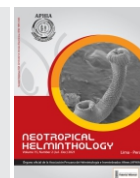




## Neotropical Helminthology



ORIGINAL ARTICLE / ARTÍCULO ORIGINAL

### HELMINTH COMMUNITY OF BACKYARD CHICKENS (*GALLUS GALLUS DOMESTICUS* LINNAEUS, 1758) IN SEROPÉDICA, RIO DE JANEIRO, BRAZIL

### COMUNIDADE DE HELMINTOS DE FRANGOS DE QUINTAL (*GALLUS GALLUS DOMESTICUS* LINNAEUS, 1758) EM SEROPÉDICA, RIO DE JANEIRO, BRASIL

### COMUNIDAD DE HELMINTOS DE POLLOS CASEROS (*GALLUS GALLUS DOMESTICUS* LINNAEUS, 1758) EN SEROPÉDICA, RIO DE JANEIRO, BRASIL

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## ABSTRACT

The domestic chicken, *Gallus gallus domesticus* Linnaeus, 1758, is an important component of the agribusiness segment, and Brazil is one of the world's largest broiler producers and exporters. The present study aimed to characterize the composition and structure of the helminth community of backyard chickens, *G. g. domesticus*, in Seropédica, Rio de Janeiro, Brazil. Fifty-five adult chickens were studied. The overall helminth species richness was 12. The nematodes *Capillaria* sp. and *Heterakis gallinarum* (Schränk, 1788), recovered from small intestine and cecum, respectively, presented the highest prevalence and mean abundance and were considered central species. In addition, these species presented the highest values of frequency of dominance and Berger. The pairs of helminth species *Amoebotaenia cuneata* (von Linstow, 1872) – *Raillietina tetragona* (Molin, 1958); *Davainea proglottina* (Davaine, 1860) – *A. cuneata*; and *H. gallinarum* – *Capillaria* sp. showed significant positive correlation between their abundance and prevalence. *Gongylonema ingluvicola* Ransom, 1904 and *H. gallinarum* showed significant correlation between host sex and helminth abundance, while there was no correlation between host sex and helminths prevalence. The knowledge of helminth community structure in free-range chickens is important to adopt better measures for control and prevention of helminth infections.

**Keywords:** birds – Cestoda – Nematoda – parasite ecology – prevalence – parasite richness

## RESUMO

As galinhas domésticas *Gallus gallus domesticus* Linnaeus, 1758, são um importante componente do segmento do agronegócio, e o Brasil é um dos maiores produtores e exportadores de frangos de corte. O presente estudo teve como objetivo caracterizar a estrutura da comunidade componente de helmintos de galinhas de fundo de quintal, *G. g. domesticus*, em Seropédica, Rio de Janeiro, Brasil. Foram estudados 55 frangos adultos. A riqueza total de helmintos foi de 12 espécies. Os nematóides *Capillaria* sp. e *Heterakis gallinarum* (Schrank, 1788), encontrados no intestino delgado e ceco, respectivamente, foram as espécies mais prevalentes e com maior abundância média e foram consideradas espécies centrais. Além disso, essas espécies apresentaram alta frequência de dominância e dominância relativa média. Os pares de espécies de helmintos *Amoebotaenia cuneata* (von Linstow, 1872) - *Raillietina tetragona* (Molin, 1958); *Davainea proglottina* (Davaine, 1860) - *A. cuneata* e *H. gallinarum* - *Capillaria* sp. mostraram correlação positiva significativa entre abundância média e prevalência parasitária. *Gongylonema ingluvicola* Ransom, 1904 and *H. gallinarum* mostraram correlação significativa entre sexo do hospedeiro e abundância de helminto, enquanto que não houve correlação no sexo do hospedeiro e a prevalência destes. O conhecimento da estrutura da comunidade de helmintos em frangos criados no quintal é importante para a adoção de medidas de controle e prevenção de helmintos.

**Palavras-chaves:** aves – Cestoda – Nematoda – ecologia parasitária – prevalência – riqueza parasitária

## RESUMEN

Las gallinas domésticas, *Gallus gallus domesticus* Linnaeus, 1758, son un componente importante del segmento de la agroindustria y Brasil es uno de los mayores productores y exportadores de pollos. El presente estudio tuvo como objetivo caracterizar la estructura comunitaria de helmintos de pollos criados en corrales, *G. g. domesticus*, en Seropédica, Rio de Janeiro, Brasil. Se estudiaron 55 aves adultas. La riqueza total de helmintos fue de 12 especies. *Capillaria* sp. y *Heterakis gallinarum* (Schrank, 1788), que se encontraron en el intestino delgado y ciego intestinal, respectivamente, fueron las especies más prevalentes con mayor abundancia promedio y fueron consideradas especies centrales. Además, estas especies mostraron los valores mayores de frecuencia de dominancia y de dominancia relativa media. Los pares de especies de helmintos, *Amoebotaenia cuneata* (von Linstow, 1872) - *Raillietina tetragona* (Molin, 1958); *Davainea proglottina* (Davaine, 1860) - *A. cuneata* y *H. gallinarum* - *Capillaria* sp. mostraron una correlación positiva significativa entre la abundancia media y la prevalencia de parásitos. *Gongylonema ingluvicola* Ransom, 1904 y *H. gallinarum* mostraron una correlación significativa entre el sexo del huésped y la abundancia de helmintos, mientras que no hubo correlación entre el sexo del huésped y la prevalencia de helmintos. El conocimiento de la estructura de la comunidad de helmintos en los pollos criados en corrales es importante para la adopción de medidas de prevención y control de helmintos.

**Palabras clave:** aves – Cestoda – Nematoda – ecología parasitaria – prevalência – riqueza parasitaria

## INTRODUCTION

The domestic chicken, *Gallus gallus domesticus* Linnaeus, 1758, is an important component of the agribusiness segment, and Brazil is one of the world's largest broiler producers and exporters

(ABPA, 2020). There is high demand for chicken meat and eggs due to high nutritional value and affordable price (Geraldo *et al.*, 2020). In the past decade, the market of organic, healthy, and sustainable products and contingent of consumers concerned about animal welfare have both increased (Miao *et al.*, 2005; Alsaffar, 2016).

Chickens under free-range production typically show signs of calmness and comfort (Bogdanov, 1997). However, chickens reared freely are more exposed to helminths than those kept in confinement (Cardozo & Yamamura, 2004; Lozano *et al.*, 2019), since they have direct contact with soil, and free access to different areas and food items (Gomes *et al.*, 2009). A high burden of gastrointestinal helminths can decrease productivity by increasing mortality, causing economic losses to breeders (Ruff, 1999).

In Brazil, many studies have reported the occurrence of helminths in *G. g. domesticus* in different regions and breeding systems (Grisi & Crvalho, 1974; Quadros *et al.*, 2015; Silva *et al.*, 2016; Silva *et al.*, 2018; Brandão-Simões *et al.*, 2020). However, little is known about the helminth community structure of *G. g. domesticus* in this condition (Silva *et al.*, 2016, 2018). Studies of parasites through an ecological approach, to detect possible patterns in the organization and composition of parasite populations, can be relevant to understand the distribution of the pathogen species in different breeding systems. Moreover, it can assist helminthic control strategies and contribute to develop biosafety programs. Here, we characterize the composition and structure of the helminth component community of backyard chickens in Seropédica, Rio de Janeiro, Brazil.

## MATERIALES Y MÉTODOS

### *Host and parasitological procedures*

Fifty-five sexually mature *G. g. domesticus* were purchased dead during 1999-2000. The study was carried out in Seropédica (22° 44' 29"S, 43° 42' 19"W), state of Rio de Janeiro, Brazil. The chickens were subjected to procedures in accordance with standard international parasitological guidelines (Yazwinski *et al.*, 2003). The gastrointestinal tract, eyeball, trachea, lungs, kidneys, and bursa of Fabricius were examined. The helminths were washed in saline solution (0.85% NaCl) and fixed in AFA (2% acetic acid,

3% formaldehyde and 95% ethanol). For microscopic studies, nematodes were cleared in lactophenol and cestodes were stained with chlorhydric carmine. The helminth parasites were identified according to Yamaguti (1961), Khalil *et al.* (1994), Vicente *et al.* (1995) and Shmidt (1986), along with specific papers.

### *Data analysis*

To analyze the helminth community structure, we considered prevalence, mean intensity and mean abundance of each species using the procedures described by Bush *et al.* (1997). Statistical analyses were performed only for the parasite species with prevalence higher than 10%.

The frequency of dominance and the Berger-Parker index of each parasite species were calculated according to Rohde *et al.* (1995) and Magurran (2004).

Species richness, helminth richness at the infracommunity level and Brillouin's diversity index (log10) were calculated. The parasite infracommunity species were classified as: a) central species (present in more than one-third of the hosts); b) secondary species (present in one- to two-thirds of the hosts); and c) satellite species (present in less than a third of the hosts) (Bush & Holmes, 1986).

Mann-Whitney *U* test values and chi-square analysis were used for comparison of parasite abundance and prevalence, respectively, and between host sex (Zar, 1999). These analyses were performed following the recommendations of Wilkinson (1990) using the SYSTAT™ statistical software.

The possible relationships between prevalence and abundance between pairs of concurrent species were determined using the chi-square test and Spearman rank correlation coefficient, respectively (Ludwig & Reynolds, 1988). Statistical significance was considered at  $P < 0.05$ . All ecological terminology was used according to Bush *et al.* (1997).

### *Ethic aspects*

All national and institutional guidelines for use of animals were followed. All chickens were purchased died.

## RESULTS

The overall helminth richness was 12 species. The helminth richness at the infracommunity level ranged of 1 to 10, with a mean of  $5.2 \pm 2.2$ . Fifty-three hosts (96%) presented two or more helminth species.

A total of 10,708 specimens were collected, with mean abundance of  $194.7 \pm 283.9$ . Nematodes were the most abundant, with 7 species representing 60.7% of all parasite specimens collected. All chickens examined were parasitized by at least one species of nematode and 85.7% (47) were parasitized by cestodes. Trematodes were not found. The nematode *Capillaria* Zeder, 1800 was not identified to the species level as cestode *Raillietina* Fuhrmann, 1920 due to the low quality of the morphological characteristics, such as scolex and mature proglottids, of the specimens collected.

The nematodes *Capillaria* sp. and *Heterakis gallinarum* (Schrank, 1788), recovered from the small intestine and cecum, respectively, presented the highest values of prevalence and mean abundance and were considered central species (Table 1). In addition, these species presented the highest values of frequency of dominance and Berger-Parker index (Table 2); while the cestode *Raillietina* sp. presented the highest value of mean intensity and was considered a secondary species along with the cestode *Amoebotaenia cuneata* (von Linstow, 1872) and the nematodes *Gongylonema ingluvicola* Ransom, 1904 and *Oxyuris mansoni*. (Cobbold, 1879) (Table 2). Secondary species were found in the small intestine, crop and eye. *Tetrameres confuse* Travassos, 1917 showed the lowest values of prevalence, mean intensity and abundance (Table 1). The Brillouin index for helminth infracommunities had mean value of  $0.43 \pm 0.19$  and maximum diversity of 0.78.

**Table 1.** Prevalence (P), mean intensity (MI), and mean abundance (MA) followed by standard error, community status (CS) and site of infection (SI) of helminth parasites of *Gallus gallus. domesticus* from Seropédica, Rio de Janeiro, Brazil.

Helminth species	P (%)	MI	MA	CS*	SI
CESTODA					
<i>Amoebotaenia cuneata</i>	41.8	45.4 $\pm$ 99.9	19.0 $\pm$ 67.6	S	Small Intestine
<i>Davainea proglottina</i>	30.9	37.2 $\pm$ 53.0	11.5 $\pm$ 33.7	Sa	Small Intestine
<i>Raillietina tetragona</i>	32.3	16.5 $\pm$ 29.2	5.4 $\pm$ 18.2	Sa	Small Intestine
<i>Raillietina echinobothrida</i>	27.3	17.5 $\pm$ 22.4	4.8 $\pm$ 13.9	Sa	Small Intestine
<i>Raillietina</i> sp.	43.6	81.9 $\pm$ 253.3	35.7 $\pm$ 170.3	S	Small Intestine
NEMATODA					
<i>Ascaridia galli</i>	21.8	37.6 $\pm$ 65.7	8.2 $\pm$ 33.6	Sa	Small Intestine
<i>Capillaria</i> sp.	90.9	46.0 $\pm$ 83.7	41.8 $\pm$ 80.9	C	Small Intestine
<i>Cheilosporura hamulosa</i>	25.4	8.9 $\pm$ 13.2	2.2 $\pm$ 7.6	Sa	Ventricle
<i>Gongylonema ingluvicola</i>	49.1	7.0 $\pm$ 8.2	3.4 $\pm$ 6.7	S	Crop
<i>Heterakis gallinarum</i>	72.7	63.1 $\pm$ 122.8	45.9 $\pm$ 108.1	C	Cecum
<i>Oxyuris mansoni</i>	56.4	27.7 $\pm$ 36.2	15.6 $\pm$ 30.4	S	Eye
<i>Tetrameres confuse</i>	23.6	4.1 $\pm$ 4.4	1.0 $\pm$ 2.7	Sa	Proventriculum

\* Central species (C). Secondary species (S) and Satellite species (Sa).

**Table 2.** Frequency of dominance and Berger Parker Index values (BPI) of the helminth species from *Gallus gallus domesticus* in the municipality of Seropédica, Rio de Janeiro, Brazil.

Helminth species	Frequency of dominance	BPI
CESTODA		
<i>Amoebotaenia cuneata</i>	5	0.08±0.16
<i>Davainea proglottina</i>	2	0.05±0.12
<i>Raillietina tetragona</i>	2	0.04±0.10
<i>Raillietina echinobothrida</i>	3	0.04±0.14
<i>Raillietina</i> sp.	7	0.12±0.24
NEMATODA		
<i>Ascaridia galli</i>	1	0.02±0.09
<i>Capillaria</i> sp.	13	0.25±0.22
<i>Cheilospirura hamulosa</i>	0	0.01±0.03
<i>Gongylonema ingluvicola</i>	0	0.03±0.06
<i>Heterakis gallinarum</i>	15	0.23±0.24
<i>Oxyspirura mansoni</i>	7	0.12±0.21
<i>Tetrameres confusa</i>	0	0.01±0.03

**Table 3.** Mann-Whitney *U* test values (*Z*, value of the normal *U* test approximation) and Chi square ( $\chi^2$ ) to evaluate the relationship between the sex of *Gallus domesticus* and the abundance and prevalence of the components of its parasitic community (*P*= level of significance).

Helminth species	<i>Z</i>	<i>P</i>	$\chi^2$	<i>P</i>
CESTODA				
<i>Amoebotaenia cuneata</i>	-0.18	0.86	0.15	0.70
<i>Davainea proglottina</i>	-0.72	0.47	0.02	0.89
<i>Raillietina tetragona</i>	-0.37	0.71	0.02	0.89
<i>Raillietina echinobothrida</i>	-1.53	0.13	0.23	0.63
<i>Raillietina</i> sp.	-0.47	0.64	0.09	0.76
NEMATODA				
<i>Ascaridia galli</i>	-0.26	0.79	0.22	0.64
<i>Capillaria</i> sp.	-1.85	0.06	0.01	0.94
<i>Cheilospirura hamulosa</i>	-0.71	0.48	0.06	0.81
<i>Gongylonema ingluvicola</i>	-2.11*	0.04	0.96	0.33
<i>Heterakis gallinarum</i>	-2.01*	0.04	0.02	0.89
<i>Oxyspirura mansoni</i>	-1.41	0.16	0.09	0.76
<i>Tetrameres confusa</i>	-0.85	0.40	0.12	0.73

\*Significant Value

*Gongylonema ingluvicula* and *H. gallinarum* showed significant correlation between host sex and helminth abundance, while there was no correlation between host sex and helminths prevalence (Table 3).

The following pairs of concurrent helminths showed significant positive correlation between their abundance and prevalence, respectively: *Amoebotaenia cuneata* – *Raillietina tetragona* (Molin, 1858) ( $r_s = 0.37$ ;  $P = 0.006$ ;  $t^2 = 8.59$ ;  $P = 0.004$ ), *Davainea proglottina* (Davaine, 1860) – *A. cuneata* ( $r_s = 0.46$ ;  $P < 0.001$ ;  $t^2 = 3.53$ ;  $P = 0.049$ ); and *H. gallinarum* – *Capillaria* sp. ( $r_s = 0.39$ ;  $P = 0.003$ ;  $t^2 = 5.6$ ;  $P = 0.024$ ). The pair *R. tetragona* – *Ascaridia galli* showed significant correlation only between abundance ( $r_s = 0.37$ ;  $P = 0.005$ ;  $t^2 = 2.22$ ;  $P = 0.136$ ).

## DISCUSSION

Several studies of the characteristics of the helminth communities of *G. g. domesticus* have been performed worldwide (Schou et al., 2007; Idika et al., 2016; Slimane, 2016; Berhe et al., 2019; Sarba et al., 2019, among others). In Brazil, most studies have focused only on the description of the helminth fauna (Costa & Freitas, 1959; Grisi & Carvalho, 1974; Gomes et al., 2009; Siqueira & Marques, 2016). More recently, knowledge about the helminth community structure of chickens raised in different production systems has increased, mainly in the state of São Paulo (Silva et al., 2018; Silva et al., 2016). In contrast, there are few reports of the helminth community structure of *G. g. domesticus* in the state of Rio de Janeiro (Grisi & Carvalho, 1974; Gomes et al., 2009). The composition and structure of the helminth community of a specific host population correspond to a pool of available parasite species in a specific locality. Moreover, the structure and composition of a helminth community can vary due to biotic and abiotic factors, such as environmental changes, parasite control, host behavior and age, among others (Poulin, 2007; Santoro et al., 2012; Simões et al., 2016; Cardoso et al., 2019).

Grisi & Cravalho (1974) reported 13 species of helminths in *G. g. domesticus* in Seropédica,

similar to the present study, in which we found 12 species in the same locale. However, some species differed between the studies. The species *Syngamus trachea* (Montagu, 1811), *Heterakis brevispiculum* Gendré, 1911, *Capillaria collaris* (Linstow, 1873) and *C. obsignata* Madsen, 1945 were not found in our study, although we found nematodes belonging to the genus *Capillaria*. Unfortunately, the specific identification was not possible. Moreover, we collected two species of cestodes (*Davainea proglottina* and *Amoebotaenia cuneata*) that were not reported by Grisi & Carvalho (1974). Neither study found trematodes. In contrast, Silva et al. (2016, 2018) reported trematodes in different regions of São Paulo state. These differences have also been observed in chickens from African and Asian countries (Berhe et al., 2019; Chege et al., 2015; Junaidu et al., 2014; Schou et al., 2007). Additionally, nematodes and cestodes have been reported with more frequency than trematodes in the component helminth community in chickens by many researchers around the world (Sarba et al., 2019; Idika et al., 2016; Slimane, 2016; Hussen et al., 2012). Variation in environmental conditions can favor or disfavor the presence of intermediate hosts responsible for cestode and trematode transmission and can also influence the occurrence of these species in the helminth community of the definitive host.

In the present study, all chickens were infected with the nematodes *H. gallinarum* and *Capillaria* sp., and both were the more prevalent (central species), with high frequency of dominance and mean values of Berger-Parker index in the helminth community. Similar findings were also observed by Gomes et al. (2009), Siqueira & Marques (2016) and Silva et al. (2018) in chickens reared in extensive, semi-intensive and/or intensive systems. Parasites with a direct life cycle have a higher probability of infecting definitive hosts since there is no intermediate host. In addition, the high fertility of these female nematodes increases the likelihood of host infection due to more eggs eliminated in the environment. The chickens in the present study were raised free outdoors and allowed to find food by foraging in the surface soil layer. Thus, these factors can partly explain the high prevalence of nematodes in the helminth community.

The influence of host sex on the helminth abundance has been observed in different helminth community structures (Simões *et al.*, 2014; Wendt *et al.*, 2018). The influence of female chickens was observed regarding two nematodes species (*H. gallinarum* and *Capillaria* sp.). However, this result must be viewed with caution, since the proportion of female chickens used in the study was much greater than of males (10: 1). In addition, many studies of the helminth community of *G. g. domesticus* have not reported significant differences in helminth infection rate between male and female animals (Poulsen *et al.*, 2000; Abdelqader *et al.*, 2008; Ebrahimi *et al.*, 2014).

The positive correlation of abundance and prevalence between pairs of cestodes and nematodes may be related to the similarities of their life cycle. The cestodes *R. tetragona*, *A. cuneata* and *D. proglottina* need an intermediate host, such as an ant, beetle or fly; earthworm; and slug respectively (Acha & Szyfres, 2003; Taylor *et al.*, 2017). One of the food items collected by the chicken was small animals found in the soil. Similarly, this can explain the positive correlation with nematodes, which have a direct life cycle and oral infection.

Free-range chickens are exposed to more parasites than those reared in intensive systems (Ruff, 1999; Lozano *et al.*, 2019) due to the direct contact with soil and intermediate hosts. Tapeworms are frequently encountered in the chickens in these production systems and most species cause low pathogenicity (Ruff, 1999). However, *R. tetragona* and *D. proglottina* are more pathogenic, reducing the performance of parasitized animals (Ruff, 1999; Nnadi & George, 2010). Moreover, parasitism by *Capillaria* spp. can also cause production losses (Ruff, 1999). Seasonality and other collection areas should be considered in future studies to measure the heterogeneity and stability of the parasite helminth community.

Knowledge of the helminth community structure in backyard chickens can be relevant to allow breeders to adopt better measures for control and prevention of helminths. Moreover, other variables like climate conditions, hygiene, age, and animal density should also be considered since they may cause influence in the parasitism. In addition, this study can also serve as a baseline for future

parasitological studies on other bird species.

## BIBLIOGRAPHIC REFERENCES

- Abdelqader, A, Gauly, M, Wollny, CBA & Abo-Shehada, MM. 2008. *Prevalence and burden of gastrointestinal helminthes among local chickens, in northern Jordan*. Preventive Veterinary Medicine, vol. 85, pp.17-22.
- ABPA. Associação Brasileira de Proteína Animal. 2020. “*Relatório Anual, 2020*” <https://abpa-br.org/relatorios>.
- Acha, B & Szyfres, P. 2003. *Zoonoses and communicable diseases common to man and animals V. 3: Parasitoses*. 3. Ed. PAHO Publication centers, Washington, p. 401.
- Alsaffar, AA. 2016. *Sustainable diets: The interaction between food industry, nutrition, health, and the environment*. Food Science and Technology International, vol. 22, pp. 102-111.
- Brandão-Simões, M, Melo, AL & Moreira, NIB. 2020. Occurrence of *Heterakis gallinarum* Schrank, 1788) (Nematoda: Heterakidae) in *Gallus gallus domesticus* Linnaeus, 1758 in Vitória, Espírito Santo, Brazil. Neotropical Helminthology, vol. 114, pp. 199-203.
- Berhe, M, Mekibib, B, Bsrat, A. & Atsbaha, G. 2019. *Gastrointestinal Helminth Parasites of Chicken under Different Management System in Mekelle Town, Tigray Region, Ethiopia*. Journal of Veterinary Medicine, vol. 2019, pp. 1-7.
- Bogdanov, IA. 1997. *Seasonal effects on free-range egg production*. World Poultry-Misset, vol. 13, pp. 47-49.
- Bush, AO & Holmes, JC. 1986. *Intestinal helminths of lesser scaup ducks: an interactive community*. Canadian Journal of Zoology, vol. 64, pp. 142-152.
- Bush, AO, Laferty, K D, Lotz, JM & Shostak, AW. 1997. *Parasitology meets ecology on its own terms: Margolis et al. Revisited*. Journal of Parasitology, vol. 83, pp. 575-583.

- Cardoso, TS, Macabu, CE, Simões, RO, Maldonado-Júnior, A, Luque, JL & Gentile, R. 2019. *Helminth Community structure of two sigmodontinae rodents in Serra dos Órgãos National Park, State of Rio de Janeiro, Brazil*. *Oecologia Australis*, vol. 23, pp. 301-314.
- Cardozo, SP & Yamamura, MH. 2004. *Parasitas em produção de frangos no sistema de criação tipo colonial/caipira no Brasil*. *Semina: Ciências Agrárias*, vol. 25, pp. 63-74.
- Chege, HW, Kemboi, DC, Bebora, LC, Maingi, N, Mbuthia, PG, Nyaga, PN, Njagi, LW & Githinji, J. 2015. *Studies on seasonal prevalence of ecto- and endo-parasites in indigenous chicken of Mbeere Subcounty, Kenya*. *Livestock Research for Rural Development*, vol. 27, pp. 113.
- Costa, HMA & Freitas, M. 1959. *Novos achados helmintológicos em animais domésticos em Minas Gerais*. *Arquivos da Escola de Veterinária da Universidade Federal de Minas Gerais*, vol. 12, pp. 293-297.
- Ebrahimi, M, Asadpour, M, Verdi, Mk & Borji, H. 2014. *Prevalence and distribution of gastrointestinal helminths in free range chickens in Mashhad, northeast of Iran*. *Science Parasitology*, vol. 15, pp.38-42.
- Geraldo, A, Valentim, JK, Zanella, J, Mendes, JP, Silva, AF, Garcia, RG, Eberhart, BS & Pantoja, JC. 2020. Profile of caipira broiler meat producers and consumers in the Alto San Francisco region-MG. *Revista online de extensão e cultura Realização*, vol. 7, pp.81-93.
- Gomes, FF, Machado, HHD, Lemos, LS, Almeida, LG & Daher, RF. 2009. *Principais parasitos intestinais diagnosticados em galinhas domésticas criadas em regime extensivo na municipalidade de Campos dos Goytacases, RJ*. *Ciência Animal Brasileira*, vol. 10, pp. 818-822.
- Grisi, L & Carvalho, LP. 1974. *Prevalência de helmintos parasitos de Galus galus domesticus L., no estado do Rio de Janeiro*. *Revista Brasileira de Biologia*, vol.34, pp. 115-118.
- Hussen, H, Chaka, H, Deneke, Y & Bitew, M. 2012. *Gastrointestinal helminths are highly prevalent in scavenging chickens of selected districts of Eastern Shewa zone., The Ethiopian Journal of Biological Sciences*, vol. 12, pp. 284-289.
- Idika, IK, Obi, C, Ezeh, IO, Iheagwam, CN, Njoku, IN & Nwosu, CO. 2016. *Gastrointestinal helminth parasites of local chickens from selected communities in Nsukka region of southeastern Nigeria*. *Journal of Parasitic Diseases*, vol. 40, pp. 1376–1380.
- Junaidu, HI, Luka, S A & Mijinyawa, A. 2014. *Prevalence of gastrointestinal helminth parasites of the domestic fowl (Gallus gallus domesticus) slaughtered in Giwa market, Giwa local government, area, Kaduna state, Nigeria*. *Journal of Natural Sciences Research*, vol. 4, pp. 120–125.
- Khalil, LF, Jones, A. & Bray, RA. 1994 *Keys to the cestode parasites of vertebrates*. CABI Publishing and the Natural History Museum.
- Lozano, J, Anaya, A, Salinero, AP, Lux Hoppe, EG, Gomes, L, Paz-Silva, A, Rebelo, MT & Madeira de Carvalho, E. 2019. *Gastrointestinal Parasites of Free-Range Chickens – A Worldwide Issue*. *Bulletin UASVM Veterinary Medicine*, vol. 76, pp. 110-117.
- Ludwig, JA & Reynolds, JF. 1988. *Statistical Ecology: a primer on methods and computing*. Wiley-Interscience Publications.
- Magurran, AE. 2004. *Measuring Biological Diversity*. Blackwell Publishing.
- Miao, Z H, Glatz, PC & Ru, YJ. 2005. *Free-range Poultry Production - A Review*. *Asian-Australasian Journal of Animal Sciences*, vol. 18, pp. 130-132.
- [Nnadi, PA & George, SO](#). 2010. *A cross-sectional survey on parasites of chickens in selected villages in the subhumid zones of South-eastern Nigeria*. *Journal of Parasitology Research*, vol. 2010, pp. 1-6.
- Poulin, R.2007. *Evolutionary ecology of parasites from individuals to communities*. Princeton University Press.
- Poulsen J, Permin, A, Hindsbo, O, Yelifari, L, Nansen, P & Bloch P. 2000. *Prevalence and distribution of gastro-intestinal helminths and haemoparasites in young scavenging chickens in upper eastern region of Ghana, West Africa*. *Preventive Veterinary Medicine*, vol.45, pp.237-245.
- Quadros, RM, Wiggers, SB, Paes, MPV &



- Marques, SMT. 2015. *Prevalência de endo e ectoparasitos de galinhas caipiras em pequenas propriedades da região serrana de Santa Catarina*. Publicações em Medicina Veterinária e Zootecnia, vol. 9, pp. 1-5.
- Rohde, K, Hayward, C & Heap, M. 1995. *Aspects of the ecology of metazoan ectoparasites of marine fishes*. International Journal of Parasitology, vol. 25, pp. 945-970.
- Ruff, MD. 1999. *Important parasites in poultry production systems*. Veterinary Parasitology, vol. 84, pp. 337-347.
- Santoro, M, Mattiucci, S, Nascetti, G, Kinsella, JM, Prisco, F, Troisi, S, D'Alessio, N, Veneziano, V & Aznar, FJ. 2012. *Helminth Communities of Owls (Strigiformes) Indicate Strong Biological and Ecological Differences from Birds of Prey (Accipitriformes and Falconiformes) in Southern Italy*. PLoS ONE, vol. 7, pp. e53375.
- Sarba, EJ, Bayu, MD, Gebremedhin, EZ, Motuma, K, Leta, S, Abdisa, K, Kebebew, G & Borena, BM. 2019. *Gastrointestinal helminths of backyard chickens in selected areas of West Shoa Zone Central, Ethiopia*. Veterinary Parasitology: Regional Studies and Reports, vol. 15, pp. 100265.
- Schou, TW, Permin, A, Juul-Madsen, HR, Sørensen, P, Labouriau, R, Nguyễn, TL, Fink, M & Pham SL. 2007. *Gastrointestinal helminths in indigenous and exotic chickens in Vietnam: association of the intensity of infection with the Major Histocompatibility Complex*. Parasitology, vol. 134, pp. 561-573.
- Silva, GS, Romera, DM, Conhalato, GS, Soares, VE & Meireles, MV. 2018. *Helminth infections in chickens (Gallus domesticus) raised in different production systems in Brazil*. Veterinary Parasitology: Regional Studies and Reports, vol. 12, pp. 55-60.
- Silva, GS, Romera, DM, Fonseca, LEC & Meireles, MV. 2016. *Helminthic parasites of chickens (Gallus domesticus) in different regions of São Paulo State, Brazil... Helminthic parasites of chickens (Gallus domesticus) in different regions of São Paulo State, Brazil*. Brazilian Journal of Poultry Science, vol. 18, pp. 163-168.
- Simões, RO, Luque, JL, Gentile, R, Rosa, MCS, Costa-Neto, S & Maldonado, A. 2016. *Biotic and abiotic effects on the intestinal helminth community of the brown rat Rattus norvegicus from Rio de Janeiro, Brazil*. Journal of Helminthology, vol. 90, pp. 21-27.
- Simões, RO, Maldonado JR., A., Olifiers, N, Garcia, JS, Bertolino, AVFA & Luque, JL. 2014. *A longitudinal study of Angiostrongylus cantonensis in an urban population of Rattus norvegicus in Brazil: the influences of seasonality and host features on the pattern of infection*. Parasites & Vectors, vol. 7, pp. 1-8.
- Siqueira, GB & Marques, SMT. 2016. *Parasitos intestinais em galinhas caipiras da região metropolitana de Porto Alegre, RS*. Publicações em Medicina Veterinária e Zootecnia, vol. 10, pp. 690-695.
- Schmidt, GD. 1986. *Handbook of Tapeworm Identification*. CRC. Press. Inc., Boca Raton.
- Slimane, BB. 2016. *Prevalence of the gastrointestinal parasites of domestic chicken Gallus domesticus Linnaeus, 1758 in Tunisia according to the agro-ecological zones*. [Journal of Parasitic Diseases](#), vol. 40, pp. 774-778.
- Taylor, MA, Coop, RL & Wall, RL. 2017. *Parasitologia Veterinária*. 4ª ed Guanabara Koogan.
- Vicente, JJ, Rodrigues, HO, Gomes, DC & Ponto, RM. 1995. *Nematóides do Brasil. Parte IV: Nematóides de aves*. Revista Brasileira de Zoologia, vol. 12, pp. 1-273.
- Wendt, EW, Monteiro, CM & Amato, SB. 2018. *Helminth fauna of Megaleporinus obtusidens (Characiformes: Anostomidae) from Lake Guaíba: analysis of the parasite community*. Parasitology Research, vol. 117, pp. 445-456.
- Wilkinson, L. 1990. *SYSTAT: The system for statistics*. Systat Inc, Illinois, p. 822.
- Yamaguti, F. 1961. *Sistema Helminthum. Vol II. The nematodes of vertebrates*. Interscience Publisher, New York, p. 1261.
- Yazwinski, TA, Chapman HD, Davis, RB, Letonja, T, Pote, L, Maes, L, Vercruyse, J & Jacobs, DE. 2003. *World Association for the Advancement of Veterinary Parasitology (WAAVP) guidelines for evaluating the effectiveness of anthelmintics in chickens*

*and turkeys*. Veterinary Parasitology, vol.116, pp.159-173.  
Zar, JH. 1999. *Bioestatistical Analysis*. Prentice-Hall, Inc., Upper Saddle River.

Received June 4, 2021.  
Accepted July 8, 2021.