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9 ORIGINAL ARTICLE / ARTÍCULO ORIGINAL

10 ZONOTIC FOCI OF *TOXOCARA* SP.: COMPARISON OF TWO TOURIST REGIONS IN  
11 THE STATE OF SUCRE, VENEZUELA

12  
13 FOCOS ZONÓTICOS DE *TOXOCARA* SP.: COMPARACIÓN DE DOS REGIONES  
14 TURÍSTICAS EN EL ESTADO SUCRE, VENEZUELA

15  
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30

31 Running Head: Zoonotic foci of *Toxocara* sp.: in state of Sucre, Venezuela

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44

45 **ABSTRACT**

46 Toxocariasis is a human infection caused by the larvae of the nematodes *Toxocara canis*  
47 (Werner, 1782) or *Toxocara cati* (Schrank, 1788), which are parasitic in animals, and  
48 represents a serious public health problem. The objective of this study was to investigate the  
49 information and prevalence of parasitic toxocariasis in the state of Sucre, Venezuela,  
50 particularly in the municipalities of Montes and Sucre, in communities considered vulnerable  
51 due to social and economic deficiencies within the epidemiological/sanitary monitoring. In these  
52 municipalities, there are many stray dogs living near homes and maintaining contact with other  
53 domestic dogs. The seroprevalence of *T. canis* was evaluated through the ELISA method. A  
54 seroprevalence of IgG anti-*T. canis* was obtained in the parish of San Juan of 90.12% and in  
55 the parish of Cumanacoa of 82.14%. In the parish of San Juan, when evaluating the prevalence  
56 of parasites by species, *T. canis* was present in 18.37% of the feces of the canines evaluated.  
57 On the other hand, in the parish of Cumanacoa, *T. canis* was present in 9.90% of the canine  
58 samples analyzed; similarly, the presence of *Toxocara* sp. in soils in both parishes. In summary,  
59 the determination of two zoonotic foci of *Toxocara* sp. in the state of Sucre, Venezuela, since  
60 all the factors necessary for the transmission of toxocariasis are present.

61 **Keywords:** Dogs – Epidemiology – Larva migrans – Parasitic disease – *Toxocara canis* –  
62 Zoonosis

63  
64 **RESUMEN**

65 La toxocariasis es una infección humana causada por las larvas de los nematodos *Toxocara*  
66 *canis* (Werner, 1782) o *Toxocara cati* (Schrank, 1788), que son parásitos en los animales, y  
67 representa un grave problema de salud pública. El objetivo de este trabajo fue investigar la  
68 información y prevalencia de toxocariasis parasitaria en el estado Sucre, Venezuela,  
69 particularmente en los municipios de Montes y Sucre, en comunidades consideradas

70 vulnerables por deficiencias sociales y económicas en el monitoreo epidemiológico/sanitario.  
71 En estos municipios hay muchos perros callejeros que viven cerca de las casas y mantienen  
72 contacto con otros perros domésticos. La seroprevalencia de *T. canis* se evaluó mediante el  
73 método ELISA. Se obtuvo una seroprevalencia de IgG anti-*T. canis* en la parroquia San Juan  
74 de 90,12% y en la parroquia Cumanacoa de 82,14%. En la parroquia San Juan, al evaluar la  
75 prevalencia de parásitos por especie, *T. canis* estuvo presente en el 18,37% de las heces de  
76 los caninos evaluados. Por otra parte, en la parroquia Cumanacoa, *T. canis* estuvo presente  
77 en el 9,90% de las muestras caninas analizadas; Asimismo, la presencia de *Toxocara* sp. en  
78 suelos de ambas parroquias. En resumen, la determinación de dos focos zoonóticos de  
79 *Toxocara* sp. en el estado Sucre, Venezuela, ya que están presentes todos los factores  
80 necesarios para la transmisión de la toxocariasis.

81 **Palabras clave:** Epidemiología – Enfermedad parasitaria – Larva migrans – Perros – *Toxocara*  
82 *canis* – Zoonosis

## 84 INTRODUCTION

85 Toxocariasis is a human infection caused by the larvae of the nematodes *Toxocara canis*  
86 (Werner, 1782) or *Toxocara cati* (Schrank, 1788), which are parasitic in animals. The clinical  
87 manifestations of the disease are usually asymptomatic, which contributes to the disease being  
88 neglected. And yet, toxocariasis has been shown to be especially prevalent among children  
89 from socioeconomically disadvantaged populations, both in the tropics and subtropics and in  
90 industrialized nations (Macpherson, 2013; Rostami *et al.*, 2019; Ma *et al.*, 2020).

91 In humans, the infection is acquired accidentally, as there is no normal development of  
92 the parasite in humans, only the larval stage survives as it is a paratenic host in this parasitosis  
93 (Taylor *et al.*, 2017). Infection occurs by ingestion of infective eggs found in poorly washed  
94 vegetables and fruit originating from a contaminated environment, geophagy, ingestion of

95 undercooked paratenic hosts (chicken, beef or lamb), putting contaminated objects in the  
96 mouth, as well as not washing hands after playing with or petting infected dogs (Holland *et al.*,  
97 1995; Vázquez *et al.*, 1997; Ulloque-Badaracco *et al.*, 2023). Children are the most infected  
98 due to their poor hygienic habits and are often in contact with dogs and puppies, as well as  
99 exposed to the pets' living environment (Cabral-Monica *et al.*, 2022).

100 Toxocariasis is a widely distributed zoonosis, in which the human need to keep pets and  
101 companion animals such as dogs and cats around ensures the persistence of the parasite and  
102 infection in humans over time. with rates ranging from 1% in Spain to 86% in Santa Lucia; from  
103 3.6% to 24.7% in Brazil; 47.5% in Colombia; from 34.9% to 66.6% in Venezuela and from  
104 37.9% (Roldán *et al.*, 2010a). In Argentina, a study carried out in children showed a 67.7%  
105 seroprevalence of anti-*Toxocara* IgG antibodies (López *et al.*, 2005), while in Paraguay a 78%  
106 seroprevalence for toxokaryosis was reported (Rivarola *et al.*, 2009). In Peru, a few reports  
107 have informed of frequencies from 7.8% to 44.92% in rural populations from different places  
108 (Roldán *et al.*, 2010a).

109 *Toxocara canis* infection in dogs has worldwide distribution rates ranging from 0 to 99.4%  
110 (Barriga, 1988; Heymann & American Public Health Association, 2004; Borges & Ferreira,  
111 2020). In Venezuela, studies on *T. canis* infection were carried out in urban areas such as  
112 Caracas, reporting prevalences of 66.6%, in rural and suburban communities of the Capital  
113 District and the State of Miranda 20% and 70% (Incani, 1996; Delgado & Rodríguez-Morales,  
114 2009). In the Brazilian Amazon, the prevalence in urban children < 5 years of age has been  
115 reported to be 21.5%, increasing to 26.7% in the rural general population, and to 26.8% or 52%  
116 in the riverine population (Oliart-Guzmán *et al.*, 2014). Furthermore, studies on canine  
117 toxocariasis carried out in Maracaibo reported a prevalence of 11.4% (Ramírez-Barrios *et al.*,  
118 2004) and 63,16% in soil samples from leisure parks (Cazorla-Perfetti *et al.*, 2007). In eastern  
119 Venezuela, particularly in the state of Anzoátegui, (Fernández *et al.*, 2009) carried out a study

120 on children aged 6 to 8 years, obtaining a prevalence of 16%, and (Gómez, 2004) carried out  
121 a study in the Nueva Toledo neighborhood of El Peñón, state of Sucre, on children aged 2 to  
122 15 years, with prevalences of 77.14%.

123 Thus, the present study evaluated the seroprevalence of toxocariasis in the municipalities  
124 of Montes and Sucre, which are regional tourist destinations in Venezuela. These regions were  
125 selected to be considered vulnerable due to social and economic deprivation, the large number  
126 of stray dogs found in the area, and the fact that most family groups have dogs as pets.

127

## 128 MATERIAL AND METHODS

### 129 Study area

130 The present study was conducted in the municipalities of Sucre and Montes, in the state  
131 of Sucre, Venezuela (Fig. 1). The municipality of Sucre is located in a mountainous area with  
132 a hot, semi-arid climate, with average temperatures of 27°C and 440 mm of precipitation with  
133 a rainy season from July to November (Pérez, 2006). The second site was Cumanacoa in the  
134 municipality of Montes. It is characterized by a hot sub-humid climate with temperatures ranging  
135 from 21°C to 27°C and annual precipitation between 900 and 1500 mm.



136

137 **Figure 1.** Map of the state of Sucre, Venezuela with the respective municipalities evaluated:  
138 Sucre (10,24198° N, 64,27980° O), Cumanacoa (10,24720° N, 63,92019° O), Google Maps.

139

#### 140 **Population sample**

141 To calculate the representative sample size, a formula proposed by Martínez (2003) for  
142 finite populations, with a known sampling frame, was used. The prevalence for toxocariosis of  
143 74.14% was considered (Gómez, 2004). In the parish of San Juan, 172 blood and faeces  
144 samples were taken from people who voluntarily attended the survey. The same occurred in  
145 the parish of Cumanacoa, where 100 blood and stool samples were taken, respectively. A  
146 clinical-epidemiological survey was administered to each individual, regardless of gender and  
147 age, in each parish, with prior informed consent (OPAS, 1993).

#### 148 **Human faecal samples**

149 All the people who attended and participated in the collection were asked to provide a  
150 stool sample for the evaluation of the different helminths including *Ascaris lumbricoides*  
151 (Linnaeus, 1758). As an ascaridium, *A. lumbricoides* cross-reacts with *T. canis* when using  
152 immunological techniques using sera to evaluate for anti-*T. canis* antibodies. Serum samples  
153 of individuals presenting with *Ascaris lumbricoides* eggs in their stool was excluded from the  
154 study.

#### 155 **Human blood samples**

156 Blood samples were obtained from the participants by venous puncture and transferred  
157 to the parasitology laboratory at the Instituto de Investigaciones en Biomedicina y Ciencias  
158 Aplicadas "Dr. Susan Tai", Universidad de Oriente (IIBCA-UDO), Venezuela, where they were  
159 processed for serum collection. The collected sera were frozen at -20°C until analysis.

#### 160 **Determination of anti-*T. canis* IgG antibodies by ELISA method**

161 A commercial *Toxocara*-IgG ELISA kit (RIDASCREEN *Toxocara*-IgG), which uses

162 excretion-secretion antigens, was used for serological diagnosis. No absorption of sera with  
163 *Ascaris suum* (Goeze, 1782) antigen was performed. However, all individuals with *A. suum*  
164 eggs in their faeces were excluded from analysis. For the evaluation and interpretation of the  
165 results, the sample was considered as positive for anti-*T. canis* antibodies when its index was  
166 higher than 1.10, as a negative sample, that with an index lower than 0.90 and as an  
167 indeterminate sample that with an index between 0.90 – 1.10 (Fernández *et al.*, 2009).

#### 168 **Dog Fecal Samples**

169 In both parishes, with the consent of the owners the dog fecal samples were collected in  
170 the morning, at midday and in the afternoon, and placed in airtight containers and transported  
171 to the laboratory. If not analyzed on the day of collection, they were stored at 4°C.

#### 172 **Soil samples**

173 Soil samples were collected from the courtyards of homes in communities from the San  
174 Juan and Cumanacoa parishes. These samples were obtained by superficially scraping the  
175 courtyards with a spatula, 5 cm from any faecal matter after a random selection of sites with  
176 bare soil. Soil samples were collected upto 10 cm deep and stored in labeled polyethylene bags  
177 at room temperature until processing (Leventhal & Cheadle, 1992).

#### 178 **Parasitological diagnosis**

179 Small portions of all the stool samples were examined by direct microscopic examination  
180 after mixing with 0.85% physiological saline and Lugol's solution (Botero & Restrepo, 1998). In  
181 addition, concentration and enrichment techniques such as: spontaneous tube sedimentation  
182 and flotation by Willis-Malloy were applied (Quinn *et al.*, 1980; Camacho *et al.*, 2006). It was  
183 also carried out with soil samples taken in each parish. These examinations were also done on  
184 soil samples taken in each parish.

#### 185 **Statistical analysis**

186 The results obtained were expressed as percentages and represented in tables and



187 graphs. The seroprevalence of *T. canis* infection was determined by applying the formula  
 188 described by Gordis (2004). The Chi-square test ( $\chi^2$ ) was used to establish the association  
 189 between the clinical-epidemiological aspects evaluated and the seroprevalence of *T. canis* in  
 190 the inhabitants of San Juan and Cumanacoa parishes (Martínez, 2003). Data was analyzed  
 191 using SPSS 18 software.

192 **Ethics aspects:** This research was approved by the IIBCAUDO Bioethics and Biosafety  
 193 Committee, (CBBIIBCAUDO-2009-09-3) year 2009, September, project 3.

194

195 **RESULTS**

196 The seroprevalence of *T. canis* was evaluated using the *Toxocara* IgG commercial  
 197 kit. Of the 172 serum samples analyzed from the parish of San Juan, 10 samples were  
 198 discarded because they belonged to people who had *A. lumbricoides* eggs in their faeces.  
 199 Of the remaining 162 sera, 146 (90.12%) were positive, 9 (5.56%) indeterminate and 7  
 200 (4.32%) negatives. The average sample index of the positive cases was 3.03 of the  
 201 indeterminate 0.99 and of the negative 0.55 (table 1). From the 100 serum samples  
 202 analyzed from the Cumanacoa parish, Montes municipality, 16 sera were discarded because  
 203 they belonged to people who had *A. lumbricoides* eggs in their stool samples. Of the 84  
 204 remaining sera, 69 (82.14%) were positive, 6 (7.14%) indeterminate and 9 (10.17%) negatives,  
 205 the average sample rate for positive cases was 2.18, for indeterminate cases 0.96 and for  
 206 negative cases 0.60 (table 1).

207 **Table 1.** Seroprevalence and summary statistics of sample rates for IgG anti-*T. canis* antibodies in  
 208 inhabitases of San Juan and Cumanacoa (Sucre and Montes municipality), Sucre state, Venezuela.

Inhabitants of San Juan / Cumanacoa						
Samples	n.	%	$\bar{X}$	SD	LL	UL
Positive	146/69	90,12/82,14	3,03/2,18	1,22/0,94	1,20/1,13	7,74/5,04
Indeterminate	9/6	5,56/7,14	0,99/0,96	0,08/0,04	0,91/0,92	1,10/1,02

Negative	7/9	4,32/10,17	0,55/0,60	0,14/0,21	0,37/0,34	0,73/0,89
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209 n: number of cases,  $\chi$ : mean, SD: standard deviation, LL: lower limit, UL: upper bound %:  
 210 percentage.

211

212 To understand the epidemiology of toxocariasis in both parishes of Sucre municipality,  
 213 Sucre state, Venezuela, the seroprevalence of anti-*T. canis* IgG was analyzed according to  
 214 gender; it was found that females were more affected in both San Juan and Cumanacoa  
 215 parishes. However, the chi-square test did not show a statistically significant association (table  
 216 2).

217

218 **Table 2.** Association between genders of inhabitants and seroprevalence of anti-*Toxocara canis* IgG  
 219 in San Juan and Cumanacoa (Sucre and Montes municipality), Sucre state, Venezuela.

IgG Anti- <i>Toxocara canis</i> in Inhabitants of San Juan / Cumanacoa								
Gender	Positives		Indeterminate		Negatives		Total	
	n	%	n	%	n	%	n	%
Females	102/45	69.86/65,22	7/3	77.80/50	2/6	28.57/66.67	111/54	68.52/64.29
Males	44/24	30.14/34,78	2/3	22.20/50	5/3	71.43/33.33	51/30	31.48/35.71
Total	146/69	100/100	9/6	100/100	7/9	100/100	162/84	100/100
$\chi^2 = 5,66$ ns $p = 0,06$ / $\chi^2 = 0,58$ ns $p = 0,75$								

220 n: number of cases    %: percentage     $\chi^2$ : chi-square    ns: not significant    p: probability

221

222 Table 3 show the seroprevalence of anti-*T. canis* IgG and its distribution according to  
 223 age group, with significant results in the Cumanacoa and San Juan parishes where the 0-10  
 224 age group was the most affected. Curiously, the chi-square test showed no significant  
 225 association within this age group from the San Juan parish, which was also the most affected.

226 **Table 3.** Association between age and seroprevalence of anti-*Toxocara canis* IgG in  
 227 inhabitants of San Juan and Cumanacoa (Sucre and Montes municipality), Sucre state,  
 228 Venezuela.

IgG Anti- <i>Toxocara canis</i> in Inhabitants of San Juan / Cumanacoa								
Age (years)	Positives		Indeterminate		Negatives		Total	
	n	%	n	%	n	%	n	%
0-10	55/38	37,67/55,07	5/4	55,56/66,67	6/4	85,71/44,44	66/46	40,74/54,76
11-20	31/13	21,23/18,84	1/0	11,11/0	0/0	0/0	32/13	19,75/15,48
21-30	21/9	14,38/13,04	1/0	11,11/0	0/3	0/33,33	22/12	13,59/14,29
31-40	13/1	8,90/1,45	0/0	0/0	1/1	14,29/11,11	14/2	8,64/2,38
41-50	4/1	2,74/1,45	0/2	0/33,33	0/1	0/11,11	4/4	2,47/4,76
51-60	11/4	7,53/5,80	2/0	22,22/0	0/0	0/0	13/4	8,02/4,76
61-70	7/3	4,80/4,35	0/0	0/0	0/0	0/0	7/3	4,32/3,57
71-80	3/0	2,05/0	0	0/0	0/0	0/0	3/0	1,85/0
81-90	1/0	0,70/0	0	0/0	0/0	0/0	1/0	0,62/0
Total	146/69	100/100	9/6	100/100	7/9	100/100	162/84	100/100

$\chi^2 = 12.80$  ns     $p = 0.69$

229 n: number of cases    %: percentage     $\chi^2$ : chi-square    ns: not significant    \*s: significant  
230 p: probability

231

232 Table 4 show the seroprevalence of anti-*T. canis* IgG and its distribution according to  
233 handwashing practices before eating, food preparations and contact with soil by individuals in  
234 both parishes. There was no statistically significant association with any of the variables  
235 assessed by the chi-square test. This was contrary to the self-declared food handling and  
236 handwashing practices. Majority of those testing positive for anti-*T. canis* IgG antibodies  
237 individuals declared washing their hands before eating (84.93% San Juan parish) and (92.75%  
238 Cumanacoa parish) or cleaning their food together with handwashing before consumptio  
239 (75.34% for San Juan parish, and 85.51% for Cumanacoa parish). We also disclose that at the  
240 time of the surveys many of the people, particularly children, had dirty fingernails. In addition,  
241 if they washed their hands or wash their food (fruits and vegetables), they did so using water  
242 from the rivers that transversed these parishes. The water used was neither treated nor boiled,  
243 further compounding to the environmental contamination of these uncooked foodstuffs with *T.*  
244 *canis* eggs and other parasites. The results from both parishes were comparable.

245 **Table 4.** Association between hand washing practices before eating, foodstuff  
246 handling parctices, contact with soil and seroprevalence of anti-Toxocara canis IgG in

247 inhabitants of San Juan and Cumanacoa (Sucre and Montes municipality), Sucre  
 248 state, Venezuela

IgG ANTI- <i>Toxocara canis</i> in Inhabitants of San Juan / Cumanacoa								
	Positives		Indeterminate		Negatives		Total	
	n	%	n	%	n	%	n	%
Hand washing before eating								
Yes	124/64	84,93/92,75	7/5	77,78/83,33	7/8	100/88,89	138/77	85,19/91,67
No	22/5	15,07/71,25	2/1	22,22/16,67	0/1	0/11,11	24/7	14,81/8,33
Total	146/69	100/100	9/6	100/100	7/9	100/100	162/84	100/100
$\chi^2= 1,62$ ns $p= 0,44$ / $\chi^2= 0,74$ ns $p= 0,69$								
Foodstuff washing and cleaning								
Yes	110/59	75,34/85,51	7/4	77,78/66,67	5/7	71,43/77,78	122/70	75,31/83,33
No	36/10	24,66/14,49	2/2	22,22/66,67	2/2	28,57/22,22	40/14	24,69/16,67
Total	146/69	100/100	9/6	100/100	7/9	100/100	162/84	100/100
$\chi^2= 0,09$ ns $p= 0,96$ / $\chi^2= 1,63$ ns $p= 0,44$								
Contact with soil								
Yes	92/47	63,01/68,12	7/3	77,78/50	6/4	85,71/44,44	105/54	64,81/64,29
No	54/22	36,99/31,88	2/3	22,22/50	1/5	14,29/55,56	57/30	35,19/35,71
Total	146/69	100/100	9/6	100/100	7/9	100/100	162/84	100/100
$\chi^2= 2,21$ ns $p= 0,33$ / $\chi^2= 2,52$ ns $p= 0,28$								

249 n: number of cases %: percentage  $\chi^2$ : chi-square ns: not significant p: probability

250

251 An analysis of anti-*T. canis* IgG seroprevalences and their distribution with regard to  
 252 presence and contact with dogs by the participants from the two parishes is shown in table 5.  
 253 The sero-conversion status was not associated with the contact with dogs.

254 **Table 5.** Association between the presence of dogs, contact with dogs and the  
 255 seroprevalence of anti-*Toxocara canis* IgG in inhabitants of San Juan and Cumanacoa  
 256 (Sucre and Montes municipality), Sucre state, Venezuela.

IgG ANTI- <i>Toxocara canis</i> in Inhabitants of San Juan / Cumanacoa								
	Positives		Indeterminate		Negatives		Total	
	n	%	n	%	n	%	n	%
Dog present								
Yes	99/23	67,81/33,33	8/3	66,89/50	4/6	57,14/66,67	111/32	68,52/38,10
No	47/46	32,19/66,67	1/3	11,11/50	3/3	42,86/33,33	51/52	31,48/61,90
Total	146/69	100/100	9/6	100/100	7/9	100/100	162/84	100/100
$\chi^2= 2,19$ ns $p= 0,33$ / $\chi^2= 4,14$ ns $p= 0,13$								
Contact with dogs								
Yes	60/48	41,10/69,57	7/6	77,78/100	3/4	42,86/44,44	70/58	43,20/69,04

No	86/21	58,90/30,43	2/0	22,22/0	4/5	57,14/55,56	92/26	56,80/30,96
Total	146/69	100/100	9/6	100/100	7/9	100/100	162/84	100/100
$\chi^2= 4,659$ ns $p= 0,09$ / $\chi^2= 5,25$ ns $p= 0,07$								

257 n: number of cases   %: percentage    $\chi^2$ : chi-square   ns: not significant   p: probability

258

259 These tables show the seroprevalence of anti-*T. canis* IgG and its distribution according

260 to age, deworming and time of deworming of pet dogs of the participants from in both parishes.

261 However, there were no significant associations established (Table 6).

262 **Table 6.** Role of age, deworming status, and time of deworming of pet dogs in the anti-Toxocara

263 canis IgG seroprevalence of inhabitants of San Juan and Cumanacoa (Sucre and Montes

264 municipality), Sucre state, Venezuela.

IgG ANTI- <i>Toxocara canis</i> in Inhabitants of San Juan / Cumanacoa								
Variables	Positives		Indeterminate		Negatives		Total	
	n	%	n	%	n	%	n	%
<b>Age of dog</b>								
6 months-1 year	41/13	41,41/56,52	4/0	50/0	4/3	100/50	49/16	44,14/50
Older than 1 year	58/10	58,59/43,48	4/3	50/100	0/3	0/50	62/16	55,86/50
Total	99/23	100/100	8/3	100/100	4/6	100/100	111/32	100/100
$\chi^2= 5,47$ ns $p= 0,06$ / $\chi^2= 3,39$ ns $p= 0,18$								
<b>Received dewormer</b>								
Yes	39/14	39,40/60,87	3/2	37,60/33,33	1/5	25/83,33	43/21	38,74/65,63
No	60/9	60,60/39,13	5/1	62,50/66,67	3/1	75/16,67	68/11	61,26/34,37
Total	99/23	100/100	8/3	100/100	4/6	100/100	111/32	100/100
$\chi^2= 0,34$ ns $p= 0,84$ / $\chi^2= 1,07$ ns $p= 0,59$								
<b>Deworming time</b>								
At birth	10/0	10,10/0	1/0	12,50/0	0/0	0/0	11/0	9,91/0
Monthly	11/0	11,11/0	0/0	0/0	0/0	0/0	11/0	9,91/0
Every 6 months	11/6	11,11/26,09	1/0	12,50/0	1/3	25/50	13/9	11,71/28,13
Annually	7/8	7,07/34,78	1/2	12,50/33,33	0/2	0/33,33	8/12	7,21/37,50
Never	60/9	60,61/39,13	5/1	62,50/66,67	3/1	75/16,67	68/11	61,26/34,37
Total	99/23	100/100	8/3	100/100	4/6	100/100	111/32	100/100
$\chi^2= 3,16$ ns $p= 0,92$ / $\chi^2= 3,35$ ns $p= 0,50$								

265 n: number of cases   %: percentage    $\chi^2$ : chi-square   ns: not significant   p: probability

266

267 It was observed that puppy owners do not provide adequate deworming treatment, of

268 which after 3- or 4-weeks puppies begin to shed *T. canis* eggs freely into the soil, where they

269 become infective. Consulted databases did not produce any published work on this parasitosis

270 that considered these epidemiological variables. Both parishes had different patterns of  
 271 behavior in relation to these items, but no statistically significant association was found in either  
 272 parish.

273 There was a statistically significant association with the collection of canine faeces by  
 274 their owners in the parish of Cumanacoa, where 91.30% of the animals defecated in the yard  
 275 and 78.26% of the owners did not collect the dog fecal matter. Thus, these faeces, together  
 276 with the evolutionary forms of parasites in the contaminated fecal material can develop and be  
 277 ready for infection (Table 7).

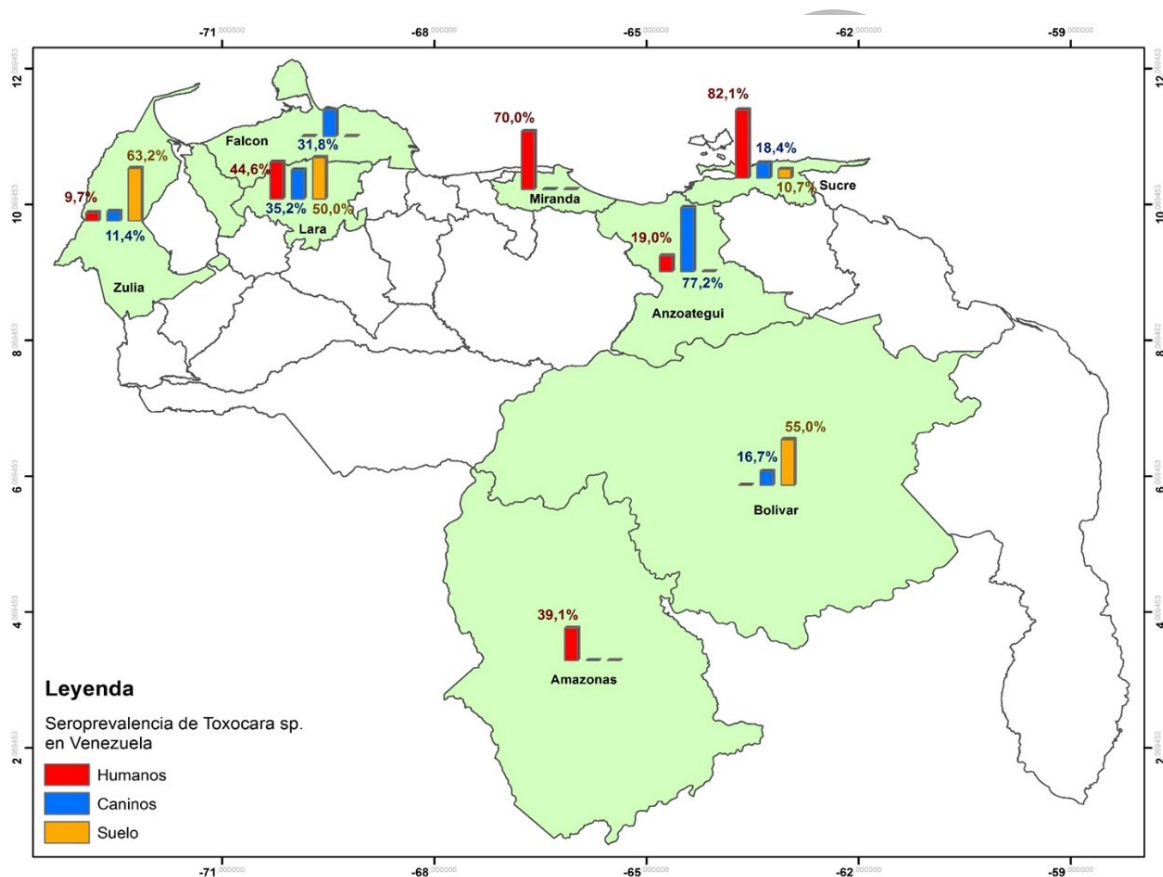
278 **Table 7.** Association between location of dog defecation, collection of dog faeces by dog  
 279 owners, the frequency of dog faeces collection and disposal, and the seroprevalence of anti-  
 280 *Toxocara canis* IgG in inhabitants of San Juan and Cumanacoa (Sucre and Montes  
 281 municipality), Sucre state, Venezuela.

IgG ANTI- <i>Toxocara canis</i> in Inhabitants of San Juan / Cumanacoa								
Variables	Positives		Indeterminate		Negatives		Total	
	n	%	n	%	n	%	n	%
Location of dog defecation								
Courtyard	81/21	81,82/91,30	7/2	87,50/66,67	4/5	100/83,3	92/28	82,88/87,50
Thicket	17/0	17,17/0	0/0	0/0	0/0	0/	17/0	15,32/0
House	1/2	1,01/8,70	1/1	12,50/33,33	0/1	0/16,67	2/4	1,80/12,50
Total	99/23	100/100	8/3	100/100	4/6	100/100	111/32	100/100
$\chi^2= 7,73$ ns $p= 0,10$ / $\chi^2= 1,59$ ns $p= 0,45$								
Collection of faeces								
Yes	51/5	51,52/21,74	6/1	75/33,33	4/5	100/66,6	61/26	54,96/81,26
No	48/18	48,48/78,26	2/2	25/66,67	0/1	0/33,33	50/6	45,05/18,75
Total	99/23	100/100	8/3	100/100	4/6	100/100	111/32	100/100
$\chi^2= 5,05$ ns $p= 0,08$ / $\chi^2= 8,01$ ns $p= 0,02$								
Contact with the site								
Yes	51/20	51,52/86,96	6/2	75/66,67	4/4	100/83,3	61/11	54,96/34,37
No	48/3	48,48/13,04	2/1	25/33,33	0/2	0/16,67	50/21	45,05/65,63
Total	99/23	100/100	8/3	100/100	4/6	100/100	111/32	100/100
$\chi^2= 5,05$ ns $p= 0,08$ / $\chi^2= 8,01$ *s $p= 0,02$								

282 n: number of cases    %: percentage     $\chi^2$ : chi-square    ns: not significant    \*s: significant    p:  
 283 probability

284

285 In summary, we have established prevalence of the *Toxocara* infection in human  
286 subjects, dogs (pets and strays) and in soil samples obtained from the vicinity of the homes  
287 and playgrounds. Thus, we have identified two zoonotic foci of potential *Toxocara* sp.  
288 transmission and maintenance parasite life cycle in Sucre state. An epidemiological map was



289 charted comparing the situation in Sucre state with other states in Venezuela (Fig. 2).

290

291 **Figure 2.** Prevalence of *Toxocara* sp. in Sucre state compared to states in Venezuela  
292 with existing reports of *Toxocara* sp. studies.

293

## 294 DISCUSSION

295 The parish of San Juan, Sucre municipality, Sucre state, Venezuela, is a rural area

296 dedicated to agriculture and animal husbandry (pigs, cattle, and poultry) usage. These activities  
297 are both a source of income and food, as well as an area of regional tourism. The municipality  
298 of Montes carries out similar economic activities, especially agriculture and livestock farming  
299 with a special emphasis on sugar cane, coffee, vegetable crop production and cattle, poultry  
300 and pig rearing.

301 We found high seroprevalence of *T. canis* (Table 1), nonetheless, globally, the  
302 prevalences reported are lower than those of this study: in Colombia, a prevalence of 39% was  
303 reported, in Argentina 37% and in Brazil 28% (Ulloque-Badaracco *et al.*, 2023). However, a  
304 study in Paraguay (78%), but not in Turkey (7.6%) and Italy (31.87%) reports prevalences  
305 similar to those obtained in this study (Rivarola *et al.*, 2009; Akdemir, 2010; Qualizza *et al.*,  
306 2011).

307 In Venezuela, most studies on *T. canis* infection in humans have been carried out in the  
308 central north, west and south of the country. In Amazonas, a seroprevalence of 19.66% was  
309 found (Oliart-Guzmán *et al.*, 2014), in preschool children in Aragua state, Venezuela, were  
310 detected in 29% (65/224) of children. The seroprevalence in the different preschools studied  
311 ranged between 4.2% and 60.6% (Martínez *et al.*, 2015). In Zulia state, García *et al.* (2004)  
312 found a prevalence of 9.72% while a study conducted in Barquisimeto, a seroprevalence of  
313 44.60% was reported (Delgado *et al.*, 2009).

314 Few studies have been conducted on *T. canis* infection in eastern Venezuela. Gómez  
315 (2004), for instance, obtained a 77.14% prevalence in Sucre State, using the ELISA method,  
316 while Fernández *et al.* (2009), using the RIDASCREEN ELISA technique, reported a  
317 prevalence of 19.0% in Anzoátegui State. It is noteworthy that participants evaluated in this  
318 study, from both parishes, have high prevalence rates, a reflection of zoonotic foci present in  
319 these communities.

320 The seroprevalence of anti-*T. canis* IgG according to gender, was found that females were



321 more affected in both San Juan and Cumanacoa parishes (Table 2). Comparable results were  
322 obtained by Espinoza *et al.* (2003) and Gómez (2004), who found a higher seroprevalence in  
323 females with 57.80% and 61.11%, respectively. However, these findings differ from those by  
324 Martín *et al.* (2008), Delgado *et al.* (2009), Akdemir (2010) and Wisniewska *et al.* (2011) who  
325 report that 57.63%, 51.86%, 77.80% and 62.14%, respectively, were in males. Nevertheless,  
326 Roldán *et al.* (2010b) mentioned that the gender of the individual seems not to be an important  
327 predisposing factor for toxocariasis in human populations.

328 The results found in the age group of both municipalities coincide with those reported by  
329 various authors such as Aguiar-Santos *et al.* (2004) who reported a prevalence of 60% in  
330 children under 10 years of age and Gómez (2004) who reported a prevalence of 40.74% in  
331 children aged 2-4 years. Similarly, López *et al.* (2005) reported 67.70% in children aged 1-14  
332 years while Fernández *et al.* (2009) recorded a prevalence of 22.50% in children aged 6 years  
333 and Rivarola *et al.* (2009) found a prevalence of 64.15% in children aged 3 to 9 years. However,  
334 none of these researchers found age-group specific differences in the occurrence of  
335 toxocariasis. García *et al.* (2004) suggested that the reduced risk of infection in older individuals  
336 might be due to improved hygienic habits and spending more time away from home thus  
337 distancing them from sources of infection. It was proposed that this parasitic infection was most  
338 likely to occur at home more than at school or at work, where sanitary and infrastructural  
339 conditions were better.

340 It is important to highlight that in the two parishes; the schools are close to rivers and  
341 surrounded by fields, where children carry out their recreational activities, and where dogs  
342 defecate openly consequently depositing parasite eggs. Additionally, many of these  
343 communities and schools lack piped water and thus draw water from the rivers for consumption  
344 by children and residents. Subsequently, residents of these communities, including school  
345 children, are exposed to contracting this parasitosis both at home and in their schools.

346           These results differ from those obtained by Acero *et al.* (2000), who found that 46.15% of  
347 children never washed their hands before eating and by Gómez (2004) who found that 62.96%  
348 of the *Toxocara* seropositive children did not wash their hands before eating, but 100% of them  
349 declared washing their food before consumption, particularly those to be eaten raw.

350           Most participants from San Juan (63,01%) and Cumanacoa (68,12%), had contact *with T.*  
351 *canis* soil contaminated, had parasitized dogs and did not wash their hands well before eating  
352 or ingesting food. These participants were predisposed to high risk of easily getting infected.  
353 These rates were comparable to results found by Acero *et al.* (2000) who reported that 53.85%  
354 of toxocara-infected children consumed soil, had poor hygiene practices, López *et al.* (2003)  
355 found that 30% of toxocariasis positive children had a history of geophagy whilst Delgado *et al.*  
356 (2009) noted that 96.3% of the infected children had a history of geophagy.

357           In both evaluated parishes, there were large tracts of land, contaminated with canine  
358 faeces, and consequently *T. canis* eggs. At the same time, these fields served as playgrounds  
359 for the children or fruit and vegetable gardens.

360           Concerning the presence of dogs in the homesteads, 67.81% of the seropositive  
361 individuals in the San Juan parish stated that they had dogs as pets, which differed from those  
362 of Cumanacoa parish (33.33%). However, it should be noted that majority of the inhabitants  
363 from both parishes declared coming in contact with other canines in the area due to the large  
364 and uncontrolled population of stray dogs found in both parishes. Gómez (2004) found that  
365 52.26% of seropositive participants had dogs while Martín *et al.* (2008) reported that 74.57%  
366 of those seropositive for IgG anti-*T. canis* antibodies owned dogs. However, in a study  
367 evaluating playgrounds in Turkey, Akdemir (2010) found that only 7.60% of those seropositive  
368 had dogs. We note that having dogs as pets was not significantly associated with testing  
369 positive for the IgG anti-*T. canis* antibodies.

370           Fan *et al.* (2004) argued that toxocariasis is not related to dog breeding, but dog breeding

371 is a condition that in most cases contributed to the occurrence of infection, especially in low  
372 socio-economic societies. Similarly, Alonso *et al.* (2004) pointed out that neither direct contact  
373 with dogs, even puppies, nor the presence of the animals in the home are sufficient conditions  
374 for acquiring *T. canis* infection; other epidemiological circumstances had to coincide to favor  
375 the risk of an infection being acquired from a contaminated environment.

376 Some studies suggest that the likelihood of infection had more to do with individual  
377 susceptibility, and behavioral and hygienic habits rather than with the environmental factors  
378 present in the population. Considering this, we sought to understand if epidemiological  
379 variables such as age of the canines present, deworming and time of deworming of the  
380 individuals who indicated that they owned dogs and the seroprevalence of anti-*T. canis* IgG.

381 The majority of the seropositive individuals in the parish of San Juan stated that their dogs  
382 defecated in the yard (81.82%), 51.52% reported collecting dog faeces and frequenting the  
383 place of disposal, 27.27% disposed of these faeces in the thicket and 15.15% accumulated  
384 them until the waste collection day. This was indicative that these individuals also had frequent  
385 and direct contact with *T. canis* contaminated dog faeces in addition to being surrounded by  
386 animal waste subsequently increasing the possibility of parasite transmission and reinfection.  
387 In the databases consulted, no published studies on this parasitosis were found that took these  
388 epidemiological variables into account.

389 One of the main sources of *T. canis* infection found within the homesteads are gardens  
390 and yards heavily contaminated with dog faeces. Additionally, the ease with which unwormed  
391 dogs access homes greatly contributes to this contamination and high likelihood of infection.  
392 It is estimated that one gram of faeces from an infected animal can harbour about 10.000 *T.*  
393 *canis* eggs, while a female parasite can shed up to 200.000 eggs daily into the environment.  
394 Larval eggs can survive up to 10 years in the environment if conditions are right, thanks to their  
395 high resistance (García *et al.*, 2004).

396 Taking into account the observations by García *et al.* (2004) and Alonso *et al.* (2004),  
397 geophagia, the high dog population, the manner in which dogs are kept without restrictions of  
398 space or specific places to defecate, and the lack of collection of excrement from stray dogs,  
399 the parishes of San Juan and Cumanacoa have become a favorable place for the transmission  
400 of this zoonosis. In these localities, the prevalent conditions expose the inhabitants, especially  
401 children, to the parasite-contaminated excrement.

402 The population sample studied corresponded to individuals from two different socio-  
403 economic and cultural backgrounds, but with the same social deficiencies. This could explain  
404 why no major differences in risk factors association were found between serology-positive and  
405 serology-negative patients.

406 Additionally, a representative sample of canine faeces was also taken and evaluated for  
407 parasitological contamination in these communities. In the parish of San Juan, 93.87% (46 of  
408 49) of the canine faecal samples were parasitized of which the prevalence of *T. canis* was  
409 18.37%. Similarly, in the parish of Cumanacoa, 65.40% (53 of 81) of the samples were  
410 parasitized, of which 9.90% were the *T. canis* species. This is indicative of which shows the  
411 risk of infection with *T. canis* in both communities. These findings are similar to those of  
412 Ramírez-Barrios *et al.* (2004) and Devera *et al.* (2008) that showed a 11.4% and 16.7%  
413 prevalence of canine intestinal parasites in Venezuela, specifically in the city of Maracaibo and  
414 Ciudad Bolívar.

415 In addition to the faecal samples, soil samples were also analyzed. Of the 35 and 28 soil  
416 samples from San Juan and Cumanacoa parishes analyzed, 5.0% and 10.71% were positive  
417 for *Toxocara* sp. Eggs, respectively. We however declare that at the time of soil collection, the  
418 weather condition in the localities was unpredictable, with constant and torrential rains, such  
419 that the parasite eggs in the soil could have been washed by Surface run off into the rivers that  
420 flow through the communities. Despite the weather conditions, the presence of *Toxocara* sp.

421 eggs in the soil of both parishes in Sucre municipality, Sucre State, Venezuela, demonstrates  
422 the high risk of contact and possible accidental ingestion by children in these communities. This  
423 subsequently predisposes them to acquiring the Visceral or Ocular Larva Migrans syndrome,  
424 in addition to advancing high infection rates in dogs. Very heterogeneous result was reported  
425 by Laiño *et al.* (2024), (1.5%) and Antonio *et al.* (2020) (32.5%).

426 Comparing with other studies around Venezuela, from the two zoonotic foci evaluated in  
427 Sucre State demonstrates that this region has one of the highest seroprevalences of  
428 toxocariasis in humans. However, Anzoátegui State reports the highest prevalence of *Toxocara*  
429 *sp.* in canines while Zulia State has the highest rate of soil infected with *Toxocara sp.* eggs  
430 (Cazorla-Perfetti *et al.*, 2007; Tortolero *et al.*, 2008; Devera, 2008; Fernández *et al.*, 2009;  
431 Nieves *et al.*, 2012; Apóstol *et al.*, 2013;). We are of the opinion that the high seroprevalence  
432 is attributed to the rural condition of both parishes evaluated in Sucre State. It is further  
433 compounded by the lack of a Zoonosis department in the local government to manage the large  
434 number of stray dogs in the communities and promote civic education on disease, dynamics of  
435 transmission, consequences of infection.

436 As a zoonosis and geohelminthiasis, this parasitosis represents a public health problem  
437 for both parishes in Sucre State. Its epidemiology depends closely on maintaining the chain of  
438 infected domestic and/or stray dogs and cats, on human contact with soil contaminated with  
439 the infected and on the general habits and attitudes that tend to acquire the infection. It is crucial  
440 that the public health authorities implement a control and health education plan to raise  
441 awareness of the forms of contagion of this parasitosis and the most appropriate way to prevent  
442 it. Primary prevention should focus on veterinarial control of pets, culling of stray dog  
443 populations and improving habits and attitudes that tend to maintain the infection in both  
444 municipalities of Sucre State, Venezuela.

445  
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447  
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473

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479 BIBLIOGRAPHIC REFERENCES

- 480 Acero, M., Muñoz, M., Flores, A., & Nicholis, R. (2000). Seroprevalencia de anticuerpos contra  
481 *Toxocara canis* y factores de riesgo en niños, Ciudad Bolívar, Bogotá. *Biomédica (Bogotá)*, 21,  
482 256-263.
- 483 Aguiar-Santos, A., Andrade, L., & Medeiros, Z. (2004). Human toxocariasis: Frequency of anti  
484 *Toxocara* antibodies in children and adolescents from an outpatient clinic for lymphatic filariasis  
485 in Recife, Northcast, Brasil. *Revista do Instituto de Medicina Tropical de São Paulo*, 46, 81-85.
- 486 Akdemir, C. (2010). Visceral larva migrans among children in Kutahya (Turkey) and an  
487 evaluation of playgrounds for *T. canis* eggs. *The Turkish Journal of Pediatrics*, 53, 158-162.
- 488 Antonio, D.R., Daniel, A.L.V., José, V.R.F., Dario, A.R.I., & Yanitza, B.M.Y. (2020). *Toxocara*  
489 spp. y otros helmintos en muestras de suelo de plazas y parques de Puerto Ordaz, Municipio  
490 Caroní, Estado Bolívar, Venezuela. *Kasmera*, 48, e48231618
- 491 Apóstol, P., Pasceri, P., & Javitt-Jiménez, M. (2013). Detección de huevos de *Toxocara* sp. en  
492 suelos de tres parques públicos de la zona centro de Barquisimeto, estado Lara. *Revista del*  
493 *Colegio de Médicos Veterinarios*, 5, 38-47.
- 494 Alonso, J., López, M., Bojanich, M., & Marull, J. (2004). Infección por *Toxocara canis* en  
495 población adulta sana de un área subtropical de Argentina. *Parasitología latinoamericana*, 59,  
496 61-64.
- 497 Barriga, O. (1988). A critical look at the importance, prevalence and control of Toxocariasis and  
498 the possibilities of immunological control. *Veterinary Parasitology*, 29, 195-234.
- 499 Borge, H., & Ferreira, J. M. (2020). Leucocorias e seus diagnósticos diferenciais: Um relato de  
500 caso. *Revista da Faculdade de Medicina de Teresópolis*, 4, 42-50.
- 501 Botero, D., & Restrepo, M. (1998). *Parasitosis humanas*. Tercera edición. Corporación para  
502 investigaciones biológicas.
- 503 Cabral-Monica, T., Evers, F., de Souza Lima Nino, B., Pinto-Ferreira, F., Breganó, J.W.,

504 Ragassi Urbano, M., Rubinsky-Elefant, G., Freire, R.L., Navarro, I.T., & Mitsuka-Breganó, R.  
505 (2022). Socioeconomic factors associated with infection by *Toxoplasma gondii* and *Toxocara*  
506 *canis* in children. *Transboundary and emerging diseases*, 69, 1589-1595.

507 Cazorla-Perfetti, D.J., Morales Moreno, P., & Acosta Quintero, M.E. (2007). Contaminación de  
508 suelos con huevos de *Toxocara* spp. (Nematoda, Ascaridida) en parques públicos de la ciudad  
509 de Coro, estado Falcón, Venezuela. *Revista Científica de la Facultad de Ciencias Veterinarias*,  
510 17, 117-122.

511 Delgado, R., Díaz, D., Garrido, N., Medina, Z., & Torres, M. (2009). *Presencia de anticuerpos*  
512 *séricos IgG anti-Toxocara canis en niños de 1 a 6 años con y sin síntomas respiratorios que*  
513 *consultan al ambulatorio urbano tipo II "El Jebe". Barquisimeto, estado Lara. Octubre 2008–*  
514 *Marzo 2009.* (Trabajo de pregrado. Departamento de Medicina Preventiva y Social.  
515 Universidad Centro Occidental. Barquisimeto, Venezuela).

516 Delgado, O., & Rodríguez-Morales, A.J. (2009). Aspectos clínicos de la toxocariasis una  
517 enfermedad desatendida en Venezuela y America Latina. *Boletín de Malariología y Salud*  
518 *Ambiental*, 49, 1-33.

519 Devera, R., Blanco, Y., Hernández, H., & Simoes, D. (2008). *Toxocara* spp. y otros helmintos  
520 en plazas y parques de Ciudad Bolívar, estado Bolívar, Venezuela. *The Journal Enfermedades*  
521 *Infecciosas y Microbiología Clínica*, 26, 23-26.

522 Espinoza, Y., Huapaya, P., Huiza, A., Jiménez, S., & Náquira, C. (2003). Toxocariasis humana:  
523 seroprevalencia en población de Lima mediante la técnica de ELISA. *Anales de la Facultad de*  
524 *Medicina*, 64, 228-232.

525 Fan, C.K., Hung, C.C., Du, W.Y., Liao, C.W., & Su, K.E. (2004). Seroepidemiology of *Toxocara*  
526 *canis* infection among mountain aboriginal schoolchildren living in contaminated districts in  
527 eastern Taiwan. *Tropical Medicine & International Health*, 9, 1312-1318.

528 Fernández, L., Pimentel, R., & Poyer, M. (2009). *Seroprevalencia de anticuerpos anti-Toxocara*



529 y alteraciones oculares en escolares de la U.E. Padre Salimero "Fe y Alegría" del municipio  
530 Sotillo del estado Anzoátegui. Trabajo de pregrado. Departamento de Medicina. Universidad  
531 de Oriente. Anzoátegui, Venezuela.

532 García, M., Díaz, O., Estevez, J., Cheng, R., Araujo, M., & Catellano, J. (2004). Prevalencia de  
533 infección por *Toxocara* en pre-escolares de una comunidad educativa de El Moján, estado  
534 Zulia, Venezuela. Resultados preliminares. *Investigación Clínica*, 45, 347-354.

535 Gómez, M. (2004). *Toxocariasis humana y canina en la localidad del barrio Nueva Toledo del*  
536 *El Peñón, parroquia Valentín Valiente, municipio Sucre, estado Sucre*. Trabajo de pregrado.  
537 Departamento de Bioanálisis. Universidad de Oriente. Sucre.

538 Gordis, L. (2004). *Epidemiology*. 3<sup>rd</sup> ed. Elsevier Saunders.

539 Heymann, D.L. (eds) (2004). *Control of communicable diseases manual: An official report of*  
540 *the american public health association*. 18<sup>th</sup> (edn). American Public Health Association,  
541 Washington DC, USA. 700.

542 Holland, C., O'Lorcain, P., Taylor, M., & Kelly, A. (1995). Seroepidemiology of toxocariasis in  
543 school children. *Parasitology*, 110, 534-542.

544 Incani, R., Aguilar C., & Davila Pacheco M. (1996). *Parasitología*. Tatum.

545 Laiño, M., Domínguez, L., Gramajo, L., Siccardi, F., Herculini, C., Beltrán, F., & Vidal, J. (2024).  
546 Contaminación fecal canina, helmintos y protozoarios zoonóticos en Buenos Aires: Asociación  
547 con variables ambientales. *Revista Argentina de Salud Pública*, 16, 67-73.

548 Leventhal, R., & Cheadle, R. (1992). *Parasitología Médica*. 3<sup>ra</sup>. Ed., Ed. Interamericana, Mc  
549 Graw-Hill.

550 López, M., Alonso, J., Bojanich, M., Chamorro, M., & Falivene, G. (2003). Aspectos  
551 inmunológicos de la infección infantil por *Toxocara canis* en el area del Gran Resistencia.  
552 *Instituto de Medicina Regional-UNNE. Resumen: M-017*.

553 López, M., Fernández, G., Bojanich, M., & Alonso, J. (2005). Infección por *Toxocara canis* en

554 una población infantil vulnerable de la ciudad de Corrientes, Argentina. *Comunicaciones*  
555 *Científicas y Tecnológicas*, 58, 425-431.

556 Ma, G., Rostami, A., Wang, T., Hofmann, A., Hotez, P.J., & Gasser, R.B. (2020). Global and  
557 regional seroprevalence estimates for human toxocariasis: A call for action. *Advances in*  
558 *Parasitology*, 109, 275–290.

559 Martín, U., Machuca, P., Demonte, M., & Contini, L. (2008). Estudio en niños con diagnóstico  
560 presuntivo de toxocariasis en Santa Fe, Argentina. *Medicina (Buenos Aires)*, 68, 353-357.

561 Martínez, C. (2003). *Estadística y muestreo*. Décimo primera edición. Ecoe ediciones.

562 Martínez, M., García, H., Figuera, L., González, V., Lamas, F., López, K., Mijares, V., Corrales,  
563 Y., Lares, M., & Ferrer, E. (2015). Seroprevalence and risk factors of toxocariasis in preschool  
564 children in Aragua state, Venezuela. *Transactions of the Royal Society of Tropical Medicine*  
565 *and Hygiene*, 109, 579-588.

566 Macpherson, C.N. (2013). The epidemiology and public health importance of toxocariasis: A  
567 zoonosis of global importance. *International journal for parasitology*, 43, 999–1008.

568 Nieves, M., Guilarte, Del Valle, Gómez, E., Díaz, A., Toledo, J., & Díaz, M. (2012). Zoonosis  
569 canina en comunidades de la parroquia San Juan, municipio Sucre, estado Sucre, Venezuela.  
570 *IX Congreso Científico Universidad de Oriente*. Cumaná, Venezuela.

571 Oliart-Guzmán, H., Delfino, B.M., Martins, A.C., Mantovani, S.A., Brana, A.M., Pereira, T.M.,  
572 Branco, F.L.C.C., Ramalho, A.A., Campos, R.G., Fontoura, P.S., de Araujo, T.S., de Oliveira,  
573 C.S.M., Muniz, P.T., Rubinsky-Elefant, G., Codeço, C.T., & da Silva-Nunes, M. (2014).  
574 Epidemiology and control of child toxocariasis in the Western Brazilian Amazon—a population-  
575 based study. *The American journal of tropical medicine and hygiene*, 90, 670–681.

576 OPAS - Organización Panamericana de la Salud. (1993). *Normas éticas internacionales para*  
577 *la investigación biomédica con sujetos humanos*. Publicación científica.

578 Camacho, G.P., Roca, D.L., Pérez, B.P., & Casanova, R.T. (2006). Aplicación de la técnica de

579 sedimentación espontánea en tubo en el diagnóstico de parásitos intestinales. *Revista*  
580 *Mexicana de Patología Clínica y Medicina de Laboratorio*, 53, 114-118.

581 Pérez, L. (2006). Corografía municipal del estado Sucre (para la guía turísticas). Biblioteca.

582 Qualizza, R., Incorvaia, C., Grande, R., Makri, E., & Allegra, L. (2011). Seroprevalence of IgG  
583 anti-*Toxocara* species antibodies in a population of patients with suspected allergy.  
584 *International Journal of General Medicine*, 4, 783-787.

585 Quinn, R., Smith, H., Bruce, R., & Gierdawood., R. (1980). On the incidence of *Toxocara* and  
586 *Toxocaris* spp ova in the environment. A comparison of flotation procedures for recovering  
587 *Toxocara* spp ova soil. *Epidemiology & Infection*, 84, 83-89.

588 Ramírez-Barrios, R.A., Barboza-Mena, G., Muñoz, J., Angulo-Cubillán, F., Hernández, E.,  
589 González, F., & Escalona, F. (2004). Prevalence of intestinal parasites in dogs under veterinary  
590 care in Maracaibo, Venezuela. *Veterinary parasitology*, 121, 11-20.

591 Rivarola, M., Vuyk, I., Riveros, M., & Canese, M. (2009). *Toxocara canis* en población  
592 pediátrica rural. *Pediatría (Asunción)*, 36, 123-126.

593 Roldán, W.H., Cavero, Y.A., Espinoza, Y.A., Jiménez, S., & Gutiérrez, C.A. (2010a).  
594 Toxocaríase humana: inquérito soroepidemiológico na cidade amazônica de Yurimaguas,  
595 Peru. *Revista do Instituto de Medicina Tropical de São Paulo*, 52, 37-42.

596 Roldán, W., Espinoza, Y., Huapaya, P., & Jiménez, S. (2010b). Diagnóstico de la toxocarosis  
597 humana. *Revista peruana de medicina experimental y salud publica*, 27, 613-620.

598 Rostami, A., Riahi, S.M., Holland, C.V., Taghipour, A., Khalili-Fomeshi, M., Fakhri, Y., Omrani,  
599 V.F., Hotez, P.J., & Gasser, R.B. (2019). Seroprevalence estimates for toxocariasis in people  
600 worldwide: A systematic review and meta-analysis. *PLoS neglected tropical diseases*, 13,  
601 e0007809.

602 Taylor, M. A., Coop, R. L., & Wall, R. L. (2017). *Parasitologia Veterinária*. Guanabara Koogan.

603 Tortolero, L., Cazorla, D., Morales, P., & Acosta, M. (2008). Prevalencia de enteroparásitos en

604 perros domiciliados de la ciudad de La Vela, estado Falcón, Venezuela. *Revista científica,*  
605 *FCV-LUZ, 18, 312-319.*

606 Ulloque-Badaracco, J. R., Hernandez-Bustamante, E. A., Alarcón-Braga, E. A., Huayta-Cortez,  
607 M., Carballo-Tello, X.L., Seminario-Amez, R.A., Rodríguez-Torres, A., Casas-Patiño, D.,  
608 Herrera-Añazco, P., & Benites-Zapata, V.A. (2023). Seroprevalence of human toxocariasis in  
609 Latin America and the Caribbean: a systematic review and meta-analysis. *Frontiers in Public*  
610 *Health, 11, 1181230.*

611 Vázquez, O., Martínez, B., Tay, J., Ruiz, A., & Pérez, A. (1997). Verduras de consumo como  
612 probable fuente de infección de *Toxocara* sp. para el hombre. *Boletín chileno de parasitología,*  
613 *52, 47-50.*

614 Wisniewska, M., Wozniakowska, T., Sobolewska, J., Markiewicz, A., & Wieczorek, M. (2011).  
615 Analysis of the course and treatment of toxocariasis in children a long-term observation.  
616 *Parasitology research, 80, 531-536.*

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