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ORIGINAL ARTICLE / ARTÍCULO ORIGINAL

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HELMINTHS INFECTING THE BOM JARDIM TOAD *RHINELLA DAPSILIS*

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FROM AN ATLANTIC FOREST ENCLAVE AT NORTHEASTERN BRAZIL

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HELMINTOS INFECTANDO EL SAPO BOM JARDIM *RHINELLA DAPSILIS* DE UN

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ENCLAVE DE LA MATA ATLÁNTICA

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HELMINTOS INFECTANDO O SAPO BOM JARDIM *RHINELLA DAPSILIS* DE UM

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ENCLAVE DA MATA ATLÂNTICA NORDESTE DO BRASIL

16

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Running Head: Helminths infecting *Rhinella dapsilis*

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30

31 ABSTRACT

32 *Rhinella dapsilis* (Myers and Carvalho, 1945) is widespread in Amazonia, from  
33 Colombia to Brazil. Despite its huge distribution, information on basic biological  
34 aspects is scarce, even more regarding helminths parasites. Herein, we present  
35 data on helminth infecting *R. dapsilis* from an Atlantic Forest enclave from  
36 Northeastern Brazil. Ten taxa were found, being nine nematodes and one  
37 acanthocephalan, *Cosmocercidae* and *Rhabdias* sp. was the most prevalent taxa.  
38 We reported six new host records and expanded the knowledge about parasitism  
39 in Neotropical anurans.

40 **Keywords:** Amphibian – Bufonidae – highland marshes – Parasites

41

42 RESUMEN

43 *Rhinella dapsilis* (Myers and Carvalho, 1945) es ampliamente distribuida desde  
44 Colombia hasta Brasil. Sin embargo, a pesar de su amplia distribución, las  
45 informaciones al respecto de los aspectos básicos de su biología son escasos,  
46 especialmente en cuanto a los parásitos helmínticos. Presentamos datos de  
47 infecciones por helmintos en *R. dapsilis* de una parte de la Mata Atlántica en el  
48 Nordeste brasileño. Diez taxones fueron encontrados, entre ellos, nueve  
49 nematodos y uno acantocéfalo. *Cosmocercidae* y *Rhabdias* sp. siendo los más  
50 frecuentes. Presentamos seis nuevos registros, expandiendo el conocimiento  
51 sobre el parasitismo para los anuros Neotropicales.

52 **Palabras-clave:** Anfibios – Bufonidae – Pantano de altitud – Parásitos

53

54 RESUMO

55 *Rhinella dapsilis* (Myers and Carvalho, 1945) é amplamente distribuída na  
56 Amazônia, desde a Colômbia até o Brasil. Apesar da ampla distribuição,  
57 informações sobre aspectos básicos de sua biologia são escassas,  
58 especialmente quanto a parasitas helmínticos. Aqui apresentamos dados de  
59 helmintos infectando *R. dapsilis* de um enclave de Mata Atlântica no nordeste  
60 brasileiro. Dez táxons foram encontrados, sendo 9 nematódeos e um  
61 acantocéfalo. Cosmocercidae e *Rhabdias* sp. foram os táxons mais  
62 prevalentes. Apresentamos seis novos registros e expandimos o  
63 conhecimento acerca de parasitismo para anuros neotropicais.

64 Palavras-chave: Anfíbios – Bufonidae – Brejos de altitude – Parasitas

65

66

INTRODUCTION

67 *Rhinella dapsilis* (Myers and Carvalho, 1945), is a leaf-litter toad widely  
68 distributed in Amazonia, occurring in Colombia, Peru, Ecuador and Brazil (Ávila  
69 *et al.*, 2018; Frost, 2024). In Brazil, *R. dapsilis* occurs in Amazonia at Acre,  
70 Amapá, Amazonas and Pará states, also being recorded in transition zones at  
71 Cerrado (Goiás, Maranhão and Tocantins states) and Atlantic Forest  
72 (Pernambuco and Alagoas states), including in a rainforest enclave inside  
73 Caatinga domain in Ceará state (Fouquet *et al.*, 2024). As many of the species  
74 within the *R. margaritifera* group, *R. dapsilis* have serious taxonomic problems,  
75 with differentiation between *R. margaritifera* being extremely difficult (Caramaschi  
76 & Pombal, 2006; Vaz-Silva *et al.*, 2015; Ávila *et al.*, 2018; Pereyra *et al.*, 2021;  
77 Fouquet *et al.*, 2024). Despite this huge distribution, very little is known about its  
78 basic biological aspects, with habitat use, diet and defensive behavior already

79 published (Brito *et al.*, 2013).

80 To date, 25 helminths are known to infect *R. dapsilis* (most of the records under  
81 the name *R. margaritifera*) from Peru, Ecuador and Brazil (Campião *et al.*, 2014;  
82 Willkens *et al.*, 2016; Chero *et al.*, 2023). Those records, however, are restricted  
83 to Amazonia, without studies from Atlantic Forest populations. This lack of  
84 knowledge difficult the understanding of host-parasite relationships (Campião *et*  
85 *al.*, 2009; Santos & Amato, 2010; Santos & Borges-Martins, 2013). Herein, we  
86 examined the helminths infecting *R. dapsilis* from an Atlantic Forest enclave in  
87 Northeastern Brazil.

## 88 MATERIAL AND METHODS

89 Specimens of *R. dapsilis* were collected at the Guaramiranga municipality (4° 15'  
90 46' S, 38° 55' 58' O), located at the Baturité massif, a rainforest enclave within  
91 Caatinga Domain, Brazil. The climate is classified as tropical sub-hot humid and  
92 tropical hot humid, with 1,737.5 mm of rainfall per year and average temperatures  
93 of 24° to 26°. The rainy season occurs from January to May and the vegetation is  
94 characterized by evergreen and seasonal evergreen forest, classified as a  
95 remnant of the Atlantic Forest (IPECE, 2017; Silvera *et al.*, 2020).

96 Specimens of *R. dapsilis* were collected from 2019 to 2024 through active  
97 searches or pitfall traps with drift fences and deposited at Coleção Herpetológica  
98 da Universidade Federal do Ceará (CHUFC-A 9975, 12286-89, 12664-68, 12670,  
99 12283-85, 12290-93). Eighteen individuals were collected, euthanized with a lethal  
100 injection of lidocaine (collecting permit SISBIO 29613-1).

101 The snout-vent length (SVL) of the specimens were measured with a digital  
102 caliper, necropsied through a midventral incision and all organs and body cavity  
103 were dissected for the search of parasitic helminths. The helminths were collected

104 and preserved according to Amato *et al.* (1991) and Andrade (2000). For  
105 identification, the nematodes were clarified in lactic acid, while acanthocephalans  
106 were stained according to the hydrochloric carmine protocols and then clarified  
107 with eugenol on temporary slides (Amato *et al.*, 1991). Helminth identification was  
108 performed following Yamaguti (1961), Sprent (1978), Vicente *et al.* (1991),  
109 Anderson (2000) and Gibbons (2010).

110 Slides were analyzed under a microscope with a computerized image analysis  
111 system. Voucher helminths were deposited at Coleção Parasitológica of the  
112 Universidade Federal do Ceará. Parasitological descriptors (prevalence, mean  
113 intensity of infection and abundance) were calculated according to the  
114 specifications of Bush *et al.* (1997).

115 **Ethic aspects:** This study was approved by the Animal Ethics Committee of the  
116 Universidade Federal do Ceará (CEUA-UFC, process # CEUA 6314010321).

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118

## RESULTS

119 A total of 770 helminths were found infecting *R. dapsilis*, comprising 10 taxa. The  
120 helminths belonged to the phylum Nematoda (767 individuals from 9 taxa) and  
121 Acanthocephala (3 individuals from 1 taxa). The parasites showed an overall prevalence  
122 of 100%, mean overall infection intensity of  $45.33 \pm 6.40$  and mean overall abundance of  
123  $45.33 \pm 6.40$ . The highest helminth abundance was Cosmocercidae, followed by  
124 *Rhabdias* sp. and *Oxysomatium petrolinensis* (Félix-Nascimento 2020) (Table 1). Most  
125 endoparasites found to *R. dapsilis* were located on the lungs (n= 324), followed by the  
126 large intestine (n= 295). By contrast, the body cavity and bladder were less infected with a  
127 total of 4 and 8 endoparasites, respectively. Six taxa were reported for the first time  
128 infecting *R. dapsilis* (Table 1).

129

## DISCUSSION

130 Most of the parasites found here belong to the family Cosmocercidae. Like many studies  
131 (Lins *et al.*, 2017; Oliveira *et al.*, 2019), Cosmocercidae gen. sp. is not identified due to  
132 difficulties in determining morphological characters or the absence of males, in which  
133 reproductive organs are essential for species determination. The species of the family,  
134 including the identified species here (*Aplectana membranosa* (Schneider 1866),  
135 *Cosmocerca parva* (Travassos, 1925) and *Oxysomatium petrolinensis*) have a direct life  
136 cycle, and their hosts become infected by ingesting infective larvae or actively  
137 penetrating larvae present in the soil (Anderson, 2000). Of the species found here, only  
138 *C. parva* have been already reported for *R. dapsilis* (as *R. margaritifera*; Campião *et al.*,  
139 2014). *A. membranosa* have been reported infecting other six bufonids (Campião *et al.*,  
140 2014) and *O. petrolinensis* were recently described infecting *Leptodactylus macrosternum*  
141 (Miranda-Ribeiro, 1926). Other cosmocercids reported infecting *R. dapsilis* (= *R.*  
142 *margaritifera*) are *Aplectana hylambatis* (Baylis, 1927), *Cosmocerca podicipinus* (Bursey  
143 *et al.*, 2001), and *Cosmocerca* sp. and *Raillietnema ibañezi* (Cordova, 1998) (Campião *et*  
144 *al.*, 2014; Chero *et al.*, 2023).

145 Nematodes of the genus *Rhabdias* are lung parasites of amphibians and reptiles, which  
146 directly infects its hosts by active penetration into the skin (Langford & Janovy, 2009;  
147 Kuzmin *et al.*, 2022). Identification is very difficult due to the high morphological similarity,  
148 and Müller *et al.* (2018) strongly suggests the use of molecular data for species  
149 recognition. This is the first record of *Rhabdias* sp. infecting *R. dapsilis*, although *R.*  
150 *sphaerocephala* have been already reported (Chero *et al.*, 2023).

151 Nematodes of the genus *Physaloptera* have been recorded infecting the stomachs of  
152 mammals, fish, reptiles, and amphibians (Anderson, 2000; Macedo *et al.*, 2023). They are  
153 usually found in larval stages parasitizing amphibians that act as intermediate or paratenic  
154 hosts (Anderson, 2000). This is the first record of larvae of *Physaloptera* sp. infecting *R.*  
155 *dapsilis*, but *Physaloptera retusa* was already reported (Campião *et al.*, 2014).

156 Nematodes of the genus *Strongyloides* can either infect their hosts direct or indirectly  
157 (Graham *et al.*, 2023). Infection occurs through skin penetration or ingestion of infected

158 prey (Mati & Melo, 2014; Hallinger *et al.*, 2020). Other bufonids have already been  
159 infected by *Strongyloides* spp. (Campião *et al.*, 2014). This is the first record of  
160 *Strongyloides* sp. infecting *R. dapsilis*.

161 *Oswaldocruzia* spp. also infects its hosts directly by ingestion of eggs or larval penetration  
162 of the host's skin (Kirillova, 2020). Many amphibians have been reported as hosts of *O.*  
163 *mazzai* (Travassos, 1935), including *R. dapsilis* (Campião *et al.*, 2014). Besides *O.*  
164 *mazzai*, other four species of *Oswaldocruzia* have been reported infecting *R. dapsilis*: *O.*  
165 *albareti* (Slimane & Desset, 1996), *O. chambrieri* (Slimane & Desset, 1993), *O. lescurei*  
166 (Slimane & Desset, 1996), *O. lopesi* (Gonçalves *et al.*, 2002), *O. proencai* (González and  
167 Hamann, 2008), *O. subauricularis* (Vicente *et al.*, 1991) (Campião *et al.*, 2014; Chero *et al.*,  
168 2023).

169 The genus *Centrorhynchus* infects birds of the Falconiformes and Strigiformes orders as  
170 definitive hosts and has invertebrates as intermediate hosts, with reptiles and amphibians  
171 serving as paratenic hosts (Oliveira *et al.*, 2024). Cystacanths of *Centrorhynchus* sp. are  
172 found in several amphibians, including bufonids (Campião *et al.*, 2014). Previously, only  
173 unidentified cystacanths have been reported infecting *R. dapsilis* (Chero *et al.*, 2023).

174 The genus *Foleyella* currently has three species infecting frogs in South America, *F.*  
175 *convoluta* (Walton, 1935), *F. scalaris* (Walton, 1935) and *F. vellardi* (Travassos, 1929) that  
176 have been recorded infecting frogs of the Hylidae, Leptodactylidae and Bufonidae in Brazil.  
177 In our study, we found a single female specimen, which makes the identification difficult at  
178 species level.

179 Despite the recent increase in studies dealing with helminth parasites of Neotropical  
180 amphibians (Mascarenhas *et al.*, 2021; Quirino *et al.*, 2023), the majority of the species are  
181 still unstudied (Campião *et al.*, 2015). Even in widely distributed species with available  
182 information of helminths, new records of parasites are often published (Campião *et al.*, 2014;  
183 2015). In our study, 60% of the helminths are new records for *R. dapsilis*, raising to 31  
184 helminth taxa actually known to infect this toad. This highlights the need for more studies  
185 dealing with helminths infecting amphibians, in order to expand the knowledge in the

186 Neotropical region.

187

188 **Author contributions: CRediT (Contributor Roles Taxonomy)**

189 **VHH** = Vitoria Hellen Holanda

190 **DBO** = Dalilange Batista Oliveira

191 **RWA** = Robson Waldemar Ávila

192

193 **Conceptualization:** VHH

194 **Data curation:** VHH, DBO, RWA

195 **Formal Analysis:** VHH, DBO, RWA

196 **Funding acquisition:** RWA

197 **Investigation:** VHH, DBO, RWA

198 **Methodology:** VHH, RWA

199 **Project administration:** VHH

200 **Resources:** VHH, DBO, RWA

201 **Software:** VHH, RWA

202 **Supervision:** RWA, DBO

203 **Validation:** VHH, DBO, RWA

204 **Visualization:** VHH, DBO, RWA

205 **Writing – original draft:** VHH, RWA

206 **Writing – review & editing:** RWA, DBO

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208

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BIBLIOGRAPHIC REFERENCES



214 Amato, J.F.R., Boeger, W.A., & Amato, S.B. (1991). *Protocolos para laboratório-coleta*  
215 *e processamento de parasitos de pescado*. Imprensa Universitária, Universidade  
216 Federal Rural do Rio de Janeiro.

217 Anderson, R.C. (eds). (2000). *Nematode parasites of vertebrates, their*  
218 *development and transmission*. Cab. International.

219 Andrade, C.M. (2000). *Meios e soluções comumente empregados em laboratórios*.  
220 Editora Universidade Rural.

221 Ávila, R.W., Pansonato, A., Perez, R., de Carvalho, V.T., Roberto, I.J., Morais,  
222 D.H., de Almeida, A.P., Rojas, R., Gordo, M., & Farias, I.P. (2018). On *Rhinella*  
223 *gildae* Vaz-Silva, Maciel, Bastos & Pombal 2015 (Anura: Bufonidae): Phylogenetic  
224 relationship, morphological variation, advertisement and release calls and  
225 geographic distribution. *Zootaxa*, 4462, 274-290.

226 Brito, L.B., Aguiar, F., Moura-Neto, C., Zucco, C.A., & Cascon, P. (2013). Diet,  
227 activity patterns, microhabitat use and defensive strategies of *Rhinella hoogmoedi*  
228 Caramaschi & Pombal, 2006 from a humid forest in northeast Brazil. *Herpetological*  
229 *Journal*, 23, 29-37.

230 Bush, A.O., Lafferty, K.D., Lotz, J.M., & Shostak, A.W. (1997). Parasitology meets  
231 ecology on its own terms: Margolis *et al.*, revisited. *The Journal of Parasitology*, 83,  
232 575-583.

233 Campião, K.M., Silva, R.J. & Ferreira, V.L. (2009). Helminth parasites of  
234 *Leptodactylus podicipinus* (Anura: Leptodactylidae) from southeastern Pantanal, State  
235 of Mato Grosso do Sul, Brazil. *Journal of Helminthology*, 83, 345-349.

236 Campião, K.M., Morais, D.H., Dias, O.T., Aguiar, A., de Melo Toledo, G., Tavares,  
237 L.E.R., & da Silva, R.J. (2014). Checklist of Helminth parasites of Amphibians from  
238 South America. *Zootaxa*, 3843, 1-93.

239 Campião, K.M., de Aquino Ribas, A.C., Morais, D.H., da Silva, R.J., & Tavares,  
240 L.E.R. (2015). How many parasites species a frog might have? Determinants of  
241 parasitediversity in South American anurans. *PLoS ONE*, *10*, e0140577.

242 Caramaschi, U., & Pombal-Jr, J.P. (2006). A New Species of *Rhinella* Fitzinger, 1826  
243 from the Atlantic Rain Forest, Eastern Brazil (Amphibia, Anura, Bufonidae). *Papéis*  
244 *Avulsos de Zoologia*, *46*, 251-259.

245 Chero, J.D., Cruces, C.L., Cacique, E.R., Ponce, J.A., Iannacone, J., Alvarino, L.,  
246 Sanchez, L., Sáez, G., Lopez, J., & da Silva, R.J. (2023). A Comprehensive Update  
247 on Helminth Parasite Biodiversity and Richness in Peruvian Amphibians. *Diversity*,  
248 *15*, 1169.

249 Fouquet, A., Ferrão, M., Rodrigues, M.T., Werneck, F.P., Prates, I., Moraes, L.J.C.,  
250 Hrbek, T., Chaparro, J.C., Lima, A.P., Perez, R., Pansonato, A., Carvalho, V.T.,  
251 Almeida, A.P., Gordo, M., Farias, I.P., Milto, K.D., Roberto, I.J., Rojas, R.R., Ron,  
252 S.R., & Ávila, R.W. (2024). Integrative species delimitation and biogeography of the  
253 *Rhinella margaritifera* species group (Amphibia, Anura, Bufonidae) suggest an  
254 intense diversification throughout Amazonia during the last 10 million years.  
255 *Systematics and Biodiversity*, *22*, <https://doi.org/10.1080/14772000.2023.2291086>

256 Frost, D.R. (2024). *Amphibian Species of the World: An Online Reference*. Version  
257 6.0. American Museum of Natural History, New York, USA.  
258 <http://research.amnh.org/herpetology/amphibia/index.html>

259 Gibbons, L. (ed.). (2010). *Keys to the Nematode Parasites of Vertebrates*.  
260 Supplementary Volume. CABI International.

261 Graham, E.A., Los Kamp, E.W., Thompson, N.M., Tillis, S.B., Childress, A.L.,  
262 Wellehan, J.F.X., Walden, H.D.S., & Ossiboff, R.J. (2023). Proliferative  
263 strongyloidiasis in a colony of colubrid snakes. *Veterinary Pathology*, *61*, 109-118.

264 Hallinger, M.J., Taubert, A., & Hermosilla, C. (2020). Occurrence of  
265 *Kalicephalus*, *Strongyloides*, and *Rhabdias* nematodes as most common  
266 gastrointestinal parasites in captive snakes of German households and  
267 zoological gardens. *Parasitology Research*, 119, 947-956.

268 IPECE (2017). Perfil Básico Municipal 2017 Fortaleza.

269 Kirillova, N.Y., Kirillov, A.A., Shchenkov, S.V., & Chikhlyayev, I.V. (2020).  
270 *Oswaldocruzia filiformis* sensu lato (Nematoda: Molineidae) from amphibians and  
271 reptiles in European Russia: Morphological and molecular data. *Nature*  
272 *Conservation Research*, 5, 41-56.

273 Kuzmin, Y., du Preez, L., Nel, T., & Svitin, R. (2022). Three new species of *Rhabdias*  
274 Stiles et Hassall, 1905 (Nematoda: Rhabdiasidae) parasitic in *Ptychadena* spp.  
275 (Amphibia: Anura: *Ptychadenidae*) and an identification key to *Rhabdias* spp. from  
276 Afrotropical anurans. *Parasitology International*, 91, 102649.

277 Langford, G.J., & Janovy, J. (2009). Comparative life cycles and life histories of  
278 north american *Rhabdias* spp. (nematoda: Rhabdiasidae): lungworms from  
279 snakes and anurans. *Journal of Parasitology*, 95, 1145-1155.

280 Lins, A.G.S., Aguiar, A., Morais, D.H., Silva, L.A.F., Ávila, R.W., & Silva, R.J.  
281 (2017). Helminth fauna of *Leptodactylus syphax* (Anura: Leptodactylidae) from  
282 Caatinga biome, northeastern Brazil. *Brazilian Journal of Veterinary*  
283 *Parasitology*, 26, 74-80.

284 Mascarenhas, W., Oliveira, C.R., Benício, R.A., Ávila, R.W., & Ribeiro, S.C. (2021).  
285 Nematodes of *Proceratophrys ararype* (Anura: Odontophrynidae), an endemic frog  
286 from the araripe plateau, northeastern Brazil. *Biota Neotropica*, 21, e20201164.

287 Mati, V.L.T., & Melo, A.L. (2014). Some aspects of the life history and morphology  
288 of *Strongyloides ophidiae* Pereira, 1929 (Rhabditida: Strongyloididae) in *Liophis*

289 *miliaris* (Squamata: Dipsadidae). *Neotropical Helminthology*, 8, 203-216.

290 Macedo, L.C., Willkens, Y., Silva, L.M.O., Gardner, S.L., Melo, F.T.V., & dos  
291 Santos, J. N. (2023). "Revisiting the past": a redescription of *Physaloptera retusa*  
292 (Nemata, Physalopteridae) from material deposited in museums and new material  
293 from Amazon lizards. *Revista Brasileira de Parasitologia Veterinaria*, 32, e017422.

294 Müller, M.I., Morais, D.H., Costa-Silva, G.J., Aguiar, A., Ávila, R.W., & da Silva, R.J.  
295 (2018). Diversity in the genus *Rhabdias* (Nematoda, Rhabdiasidae): Evidence for  
296 cryptic speciation. *Zoologica Scripta*, 47, 595-607.

297 Oliveira, C.R., Ávila, R.W., & Morais, D.H. (2019). Helminths Associated with Three  
298 *Physalaemus* Species (Anura: Leptodactylidae) from Caatinga Biome, Brazil. *Acta*  
299 *Parasitologica*, 64, 205-212.

300 Oliveira, R.J., Mascarenhas, C.S., & Müller, G. (2024). *Centrorhynchus* spp.  
301 (Acanthocephala) in South America: new anuran record and checklist of vertebrate  
302 hosts. *Revista Brasileira de Parasitologia Veterinaria*, 33, e015823.

303 Pereyra, M.O., Blotto, B.L., Baldo, D., Chaparro, J.C., Ron, S.R., Elias-Costa, A.J.,  
304 Iglesias, P.P., Venegas, P.J., Thomé, M.T.C., Ospina-Sarria, J.J., Maciel, N.M.,  
305 Rada, M., Kolenc, F., Borteiro, C., Rivera-Correa, M., Rojas-Runjaic, F.J.M.,  
306 Moravec, J., De la Riva, I., Wheeler, W.C., Castroviejo-Fisher, S., Grant, T.,  
307 Haddad, C.F.B., & Faivovich, J. (2021). Evolution in the genus *Rhinella*: A total  
308 evidence phylogenetic analysis of Neotropical True Toads (Anura: Bufonidae).  
309 *Bulletin of the American Museum of Natural History*, 447, 1-156.

310 Quirino, T.F., Batista-Oliveira, D., Calixto, S.M., & Ávila, R.W. (2023). Helminths  
311 que infectan el escuerzo de Carvalho *Odontophrynus carvalhoi* del estado  
312 brasileño de Ceará. *Neotropical Helminthology*, 17, 247-258.

313 Santos, V.G.T., & Amato, S.B. (2010) Helminth fauna of *Rhinella fernandezae*

314 (Anura: Bufonidae) from the Rio Grande do Sul Coastland, Brazil: Analysis the  
315 parasite community. *Journal of Parasitology*, 96, 823-826.

316 Santos, V.G.T., Amato, S.B., & Borges-Martins, M. (2013). Community structure of  
317 helminth parasites of the Cururu toad, *Rhinella icterica* (Anura: Bufonidae) from  
318 southern Brazil. *Parasitology Research*, 112, 1097-1103.

319 Silvera, A.P., Loiola, M.I.B., dos Santos Gomes, V., Lima-Verde, L.W., Oliveira,  
320 T.S., Silva, E.F., Otutumi, A.T., Ribeiro, K.A., da Silva Xavier, F.A., Bruno, M.M.A.,  
321 Souza, S.S.G., & Araújo, F.S. (2020). Flora of Baturite, Ceara: A wet Island in the  
322 Brazilian Semiarid. *Floresta e Ambiente*, 27, e20180320.

323 Sprent, J.F.A. (1978). Ascaridoid nematodes of amphibians and reptiles:  
324 *Gedoelestascaris* n.g. and *Ortleppascaris* n.g. *Journal of Helminthology*, 52,  
325 261-282.

326 Sulieman, Y., Afifi, A., Awad, H.M., & Pongsakul, T. (2015). Helminth parasites of  
327 the subdesert toad, *Amietophrynus (Bufo) xeros* (Anura: Bufonidae). *International*  
328 *Journal of Research*. Granthaalayah, 3, 75-83.

329 Torres, P., & Puga, S. (1996). Occurrence of cystacanths of *Centrorhynchus* sp.  
330 (Acanthocephala: Centrorhynchidae) in toads of the genus *Eupsophus* in Chile.  
331 *Memórias do Instituto Oswaldo Cruz*, 91, 717-719.

332 Travassos, L. (1929). Filaridés des batraciens du Brésil. *Comptes rendus des*  
333 *séances de la Société de Biologie*, 100, 967-968.

334 Vaz-Silva, W., Maciel, N.M., Bastos, R.P., & Pombal, J.P. (2015). Revealing Two  
335 New Species of the *Rhinella margaritifera* Species Group (Anura, Bufonidae): An  
336 Enigmatic Taxonomic Group of Neotropical Toads. *Herpetologica*, 71, 212-222.

337 Vicente, J.J., Rodrigues, H.O., Gomes, D.C., & Pinto, R.M. (1991). Nematoides do  
338 Brasil. Parte II: Nematoides de anfíbios. *Revista Brasileira de Zoologia*, 7, 549-626.

339 Willkens, Y., Santos, A.M., Nascimento, J., Maschio, G.F., & Melo, F.T.V. (2016).  
340 Redescription of *Oswaldocruzia chambrieri* (Strongylida: Molineidae) from  
341 *Rhinella margaritifera* (Anura: Bufonidae) in Caxiuanã National Forest, Brazil.  
342 *Acta Parasitologica*, 61, 567-575.  
343 Yamaguti, S. (1961). *Systema Helminthum – Nematodes*. Interscience Publishers.  
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346 **Table 1.** Parasitological indices of helminths in *Rhinella dapsilis* from Baturité Massif.  
 347 Development stage (DS), Larvae (L), Adult (A), Prevalence values (P%), number of  
 348 endoparasites (NE), mean intensity of infection and standard deviation (MII  $\pm$  SD), mean  
 349 abundance (MA), site of infection (Site): BC=Body cavity; ST= stomach; LI= large  
 350 intestine; SI= small intestine, LU= lungs; UB= Urinary Bladder. \* New record for *R.*  
 351 *dapsilis*.

Endoparasites	DS	P%	NE	MI $\pm$ SD	MA	Site
<b>Acanthocephala</b>						
<i>Centrorhynchus</i> sp.*	L	11.1	3	-	-	BC
<b>Nematoda</b>						
<i>Aplectana membranosa</i> *	A	11.1	9	-	-	SI/ LI
<i>Cosmocerca parva</i>	A	11.1	10	-	-	SI/ LI
Cosmocercidae gen. sp.	A	77.8	292	19.5 $\pm$ 13.5	16.2 $\pm$ 14.3	SI/ LI/UB
<i>Foleyella</i> sp.*	A	5.5	1	-	-	ST
<i>Oswaldocruzia mazzai</i>	A	16.7	51	17 $\pm$ 12.3	2.8 $\pm$ 7.8	LU
<i>Oxysomatium petrolinensis</i> *	A	5.5	84	-	-	LI
<i>Physaloptera</i> sp.	L	61.1	38	2.6 $\pm$ 2.2	1.6 $\pm$ 2.1	ST/SI/ LI
<i>Rhabdias</i> sp.*	A	83.3	273	16.6 $\pm$ 11.2	15.7 $\pm$ 11.6	ST/ LU
<i>Strongyloides</i> sp.*	A	5.5	9	-	-	SI

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