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9 ORIGINAL ARTICLE / ARTÍCULO ORIGINAL

10 A NEW SPECIES OF *RHINOXENOIDES* (DACTYLOGYRIDAE) PARASITIZING
11 *TRIPORTHEUS ANGULATUS* (SPIX & AGASSIZ, 1829) (CHARACIFORMES)
12 FROM THE JURUÁ RIVER BASIN, BRAZIL

13 UNA NUEVA ESPECIE DE *RHINOXENOIDES* (DACTYLOGYRIDAE) QUE
14 PARASITA A *TRIPORTHEUS ANGULATUS* (SPIX & AGASSIZ, 1829)
15 (CHARACIFORMES) EN LA CUENCA DEL RÍO JURUÁ, BRASIL

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26
27 Running Head: New species of *Rhinoxenoides*

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29
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33 **ABSTRACT**

34 A new species of *Rhinoxenoides* was described parasitizing the gill filaments of
35 neotropical triportheid fishes. The new species was described from *Triportheus angulatus*
36 (Spix & Agassiz, 1829) from the Juruá River, State of Acre, Brazil. *Rhinoxenoides*
37 *cruzeirensis* n. sp. resembles *Rhinoxenoides horacioschneideri* Santos Neto, Costa,
38 Soares & Domingues, 2018, mainly by presenting a coiled male copulatory organ (MCO)
39 with clockwise rings; accessory piece with articulation process extending within the coils
40 to the base of MCO; dorsal anchor with well-developed superficial root, and dorsal bar
41 absent. However, it differs from this by number of the coils of the MCO; accessory piece
42 formed by a straight piece that expands in the middle, where it folds over itself; ventral
43 bar short and robust; dorsal anchor with a well-developed superficial root twice as long
44 as the deep root, with a small wing-shaped extension in the distal portion.

45 **Keywords:** Dactylogyridae – Neotropical region – Parasites of fishes – *Rhinoxenoides* –
46 South America – Triportheidae

47

48 **RESUMEN**

49 Se describió una nueva especie de *Rhinoxenoides* parasitando los filamentos branquiales
50 de peces triportheidos neotropicales. La nueva especie fue descrita de *Triportheus*
51 *angulatus* (Spix & Agassiz, 1829) del río Juruá, estado de Acre, Brasil. *Rhinoxenoides*
52 *cruzeirensis* n. sp. se asemeja a *Rhinoxenoides horacioschneideri* Santos Neto, Costa,
53 Soares & Domingues, 2018, principalmente por presentar organo copulatorio masculino
54 (OCM) con anillos espirales en sentido horario; pieza accesoria con proceso de
55 articulación que se extiende dentro de espirales hasta la base de OCM; ancla dorsal con
56 raíz superficial bien desarrollada y barra dorsal ausente. Pero se diferencia de este por el
57 número de espirales de la OCM; pieza accesoria formada por una pieza recta que se
58 expande en el centro, donde se pliega sobre sí misma; barra ventral corta y robusta; ancla
59 dorsal con una raíz superficial bien desarrollada dos veces más larga que la raíz profunda
60 con una pequeña extensión en forma de ala en la porción distal.

61 **Palabras clave:** Dactylogyridae – Parásitos de peces – Región neotropical –
62 *Rhinoxenoides* – Sudamérica – Triportheidae

63

64 **INTRODUCTION**

65 The Amazon biome boasts the largest hydrographic system in the world and the largest
66 freshwater reserve, containing around 20% of the global freshwater, and is considered

67 one of the richest ecosystems on the planet. The Amazon hydrographic region represents
68 49.29% of the Brazilian territory, covering all or part of the states of Acre, Amazonas,
69 Roraima, Rondônia, Pará, Amapá, Maranhão, Tocantins, and Mato Grosso (Tosta &
70 Coutinho 2015; IBGE, 2019; Silva & Bampi, 2019).

71 Parasite diversity in Amazonian fishes is likely underestimated, given the
72 region's rich ichthyofauna and high levels of endemism. This unique combination
73 supports a remarkable variety of helminth species (Luque *et al.*, 2017). Understanding
74 fish parasite diversity could be an important tool for conserving global biodiversity
75 (Luque *et al.*, 2017).

76 The Juruá River Basin belongs to the great Amazon Basin and is located between
77 the states of Acre and Amazonas. It is drained by extensive rivers in a general southwest-
78 north-east direction, with rivers of great importance throughout its course, such as the
79 Juruá River, which rises in the Peruvian Andes and flows into the Solimões River. This
80 river is one of the main tributaries of the right bank of the Amazon River and is considered
81 one of the most winding in the world, forming huge flood grids and thousands of lakes
82 (ACRE, 2012; Costa *et al.*, 2012; Sousa & Oliveira, 2016, Mota da Silva, 2020).

83 Although there is significant diversity of helminth species recorded in the
84 Amazon, knowledge about parasite richness remains incomplete in certain areas. Most
85 research focuses on the Central and Eastern Amazon, while the southwestern Amazon
86 has received limited attention. Studies in the Juruá River basin have primarily
87 documented parasitic fauna in aquaculture systems, leading to a substantial knowledge
88 gap regarding the biodiversity of parasites associated with fish in natural systems in this
89 southwestern Amazon region (Virgilio *et al.*, 2021).

90 Recent research on the Juruá River has identified five species of
91 monopisthocotylian, including three *Cosmetocleithrum* spp. parasitizing the catfish
92 *Oxydoras niger* (Valenciennes, 1821) in the Juruá River: *Cosmetocleithrum*
93 *basicomplexum* Silva, Meneses, Martins, Cohen, Costa & Justo, 2023, *Cosmetocleithrum*
94 *confusus* Kritsky, Thatcher & Boeger, 1986, *Cosmetocleithrum sacciforme* Silva,
95 Meneses, Martins, Cohen, Costa & Justo, 2023, and *Unibarra juruaensis* Justo, Martins
96 & Cohen, 2023 and *Unibarra paranoplatensis* (Suriano & Incorvaia, 1995) parasitizing
97 *Pimelodus blochii* Valenciennes, 1840 (Justo *et al.*, 2023; Silva *et al.*, 2023).

98 Within the order Characiformes, Triportheidae includes five genera and 23 species
99 (Froese & Pauly, 2024), among which *Triportheus angulatus* (Spix & Agassiz, 1829), a
100 benthopelagic freshwater fish, distributed in the Amazon River basin in South America

101 and exclusive to the Neotropical region (Malabarba, 2004, Froese & Pauly, 2024). These
102 species are exploited by subsistence fishing and have become an alternative source of fish
103 since the natural stocks of other commercially important fish species in the Amazon have
104 declined over time (Cajado *et al.*, 2023).

105 To date, 29 species of Monopisthocotyla have been described in South America
106 from Triportheidae hosts, with the majority of these species classified under the genus
107 *Anacanthorus* Mizelle & Price, 1965 (see Table).

108 In continuation of the studies carried out in the Juruá River Basin in Brazil, aiming
109 to describe the fauna of helminths parasitizing freshwater fish, a new species of
110 *Rhinoxenoides* Santos Neto, Costa, Soares & Domingues, 2018 was found infesting the
111 gills of *T. angulatus*. The new species described here enhances our understanding of the
112 diversity of monopisthocotylian parasites in triportheid fishes, as well as offering valuable
113 insights into the region's biodiversity, and underscoring the importance of expanding
114 research across the Amazon basin.

115

116 MATERIAL AND METHODS

117

118 In January 2024, two specimens of *T. angulatus* were captured with gill nets and a hook
119 and line from Juruá River, Acre, Brazil (7°40'34.1"S, 72°39'39.5"W) (Fig. 1). All
120 collections obtained environmental licensing through the Biodiversity Authorization and
121 Information System (SISBIO, 396871-1). The gills were removed and placed in vials
122 containing hot water (~65°C) and they were then vigorously shaken to detach the parasites
123 from the gill filaments. Absolute ethanol was added to reach a concentration of 70%. The
124 parasites were collected from the sediment and gill arches in the laboratory with the aid
125 of a stereoscopic microscope. The specimens were mounted unstained in Hoyer's
126 medium for the study of the sclerotized parts. Photomicrographs and drawings were taken
127 using a Zeiss® Axioskop microscope micrographic system, with a differential
128 interference contrast (DIC) apparatus and an Olympus BX 41 microscope with phase
129 contrast, equipped with a camera lucida. All measurements are in micrometers, and the
130 means are followed by the range in parentheses and by the number of specimens measured
131 when more than two. Dimensions of organs and other structures represent the greatest
132 distance; lengths of curved or bent structures (bars and accessory piece) represent the
133 straight-line distances between extreme ends. The numbering of hook pairs follows

134 Mizelle & Price (1963). Holotype and paratypes were deposited in the helminthological
135 collection of the Instituto Oswaldo Cruz (CHIOC).

136 **Ethical aspects:** All applicable institutional, national and international guidelines for the
137 care and use of animals were followed.

138

139 **RESULTS**

140

141 **TAXONOMY**

142 Class Monopisthocotyla Brabec, Salomaki, Kolisko, Scholz & Kuchta, 2023

143 Order Dactylogyridea Bychowsky, 1937

144 Dactylogyridae Bychowsky, 1933

145 *Rhinoxenoides* Santos Neto, Costa, Soares & Domingues, 2018

146

147 *Rhinoxenoides cruzeirensis* n. sp. (Fig. 2 A-G)

148 **Type-host:** *Triporthesus angulatus* (Spix & Agassiz, 1829) (Characiformes:
149 Triporthesidae).

150 **Site in host:** Gill lamellae.

151 **Type-locality:** Juruá River, Acre, Brazil (7°40'34.1"S, 72°39'39.5"W).

152 **Parasitological indices:** Total number of hosts: 2; number of infected hosts: 1; total
153 number of parasites: 11.

154 **Deposited material:** Holotype (CHIOC 40615a); Paratypes (CHIOC 40615b-i).

155 **Etymology:** The specific name is dedicated to the municipality of Cruzeiro do Sul, in the
156 state of Acre, Brazil, where the parasite was recovered.

157

158 Description (Based on 11 specimens, mounted in Hoyer's medium). Body fusiform,
159 elongated, comprising cephalic region, trunk, and haptor. Tegument thin, smooth. Body
160 509 (415–625; n = 10) long, 74 (60–85; n = 11) wide at the level of germarium. Anterior
161 region with four cephalic lobes well developed, two terminal, two bilateral; three pairs of
162 head organs; cephalic glands indistinct. Two pairs of eyespots, posterior pair larger and
163 closer together than anterior pair; accessory granules absent (Fig. 2A). Pharynx spherical
164 20 (18–22; n = 6) in diameter. Esophagus short. Two intestinal ceca confluent posteriorly
165 to testis, lacking diverticula. Gonads intercecal, tandem. Germarium ventral to testis, 74
166 (70–80; n = 4) long. Vagina and vaginal canal slightly sclerotized; vaginal aperture
167 sinistroversal. Seminal receptacle spherical. Mehlis' gland, uterus, oviduct, and ootype

168 not observed. Egg 72 long by 45 wide (n = 1). Vitellaria extends throughout the trunk,
169 except in areas of other reproductive organs. Prostatic reservoir elongated posterior to
170 male copulatory organ (MCO), 41 (40–42; n = 3) long. Copulatory complex comprising
171 MCO and accessory piece. Coiled MCO with 1½ clockwise rings, 17 (15–20; n = 5)
172 proximal ring diameter; accessory piece formed by a straight piece that expands in the
173 middle, where it folds over itself; in the proximal portion it articulates with the base of
174 the MCO, 40 (35–44; n = 4) long, through a copulatory ligament (Fig. 2B). Peduncle
175 inconspicuous. Haptor subtriangular, 65 (52–73; n = 8) wide (Fig. 2A). Ventral bar short
176 and wide, 19 (15–22; n = 9) long (Fig. 2C). Dorsal bar absent. Ventral and dorsal anchor
177 dissimilar in size and shape. Ventral anchor with superficial root elongated, well
178 developed; deep root short, rounded; evenly curved shaft, point not passing from the level
179 of the type of superficial root, 69 (65–72; n = 16) long, base 28 (24–31; n = 13) (Fig. 2D).
180 Dorsal anchor with a well-developed superficial root, slender, twice the size of the deep
181 root and a small wing-shaped extension in the distal portion; deep root elongated, rounded
182 at the tip; straight shaft, short point hook-shaped, 55 (53–58; n = 17) long, base 10 (8–11;
183 n = 7) (Fig. 2E). Hooks similar in shape, shank divided into two subunits, distal part of
184 shank inflated; filamentous hook (FH) loop extending close to the beginning of shank
185 dilation, erect thumb, curved long shaft, delicate point. Hook pairs 2, 3, 4, 6, and 7, 21
186 (20–22; n = 8) long (Fig. 2F), hook pairs 1 and 5, 16 (15–16; n = 4) long (Fig 2G).

187

188 **Remarks:** The new species was allocated in *Rhinoxenoides* as it shares characteristics of
189 the genus, such as MCO formed by a coiled tube with clockwise rings articulated to the
190 accessory piece by copulatory ligament; dorsal anchor with superficial root twice the size
191 of the deep root; and by the absence of dorsal bar. *Rhinoxenoides cruzeirensis* n. sp. is
192 similar to *Rhinoxenoides horacioschneideri* Santos Neto, Costa, Soares & Domingues,
193 2018, the only species of the genus, by presenting a well-developed ventral anchor with
194 a superficial truncated root; prostatic reservoir elongated, not divided into regions; vagina
195 sinistral; Coiled MCO with clockwise rings; accessory piece with articulation process
196 extending within coils to the base of MCO; dorsal anchor with well-developed superficial
197 root comprising three times the total length, and dorsal bar with enlarged ends. The new
198 species differs from *R. horacioschneideri* by presenting an Coiled MCO with 1½
199 clockwise rings; accessory piece formed by a straight piece that expands slightly at its
200 center, where it bends into the shape of an elbow (2½ coils; comprising variable distal
201 sheath in *R. horacioschneideri*); ventral bar short and robust (V-shaped in *R.*

202 *horacioschneideri*); dorsal anchor with a well-developed superficial root twice as long as
203 the deep root and a small wing-shaped extension in the distal portion (three times longer
204 than the deep root; small wing-shaped extension in the distal portion absent in *R.*
205 *horacioschneideri*) and by the hook size (1-3 and 5 smaller than the others in *R.*
206 *horacioschneideri*). In the original description, the authors stated that the species present
207 the dorsal anchor with superficial root twice the size of deep root, but in the original
208 drawing the superficial root is shown as three times as long as deep root, feature
209 confirmed by examining the type material deposited in the Helminthological Collection
210 of Oswaldo Cruz Institute CHIOC 39025a (holotype); 39025b-c (paratypes); 39026
211 (paratype), and 39027a-d (vouchers).

212

213 **DISCUSSION**

214 *Rhinoxenoides* Santos Neto, Costa, Soares & Domingues, 2018 was erected by Santos-
215 Neto *et al.* (2018) to accommodate *R. horacioschneideri* described from *Acestrorhynchus*
216 *falcatus* (Bloch, 1794). It is characterized by the morphology of copulatory complex,
217 composed of a coiled tube with clockwise rings articulated to the accessory piece by
218 copulatory ligament; by the morphology of haptoral sclerites as dorsal anchor with the
219 superficial root twice as long as the deep root; straight shaft, curved point; and by the
220 absence of dorsal bar (Santos-Neto *et al.*, 2018). The monotypic genus presents shared
221 characteristics, such as the presence of a copulatory ligament and dorsal anchors with a
222 straight long shaft and absence of a dorsal bar, with *Rhinoxenus* Kritsky, Boeger &
223 Thatcher, 1988, but differ by the coiled MCO with counterclockwise coils, absence of
224 superficial and deep roots in both pairs of anchors, and dorsal anchor with elongate,
225 straight shaft in *Rhinoxenus*. *Protorhinoxenus* Domingues & Boeger, 2002, a genus
226 closely related to *Rhinoxenus* was proposed by Domingues & Boeger (2002). Both genera
227 share morphological features such as a coiled MCO with counterclockwise coils, absence
228 of superficial and deep roots in both pairs of anchors, and dorsal anchors with elongated
229 and straight shafts. *Rhinoxenus* and *Rhinoxenoides* share the character of the absence of
230 a dorsal bar, differing from *Protorhinoxenus* in this feature. According to Santos-Neto *et*
231 *al.* (2018), the absence of dorsal bar could be interpreted as an independent secondary
232 loss in *Rhinoxenoides* and *Rhinoxenus*, or its absence could be shared by the three genera
233 as a synapomorphy with secondary acquisition in *Protorhinoxenus*. These authors
234 proposed a cladistic hypothesis, in which species of *Protorhinoxenus*, *Rhinoxenoides*, and
235 *Rhinoxenus* share the presence of a copulatory ligament and dorsal anchors with a straight

236 long shaft. These features are also observed in the new species of *Rhinoxenoides* proposed
237 herein. The authors stated that species of *Protorhinoxenus*, *Rhinoxenoides*, and
238 *Rhinoxenus* seem to be exclusively found infecting characiform fishes from South
239 America. According to our results, by proposing another species of *Rhinoxenoides*
240 parasitizing a characiform fish belonging to Triportheidae, while the previously reported
241 species of the genus was collected from Acestrorhynchidae, propose that members of
242 these three other genera are not restricted to members of one host family within
243 Characiformes. The finding shows that the parasite faunas of fishes need to be further
244 characterized through additional sampling for monopisthocotylian of species of all
245 characiform families, and that cladistics studies are necessary, to elucidate host-parasite
246 relationships in the freshwater fishes of the Neotropical Region.

247

248 **Author contributions: CRediT (Contributor Roles Taxonomy)**

249

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253

254 **Conceptualization:** SCC

255 **Data curation:** SCC, WMOM, MCNJ

256 **Formal Analysis:** SCC, WMOM, MCNJ

257 **Funding acquisition:** SCC

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259 **Methodology:** WMOM, MCNJ

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262 **Software:** MCNJ

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264 **Validation:** SCC, WMOM, MCNJ

265 **Visualization:** SCC, WMOM, MCNJ

266 **Writing – original draft:** SCC, WMOM, MCNJ

267 **Writing – review & editing:** SCC, MCNJ

268

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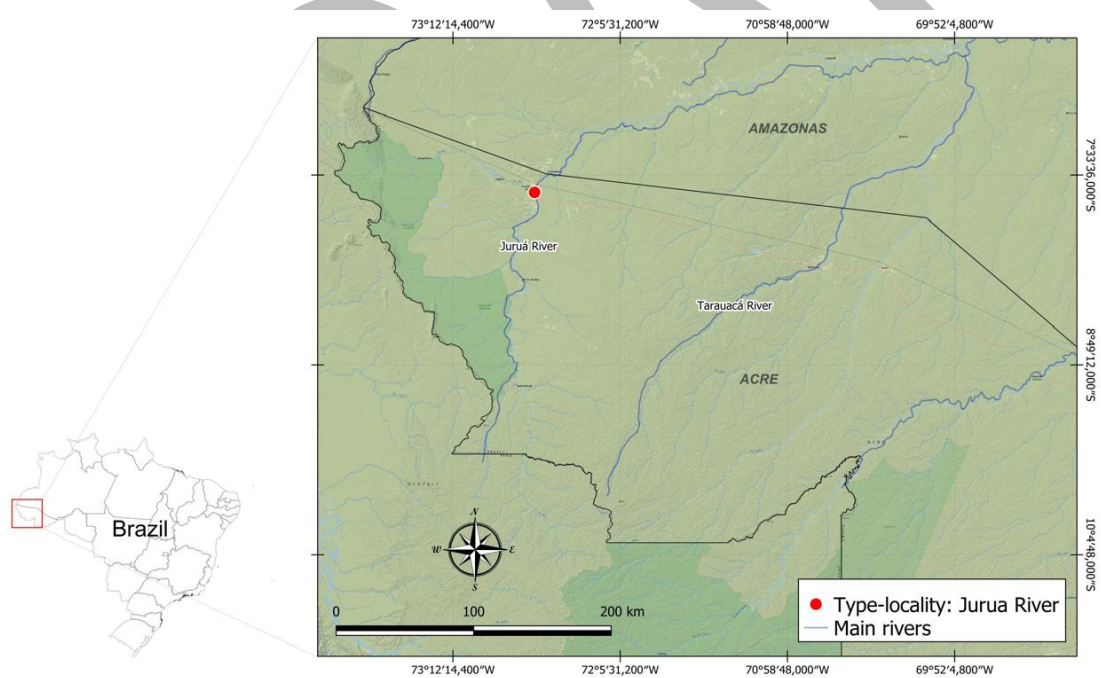
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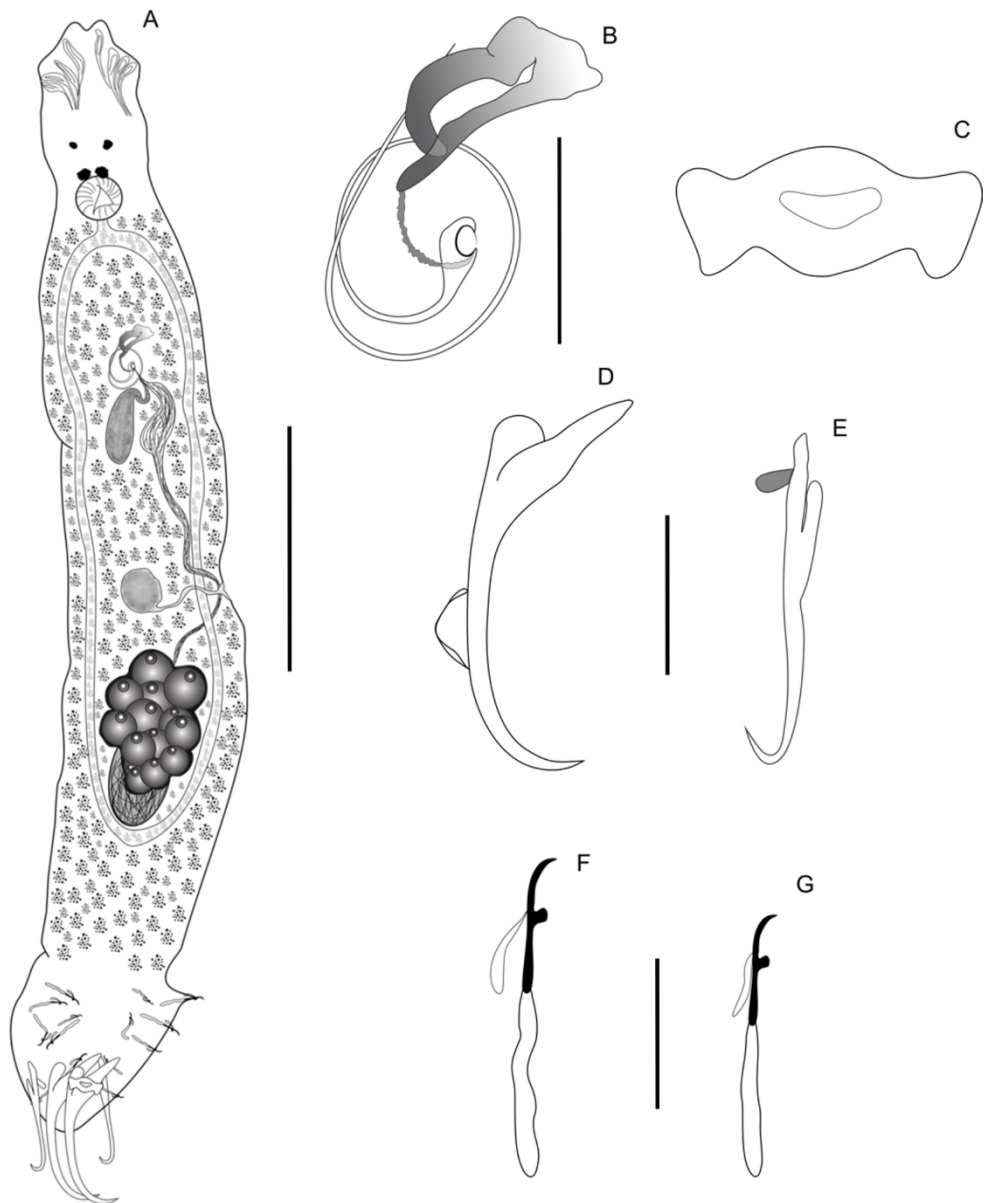
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352 **Figure 1.** Location of the study area, Juruá River, Acre, Brazil (7°40'34.1\"S,
353 72°39'39.5\"W) Cruzeiro do Sul, Acre State, Brazil.



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355 **Figure 2.** *Rhinoxenoides cruzeirensis* n. sp. from *Triportheus angulatus*. A, Total ventral
 356 view (composite); B, Copulatory complex ventral view; C, Ventral bar; D, Ventral
 357 anchor; E, Dorsal anchor; F, Hook pairs 2-4,6,7. G, Hook pairs 1,5. Scale bars: A, 100
 358 μm , B, 20 μm , C, F,G, 10 μm , D, E, 30 μm .

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360

361 **Table 1.** Species of Monopisthocotyla reported in South America from *Tripottheus* spp. hosts.

362

MONOPISTHOCOTLA	HOST	COUNTRY	REFERENCE
<i>Anacanthorus acuminatus</i> Kritsky, Boeger & Van Every, 1992	<i>T. albus</i> , <i>T. angulatus</i> , <i>T. elongatus</i>	BR	Kritsky <i>et al.</i> (1992); Moreira <i>et al.</i> (2017)
<i>Anacanthorus alatus</i> Kritsky, Boeger & Van Every, 1992	<i>T. albus</i> , <i>T. elongatus</i>	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus andersoni</i> Kritsky, Boeger & Van Every, 1992	<i>T. angulatus</i>	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus bellus</i> Kritsky, Boeger & Van Every, 1992	<i>T. albus</i> , <i>T. elongatus</i> , <i>Tripottheus</i> sp.	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus calophallus</i> Kritsky, Boeger & Van Every, 1992	<i>T. elongatus</i>	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus carinatus</i> Kritsky, Boeger & Van Every, 1992	<i>T. angulatus</i>	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus chaunophallus</i> Kritsky, Boeger & Van Every, 1992	<i>T. angulatus</i>	BR	Kritsky <i>et al.</i> (1992); Moreira <i>et al.</i> (2017)
<i>Anacanthorus chelophorus</i> Kritsky, Boeger & Van Every, 1992	<i>T. angulatus</i> , <i>Tripottheus</i> sp.	BR	Kritsky <i>et al.</i> (1992); Moreira <i>et al.</i> (2017)
<i>Anacanthorus cornutus</i> Kritsky, Boeger & Van Every, 1992	<i>T. angulatus</i>	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus euryphallus</i> Kritsky, Boeger & Van Every, 1992	<i>T. albus</i> , <i>T. angulatus</i> , <i>T. elongatus</i>	BR	Kritsky <i>et al.</i> (1992); Moreira <i>et al.</i> (2017)
<i>Anacanthorus formosus</i> Kritsky, Boeger & Van Every, 1992	<i>T. elongatus</i> , <i>Tripottheus</i> sp.	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus furculus</i> Kritsky, Boeger & Van Every, 1992	<i>T. elongatus</i> , <i>T. rotundatus</i>	BR	Kritsky <i>et al.</i> (1992); Santos & Tavares-Dias (2017)
<i>Anacanthorus glyptophallus</i> Kritsky, Boeger & Van Every, 1992	<i>T. angulatus</i>	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus lygophallus</i> Kritsky, Boeger & Van Every, 1992	<i>T. angulatus</i>	BR	Kritsky <i>et al.</i> (1992); Moreira <i>et al.</i> (2017)

<i>Anacanthorus nanus</i> Kritsky, Boeger & Van Every, 1992	<i>T. angulatus</i>	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus pelorophallus</i> Kritsky, Boeger & Van Every, 1992	<i>T. elongatus</i>	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus pithophallus</i> Kritsky, Boeger & Van Every, 1992	<i>T. angulatus</i> , <i>T. curtus</i> , <i>T. rotundatus</i>	BR	Kritsky <i>et al.</i> (1992); Oliveira <i>et al.</i> (2016); Moreira <i>et al.</i> (2017), Santos & Tavares-Dias (2017)
<i>Anacanthorus quinqueramus</i> Kritsky, Boeger & Van Every, 1992	<i>T. albus</i> , <i>T. elongatus</i> , <i>Triportheus</i> sp.	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus ramulosus</i> Kritsky, Boeger & Van Every, 1992	<i>T. albus</i> , <i>T. elongatus</i>	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus strongylophallus</i> Kritsky, Boeger & Van Every, 1992	<i>T. elongatus</i>	BR	Kritsky <i>et al.</i> (1992)
<i>Anacanthorus tricornis</i> Kritsky, Boeger & Van Every, 1992	<i>T. angulatus</i> , <i>T. elongatus</i>	BR	Kritsky <i>et al.</i> (1992)
<i>Ancistrohaptor falcatum</i> Agarwal & Kritsky, 1998	<i>T. elongatus</i>	BR	Agarwal & Kritsky (1998)
<i>Ancistrohaptor falciferum</i> Agarwal & Kritsky, 1998	<i>T. albus</i> , <i>T. angulatus</i> , <i>T. elongatus</i> , <i>Triportheus</i> sp.	BR	Agarwal & Kritsky (1998); Moreira <i>et al.</i> (2017)
<i>Ancistrohaptor falcunculum</i> Agarwal & Kritsky, 1998	<i>T. albus</i> , <i>T. angulatus</i> , <i>T. elongatus</i>	BR	Agarwal & Kritsky (1998); Moreira <i>et al.</i> (2017)
<i>Ancistrohaptor forficata</i> Diniz, Sousa, Yamada & Yamada, 2025	<i>T. signatus</i>	BR	Diniz <i>et al.</i> (2025)
<i>Jainus iquitensis</i> Morey, Viana, Chota & Chero, 2025	<i>T. angulatus</i>	PE	Morey <i>et al.</i> (2025)
<i>Jainus loretoensis</i> Morey, Viana, Chota & Chero, 2025	<i>T. angulatus</i>	PE	Morey <i>et al.</i> (2025)
<i>Jainus sardinae</i> Morey, Viana, Chota & Chero, 2025	<i>T. angulatus</i>	PE	Morey <i>et al.</i> (2025)
<i>Rhinoxenus anaclaudiae</i> Domingues & Boeger, 2005	<i>T. angulatus</i> , <i>T. nematurus</i> , <i>Triportheus</i> sp.	BR	Domingues & Boeger (2005); Moreira <i>et al.</i> (2017)

363 BR: Brazil; PE: Peru.

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