504 River, Brazil. Part 2. Seuratoidea, Ascaridoidea, Habronematoidea and Acuarioidea. *Folia*
505 *Parasitologica*, 40, 115–134.
- 506 Moravec, F. & Little, M.D. (1988). *Granulinema* gen. n., a new dracunculoid genus with two new
507 species (*G. carcharhini* sp. n. and *G. simile* sp. n.) from the bull shark, *Carcharhinus leucas*
508 (Valenciennes), from Louisiana, USA. *Folia Parasitologica*, 35, 113–120.
- 509 Oldewage, H.W. (1993). Three species of piscine parasitic copepods from southern African coastal
510 waters. *South African Journal of Zoology*, 28, 113–121.
- 511 Oosthuizen, G., Acosta, A.A., Smit, N.J., & Schaeffner, B.C. (2021). A new species of *Grillotia*
512 *Guiart, 1927* (Cestoda: Trypanorhyncha) from the spotted skate, *Raja straeleni* Poll, in South
513 Africa. *Parasitology International*, 82, 102307.

- 514 Palm, H.W. (2011). Fish parasites as biological indicators in a changing world: Can we monitor
515 environmental impact and climate change? In: Mehlhorn, H. (Ed.). *Progress in Parasitology*.
516 *Parasitology Research Monographs*. Springer Verlag.
- 517 Pozos-Carré, D., Uscanga-Alvarado, D., Mendoza-Chacón, C. & Méndez, O. (2020). Helminths
518 parásitos intestinales de la raya *Hypanus americanus* (Hildebrand & Schroeder, 1928) en
519 Chachalacas, Veracruz, México. *Neotropical Helminthology*, 14, 67–73.
- 520 Rodríguez-Santiago, M., Morales-Serna, F., Gómez, S. & Grano-Maldonado, M. I. (2016). New
521 records of parasitic copepods (Copepoda: Pandaridae, Eudactylinidae, Caligidae) on
522 elasmobranchs (Chondrichthyes) in the Gulf of Mexico. *Ciencia Pesquera*, (Special Issue), 15–
523 21.
- 524 Rodríguez-Santiago M.A., Gómez, S., & Grano-Maldonado, M.I. (2015). New records of parasitic
525 copepods (Copepoda: Pandaridae, Eudactylinidae, caligidae) on five shark species (Pisces:
526 Elasmobranchia) in the Gulf of Mexico. *Neotropical Helminthology*, 9, 177–182.
- 527 Rodríguez-Santiago, M.A., Méndez, O., Mandujano-Solís, R.E., & Caballero-Vázquez, A. (2019).
528 Parásitos de elasmobranquios (tiburones y rayas) del sur del Golfo de México: un mundo
529 microscópico desconocido. *Sociedad Ictiológica Mexicana*, 45 p.
- 530 SAGARPA. (2008). *Guía para la identificación de las especies de tiburones de importancia*
531 *comercial en el Golfo de México, términos técnicos y principales medidas*. 3ª edición.
532 CONAPESCA.
- 533 Salgado-Maldonado, G. (1979). *Procedimientos y técnicas generales empleados en los estudios*
534 *helmintológicos*. Departamento de Pesca, Dirección General de Acuicultura, México, 55 pp.
- 535 Solano-Barquero, A., Rojas, A., & Cortés, J. (2023). Metazoan marine parasites of Costa Rica: A
536 review. *Parasitologia*, 3, 116-141.
- 537 Sures, B., Nachev, M., Selbach, C., & Marcogliese, D. J. (2017). Parasite responses to pollution: what
538 we know and where we go in 'Environmental Parasitology'. *Parasites & Vectors*, 10, 65.
- 539 Vidal-Martínez, V.M., Torres-Irineo, E., Romero, D., Gold-Bouchot, G., Martínez-Meyer, E.,
540 Valdés-Lozano, D., & Aguirre-Macedo, L. (2015). Environmental and anthropogenic factors
541 affecting the probability of occurrence of *Oncomegas wagneri* (Cestoda: Trypanorhyncha) in
542 the southern Gulf of Mexico. *Parasites & Vectors*, 8, 609.
- 543 Walter, T.C., & Boxshall, G. (2018). *World of Copepods database*. World Register of Marine Species.
- 544 Watson, D.E. & Thorson, T.B. (1976). Helminths from elasmobranchs in Central American fresh
545 waters. In: *Investigations of the ichthyofauna of Nicaraguan lakes*, Thomas B. Thorson (ed.).
546 University of Nebraska-Lincoln, 52 pp.

547 Wilkinson, T., Wiken, E., Bezaury-Creel, J., Hourigan, T., Agardy, T., Herrmann, H., Janishevski,
548 L., Madden, C., Morgan, L., & Padilla, M. (2009). *Ecorregiones marinas de América del Norte*.
549 Montreal: Comisión para la Cooperación Ambiental, 200 pp.

550 Yamaguti, S. (1959). *Systema helminthum. Vol. II. The cestodes of vertebrates*. Interscience
551 Publishers. 860p.

552 Yamaguti, S. (1961). *Systema helminthum. Volume III. The nematodes of vertebrates*. Interscience
553 Publishers. 679 pp.

554 Yamaguti, S. (1963). *Systema helminthum. Volume IV. Monogenea and Aspidocotylea*. Interscience
555 Publishers. 699 p.

556 Yamaguti, S. (1971). *Synopsis of digenetic trematodes of vertebrates. Vols I and II*. Keigaku
557 Publishing. 1074 p.

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